Plasma protein turnover (albumin, transferrin, IgG, IgM) in Ménétrier's disease (giant hypertrophic gastritis): Evidence of non-selective protein loss

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SUMMARY Ten cases of Ménétrier's disease ('giant hypertrophic gastritis') have been studied with radioiodine-labelled albumin (all 10 cases), IgG (eight cases), transferrin (two cases), and IgM (six cases). Abnormal gastric loss of plasma protein was present in all cases as demonstrated by $^{59}$Fe-iron dextran (eight cases), $^{51}$Cr-albumin (one case), and $^{131}$I-polyvinylpyrrolidone (one case).

None of the patients had more distal gastrointestinal lesions. The synthetic rate of the proteins studied was normal or slightly elevated.

The fractional catabolic rate of the proteins was increased. The increase above the normal mean was similar for albumin, transferrin, and IgG. Since the molecular weight of IgG is more than twice that of albumin and transferrin, it is concluded that the protein loss in Ménétrier's disease is non-selective in the sense that it affects a similar fraction of the intravascular masses of all plasma proteins. IgM catabolism was strongly accelerated. Simultaneous studies with $^{59}$Fe-iron dextran, radioiodine-labelled albumin or IgG showed that IgM hypercatabolism could only partly be due to abnormal gastric protein loss, since IgM catabolism was significantly more raised than that of albumin and IgG. Faecal radioiodine excretion was normal in most patients after the injection of radioiodine-labelled proteins. It confirms a previous observation that increased gastrointestinal $^{59}$Fe clearance after injection of $^{59}$Fe-iron dextran associated with normal faecal radiiodine excretion after the injection of radioiodine-labelled proteins permits of a diagnosis of protein loss in the stomach.

In 1888 Ménétrier described a peculiar lesion of the stomach consisting of widespread hypertrophy of the mucosal folds (polyadénomes en nappe'). He was credited eponymously for his observation (Ménétrier's disease). Some 200 cases with a similar lesion have been published in this century. A different term has been used in several instances, eg, giant hypertrophic gastritis, giant mucosal rugae, protein-losing gastropathy, a fact which suggests that the pathoanatomical lesion may not represent a nosological unity. The course of the disease is most often benign. In rare cases carcinoma develops. Severe haemorrhage or oedema due to hypoproteinaemia occasionally requires partial or total gastrectomy.

Hypoproteinaemia is frequent, and occurs in about 70% of the cases (Jarnum, 1963). Serum albumin is depressed, but even the concentration of $\beta$- and $\gamma$-globulins may be lowered. Using $^{131}$I-albumin

Citrin, Sterling and Halsted (1957) showed that abnormal gastric plasma protein loss was responsible for the hypoproteinaemia. They were able to quantify the albumin loss directly by analyses of gastric secretions aspirated continuously over a prolonged period (24 hours). However, the method is laborious, unpleasant for the patient, and probably inferior to more recent methods, which apply isotope-labelled macromolecular compounds in which the isotope, as opposed to $^{131}$I in $^{131}$I-albumin, is not absorbed from the lumen of the gastrointestinal tract. $^{67}$Cu-labelled coeulioplasmin fulfils this condition (Waldmann, Morell, Wochner, Strobe, and Sternlieb, 1967) but the tracer is expensive. $^{51}$Cr-albumin (Waldmann, 1966) and $^{59}$Fe-iron dextran (Jarnum, Westergaard, Yssing, and Jensen, 1968) also seem to be valid test substances for the detection and quantitation of gastrointestinal protein loss. However, no tracer substance can accurately reflect the loss of some 100 plasma proteins of widely differing molecular size unless the

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protein loss is a ‘bulk loss’, which evenly comprises all plasma proteins.

In the present report 10 patients with Ménétrie’s disease were studied. 59Fe-iron dextran, 131I-albumin (MW (molecular weight) about 68000), 131I-transferrin (MW about 70000), 125I-IgG (MW about 160000), and 125I-IgM (MW about 920000) were used to assess whether a bulk loss took place.

Methods

SERUM PROTEIN DETERMINATION
The serum concentration of albumin, IgG, and IgM was determined by an immunochemical method (Mancini, Carbonara, and Heremans, 1965).

GASTROINTESTINAL PROTEIN LOSS
This was assessed by means of 59Fe-labelled iron dextran obtained from Pharmacia, Copenhagen, and manufactured at the Radiochemical Centre, Amersham, England (Jarnum et al, 1968).

TURNOVER STUDIES
Metabolically homogeneous 131I-labelled human albumin was obtained from Institutte for Atomenergi, Kjeller, Norway (code MISN).

Immunoglobulin-G was prepared from normal human serum by means of DEAE cellulose chromatography (Peterson and Sober, 1960).

ISOLATION AND LABELLING OF IgM
IgM was isolated from normal human serum by means of Sepharose 4B gel filtration (Jensen, 1969, 1970).

RADIOIODINE LABELLING OF IMMUNOGLOBULINS
IgG and IgM were labelled with 131I according to the ICI method of McFarlane (1958). On an average the labelled preparations contained one atom of iodine per molecule. More than 99.5% of the radioactivity was precipitable by trichloroacetic acid.

PROCEDURE
Thyroid uptake of radioiodide was prevented by daily oral administration of 50 mg potassium iodide from the day before and during the turnover study. Studies with 59Fe-, 131I-, and 125I-labelled preparations were performed simultaneously. Serum protein determinations were performed each day and used to correct the serum radioactivity for minor fluctuations in protein concentration. The patients were admitted to the ward during the study. None was confined to bed. Daily control of body weight and temperature and regular estimation of haemoglobin and serum proteins ensured that the patients were in a metabolically steady state condition.

Weighed amounts of the labelled preparations were administered intravenously. About 0.04 µCi of 59Fe, 0.4 µCi of 131I, and 0.15 µCi of 125I per kg were given to the patients. Radioactivity in serum was recorded 10 minutes after the injection and subsequently at daily intervals. In addition 59Fe and 131I activity was recorded by a whole body counter (Jarnum et al, 1968), which permitted the investigations to conclude within eight to 12 days.

Stools were collected until they became red after the oral administration of 1 g of carmin 96 hours after the injection. Urine was collected in 24-hour specimens during the studies. Radioactivity in serum and urine was measured in 3 ml samples by means of a three-channel gammaspectrometer (Auto-Gamma-spectrometer, Packard). Stool activity was measured in 24-hour samples in a large volume counter (Armac, Packard).

CALCULATION OF THE METABOLIC DATA
The faecal excretion of 59Fe was expressed as a percentage of the injected dose, and the gastrointestinal clearance of 59Fe-labelled iron dextran was calculated as the percentage of intravascular 59Fe excreted in the stools per day (Jarnum et al, 1968).

Albumin and IgG turnover was calculated by mathematical analysis of the multiexponential curve formed when plasma radioactivity was plotted against time in a semilogarithmic system (Nosslin, 1966). Synthesis was deduced from the calculated catabolic rate, which is only permissible if a steady state is prevailing throughout the study. This was actually the case as judged from the patients’ body weights, haemoglobin, and serum protein values.

The turnover of labelled IgM was calculated from plots of serum activity and retained activity (as determined by whole body counting) against time in a semilogarithmic system. According to Berson, Yalow, Schreiber, and Post (1953), the intravascular fraction of the labelled protein is equal to the ratio between the intercepts at zero time of the parallel final slopes of the serum and whole body curves, and the fractional catabolic rate is the slope constant divided by this ratio. Plasma volumes were determined by simple isotope dilution. The fractional catabolic rate is the percentage of the intravascular mass of protein degraded per day. The intravascular fraction denotes the fraction of the total body mass localized intravascularly. The intravascular mass equals plasma volume x serum concentration. The synthesis rate denotes the absolute amount of protein which is synthesized (or degraded) per day. It equals intravascular mass x fractional catabolic rate.
Faecal $^{131}$I excretion during the first four or five days of the study was calculated as a percentage of the injected dose. All patients were studied simultaneously, or separated by an interval of one or two weeks, with $^{59}$Fe-iron dextran (for assessment of gastrointestinal protein loss) and radiiodine ($^{125}$I or $^{131}$I) labelled albumin and IgG and IgM.

**Gastric Protein Clearance**
In five patients gastric aspiration was continuous over one to five hours from one to nine days after the injection of labelled proteins. The amount of protein-bound radioactivity per minute was related to the simultaneous plasma concentration of labelled protein, and the gastric 'clearance' of labelled protein was calculated. Gastric protein clearance was related to total metabolic clearance which was obtained from the product of plasma volume (ml) and the fractional catabolic rate (percentage per day).

In one patient (case 8) clearance studies of gastric IgG and albumin clearance were done following hexamethonium-induced 'vagotomy'. In a subsequent study of the same patient during an augmented histamine test and after an additional injection of $^{125}$I albumin, the proteolysis of the gastric juice was reduced by instillation of phosphate buffer (sodium phosphate, ionic strength 0.2, $pH$ 7.2) every five to 10 minutes.

**Case Material**
The diagnosis, Ménétrier's disease, was considered to comprise 'giant rugae' as well as giant rugae plus gastric polyps. It was based on radiography, the macroscopic appearance of the gastric mucosa on gastroscopy or laparotomy, and the microscopic picture of a biopsy or a surgical specimen. Ten patients were studied, three females and seven males, with an age range from 29 to 62 years. Laboratory and clinical findings are summarized in Tables I and II. Three patients (cases 8, 9, 10) suffered from additional diseases which are known to influence plasma protein turnover (sarcoidosis in prednisone treatment (case 8), nephrotic syndrome (case 9), and pulmonary carcinoma (case 10)).

The disease turned out to be transitory in the three females. Gastrointestinal plasma protein loss was estimated by means of $^{59}$Fe-iron dextran in eight patients (Jarnum et al, 1968), $^{51}$Cr-albumin in one patient (Waldmann, 1966), and $^{131}$I-polyvinylpyrrolidone in one patient (Gordon, 1959). Albumin turnover was studied in 10 patients. Case 4 was studied twice, before and after gastric resection. $^{125}$I-IgG turnover was studied in eight patients, and $^{131}$I-IgM turnover in six patients (cases 1, 3, 5, 6, 9, 10).

For comparison transferrin turnover determined in two patients (cases 7 and 9) has been included. The results have been published elsewhere (Jensen, Bro-Jørgensen, Jarnum, Olesen, and Yssing, 1968).

Control studies were carried out in normal persons or patients with disorders without known influence on serum protein turnover.

**Results**

**Abnormal Plasma Protein Loss**
Increased faecal $^{59}$Fe excretion and clearance was present in all patients studied (Table III). In patient no. 4 gastrointestinal plasma protein loss was demonstrated as increased faecal $^{131}$I excretion after

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Sex and Age (years)</th>
<th>Height (cm)</th>
<th>Weight (kg)</th>
<th>Associated Disease</th>
<th>Haemoglobin Blood in Stools (g/l)</th>
<th>Peak Acid Output (m-equiv/l H$^+$ per hour) (augmented histamine or pentagastrine)</th>
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<tbody>
<tr>
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<td>31</td>
<td>147</td>
<td>46</td>
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<tr>
<td>2</td>
<td>M</td>
<td>41</td>
<td>181</td>
<td>88</td>
<td>Multiple skeletal malformations</td>
<td>123 0 22 0</td>
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<tr>
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<td>M</td>
<td>57</td>
<td>171</td>
<td>59</td>
<td>Prostatic hypertrophy</td>
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</tr>
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<td>4</td>
<td>M</td>
<td>62</td>
<td>169</td>
<td>63</td>
<td>Subclavian artery stenosis</td>
<td>147 0</td>
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<td></td>
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<td></td>
<td></td>
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</tr>
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<td>Transitory Ménétrier's Disease</td>
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<tr>
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<tr>
<td>6</td>
<td>F</td>
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<td>151</td>
<td>45</td>
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<td>F</td>
<td>52</td>
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<td>57</td>
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</tr>
<tr>
<td>8</td>
<td>M</td>
<td>32</td>
<td>174</td>
<td>66</td>
<td>Sarcoïdosis in prednisone</td>
<td>135 + 3 2</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>treatment</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>M</td>
<td>56</td>
<td>175</td>
<td>79</td>
<td>Nephrotic syndrome, coronary</td>
<td>142 0 38 0</td>
</tr>
<tr>
<td>10</td>
<td>M</td>
<td>61</td>
<td>170</td>
<td>83</td>
<td>Anaplastic pulmonary carcinoma</td>
<td>115 + + + 0</td>
</tr>
</tbody>
</table>

Table I  Ten cases of Ménétrier's disease
Plasma protein turnover in Ménétrier’s disease

Table III

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Barium Meal Examination</th>
<th>Gastroscopic Findings</th>
<th>Laparotomy Findings</th>
<th>Gastric Biopsy or Surgical Specimen</th>
<th>Treatment</th>
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</thead>
<tbody>
<tr>
<td>1</td>
<td>Large mucosal folds</td>
<td>Irregular mucosa with polyps</td>
<td>Large mucosal folds + polyps</td>
<td>Polyps</td>
<td>Palliation</td>
</tr>
<tr>
<td>2</td>
<td>Large mucosal folds + proximal polyps</td>
<td>Large mucosal folds + polyps</td>
<td>Chronic atrophic gastritis in corpus</td>
<td>Anticoagulant treatment</td>
<td>Subtotal gastrectomy</td>
</tr>
<tr>
<td>3</td>
<td>Large mucosal folds in fundus and corpus</td>
<td>Large mucosal folds + polyps</td>
<td>Hyperplasia and polyps</td>
<td>Anticoagulant treatment</td>
<td>Subtotal gastrectomy</td>
</tr>
</tbody>
</table>

Transitory Ménétrier’s Disease

5 Large mucosal folds in corpus | Large mucosal folds in corpus | Chronic atrophic gastritis in corpus | 0 |

6 Large mucosal folds in corpus and fundus | Normal mucosa | Normal mucosa | 0 |

7 Large mucosal folds in corpus and fundus | ‘Thick mucosa’ | Polyps and irregular glands | Diuretics |

Cases with Associated Diseases Affecting Plasma Protein Turnover

8 Giant mucosal folds | Large mucosal folds + polyps | Polyps with eosinophilia | Total gastrectomy |

9 Large mucosal folds + polyps | Large mucosal folds in corpus | Chronic atrophic gastritis in corpus | 0 |

10 Large mucosal folds in corpus + polyps | Large mucosal folds in corpus | Chronic atrophic gastritis in corpus | Pneumonectomy |

Table II  Ten cases of Ménétrier’s disease

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Serum Albumin (g/l) (34-51)</th>
<th>Fractional Catabolic Rate (%) per day (7-1-10.3)</th>
<th>Distribution Ratio Intra-vascular Mass as % of Total Mass (37-52)</th>
<th>Synthesis Rate (g/175 cm/6) (7.3-14.4)</th>
<th>Faecal Radioiodine Excretion (% of injected dose in 4-5 days) (&lt;0-40%)</th>
<th>Faecal <strong>Fe</strong> Excretion after Intravenous Injection of <strong>Fe</strong>-iron dextran (% of injected dose in 4-5 days) (&lt;0-8%)</th>
<th>Gastrointestinal <strong>Fe</strong>-clearance (% of intravascular pool per day) (&lt;0-8%)</th>
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</thead>
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<tr>
<td>1</td>
<td>29.3</td>
<td>16.2</td>
<td>50.2</td>
<td>14.8</td>
<td>Unknown</td>
<td>4.3</td>
<td>7.5</td>
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<tr>
<td>2</td>
<td>41.3</td>
<td>13.3</td>
<td>61.2</td>
<td>19.5</td>
<td>0.13</td>
<td>3.9</td>
<td>4.4</td>
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<tr>
<td>3</td>
<td>37.2</td>
<td>14.3</td>
<td>42.0</td>
<td>11.0</td>
<td>0.51</td>
<td>6.8</td>
<td>10.9</td>
</tr>
<tr>
<td>4</td>
<td>26.1</td>
<td>13.4</td>
<td>44.0</td>
<td>10.1</td>
<td>0.47</td>
<td>(4.8)*</td>
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<td>5</td>
<td>37.8</td>
<td>8.0</td>
<td>33.6</td>
<td>10.0</td>
<td>0.14</td>
<td>(1.0)*</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>23.3</td>
<td>17.6</td>
<td>47.3</td>
<td>11.3</td>
<td>0.29</td>
<td>11.0</td>
<td>15.9</td>
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<tr>
<td>7</td>
<td>29.8</td>
<td>13.9</td>
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<td>8</td>
<td>23.8</td>
<td>24.5</td>
<td>56.2</td>
<td>16.4</td>
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<td>8.7</td>
<td>13.4</td>
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<tr>
<td>9</td>
<td>27.8</td>
<td>23.4</td>
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<td>21.5</td>
<td>0.25</td>
<td>(7.5)*</td>
<td>(11-1)*</td>
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<tr>
<td>10</td>
<td>26.1</td>
<td>23.9 (18.2)</td>
<td>51.7</td>
<td>20.9</td>
<td>0.19</td>
<td>&gt;3.4</td>
<td>&gt;5.3</td>
</tr>
<tr>
<td>11</td>
<td>27.4</td>
<td>16.0</td>
<td>56.4</td>
<td>16.7</td>
<td>0.53</td>
<td>7.2</td>
<td>8.3</td>
</tr>
</tbody>
</table>

Table III  Albumin turnover and gastric protein loss (**Fe**-iron dextran) in Ménétrier’s disease

1 After subtotal gastrectomy.
2 Faecal **131**I excretion (percentage of injected dose) following intravenous injection of **131**I-polyvinylpyrrolidone (**131**I-PVP). Normal value: <1%.
3 Faecal **51**Cr-excretion and clearance following intravenous injection of **51**Cr-albumin. Normal value for both: <1%.
4 Corrected for proteinuria. Normal range is given in parentheses.

the injection of **131**I-polyvinylpyrrolidone and in case 8 as increased **51**Cr excretion after **51**Cr-albumin. Thus an abnormal protein loss occurred in all 10 patients.

**ALBUMIN**

Serum albumin levels were lowered in all patients except patient no. 2 (Table III). He was also the patient who presented one of the lowest gastrointestinal losses as judged from faecal **55**Fe clearance (Table III). One patient (case 4) attained a normal serum albumin level following gastric resection.

Hypoalbuminaemia was due to accelerated fractional catabolism, which was present in all patients. There was a trend towards a hyperbolic relation between serum albumin and the fractional catabolic rate (Fig. 1). Gastrointestinal **55**Fe clearance seemed to be positively related to the fractional catabolic rate of albumin (Fig. 2). However, the correlation was not statistically significant (0.10 > p > 0.05). Albumin synthesis was elevated in six patients (from 14.8 to 21.5 g per 175 cm per day) and normal in the remaining four. Because of varying degrees of edema and malnutrition, the synthetic rate is related to body height (recalculated to 175 cm of body height) and not to body weight.
**Fig. 1** The inverse relation between serum concentration of albumin and IgG and their fractional catabolic rates.

- ○ = Cases 8-9-10 with associated diseases which might influence protein turnover (see text).
- ● = Normal range.

**TRANFERRIN**

In both cases studied the serum concentration was normal and the fractional catabolic rate increased (Table IV).

**IgG**

Serum concentration was slightly decreased in six, and normal in two of the eight cases studied (Table V). The fractional catabolic rate was increased in all subjects, and at the same time it was negatively and significantly ($r = -0.89, p < 0.01$) related to serum IgG concentration (Fig. 1). There was no statistically significant correlation between gastrointestinal $^{59}$Fe clearance and the IgG degradation rate (Fig. 2).

IgG synthesis was slightly elevated in five patients and high normal in the remaining three.

**IGM turnover**

Serum IgM was depressed in two patients (cases 1

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Serum Transferrin (g/l) (1.80-2.52)</th>
<th>Fractional Catabolic Rate (% per day) (14-9-19.5)</th>
<th>Distribution Ratio (5%) (45-54)</th>
<th>Synthesis Rate (g/175 cm/d) (0.80-1.79)</th>
<th>Faecal Radiiodine Excretion % (&lt;0.4)</th>
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<tr>
<td>7</td>
<td>1.86</td>
<td>27.9</td>
<td>63.0</td>
<td>1.32</td>
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<tr>
<td>9</td>
<td>2.26</td>
<td>3.94 (3.8)</td>
<td>53.7</td>
<td>2.39</td>
<td>0.22</td>
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</table>

**Table IV** Transferrin turnover in Ménétrier's disease

1Corrected for proteinuria.
**Plasma protein turnover in Ménétrier’s disease**

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Serum IgG (g/l) (6-9-14-2)</th>
<th>Fractional Catabolic Rate (% per day) (4-0-9-6)</th>
<th>Distribution Ratio (% (44-72))</th>
<th>Synthesis Rate (g/l75 cm/d) (0-93-3-28)</th>
<th>Faecal Radioiodine Excretion (%) (&lt;about 0-8)</th>
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<tbody>
<tr>
<td>1</td>
<td>5-7</td>
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<td>78-8</td>
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<td>73-5</td>
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**Table V  IgG turnover in Ménétrier’s disease**

*Corrected for proteinuria.

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Serum IgM (g/l) (0-23-1-33)</th>
<th>Fractional Catabolic Rate (% per day) (8-14)</th>
<th>Distribution Ratio Intravascular Mass as % of Total Mass (51-97)</th>
<th>Synthesis Rate (mg/l75 cm/day) (54-563)</th>
<th>Faecal Radioiodine Excretion (%) (&lt;about 0-8%)</th>
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<td>543</td>
<td>0-46</td>
</tr>
</tbody>
</table>

**Table VI  IgM turnover in Ménétrier’s disease**

*Corrected for proteinuria. Normal values are given in parentheses.

and 9, Table VI) and low normal in the remaining cases. The fractional catabolic rate was markedly elevated, from 26·7 to 31·8% of the intravascular mass per day or almost threefold the normal mean of 11% per day. The distribution of the protein was normal. On average, 68% of the total IgM mass was located intravascularly. IgM synthesis was normal.

**Protein Interrelations**

**Catabolic rates**

Patients studied with labelled transferrin or IgG were all studied simultaneously with labelled albumin. From Fig. 3 it appears that the increase in catabolism above the normal mean (percentage of intravascular mass per day) was very similar for all three proteins. The positive correlation between the catabolic rates for IgG and albumin was highly significant (r = 0·73, p < 0·01).

Comparing the fractional catabolic rates of albumin, IgG, and IgM it was obvious that IgM catabolism was relatively more increased than that of the other two proteins (Fig. 4). The average value of IgM catabolism was 29·1% of the intravascular mass per day (264% of the normal mean), and the corresponding values for albumin and IgG were 16·9 and 17·8% per day (194 and 262% of the normal mean), respectively.

**Synthetic rates**

Synthetic rates of albumin and IgG seemed to be positively correlated. However, the relationship was not significant. The distribution ratios of albumin, transferrin, and IgG were either high normal or raised in the sense that the fraction of the total mass present in the vascular bed was higher than normal. Some covariation in the distribution ratios of albumin and IgG was observed, but it did not reach a significant level. No correlation existed between the catabolic rate and the distribution ratio of albumin or IgG.

**Gastric Protein Clearance**

In five patients gastric protein clearance was assessed. However, in two of them (cases 1 and 7) no protein-bound activity was detected in gastric aspirations. In one case (no. 9) the clearance of protein-bound radioactivity was equal to 15 and 19% of the total metabolic clearance of albumin and IgG, respectively (Table VII).

In patient no. 10, who was the only one with achlorhydria, the gastric clearance of IgG amounted to 47% of the total metabolic clearance.

Case 8 had a gastric albumin clearance during the augmented histamine test equal to 71% of the total metabolic clearance (Fig. 5), whereas gastric albumin and IgG clearances after hexamethonium
Fig. 3 Albumin catabolism compared with the catabolic rate of transferrin (upper part) and IgG (lower part). The line \( x = y \pm a \) has been drawn in such a way that it traverses the normal mean of the catabolic rates of all proteins and predicts the increase in catabolism if a 'bulk loss' occurs, i.e., if albumin catabolism is increased 10% above normal mean, transferrin of IgG catabolism is also increased 10% above normal mean. \( \bigcirc \) = Normal range.

Fig. 4 Fractional catabolic rates of albumin, IgG, and IgM in six cases of Ménétrier's disease. The studies were performed simultaneously or separated by an interval of one to two weeks. Each patient has his own connecting line.

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Total Metabolic Clearance (( \mu l ) per minute)</th>
<th>Gastric Clearance of Protein-bound Radioiodine ( \mu l ) per Minute</th>
<th>Percentage of Total Metabolic Clearance</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Albumin</td>
<td>IgG</td>
</tr>
<tr>
<td>1</td>
<td>293</td>
<td>299</td>
<td>0</td>
</tr>
<tr>
<td>7</td>
<td>401</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>8</td>
<td>Augmented histamine</td>
<td>534</td>
<td>464</td>
</tr>
<tr>
<td>9</td>
<td>Hexamethonium</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>10</td>
<td>Normal values</td>
<td>433</td>
<td>459</td>
</tr>
<tr>
<td>10</td>
<td>Normal mean</td>
<td>469</td>
<td>218</td>
</tr>
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<td></td>
<td>150-359</td>
<td>95-292</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>255</td>
<td>193</td>
<td>0</td>
</tr>
</tbody>
</table>

Table VII Gastric protein clearance in Ménétrier's disease
Plasma protein turnover in Ménétrier’s disease

The common feature in the 10 cases of Ménétrier’s disease studied was the presence of hypertrophic (‘giant’) rugae of the stomach. None of them presented evidence of a more distal gastrointestinal lesion, and none suffered from malabsorption.

The diagnosis was based on a barium meal and gastroscopy. In two cases (nos. 4 and 8) it was confirmed at laparotomy. In three cases the disease turned out to be transitory, since the symptoms vanished, serum protein rose to normal values, and the radiographic picture of the stomach returned to normal. This rather unusual course of the disease will be described in detail elsewhere.

It is not the intention of the present report to contribute to the definition of ‘hypertrophic gastritis’ which is still in dispute. Our purpose was to elucidate the mechanism of abnormal protein loss which probably occurs in about 70% of patients with giant rugae in the stomach (Jarnum, 1963).

In the present report an abnormal protein loss was detected in all patients by means of $^{59}$Fe-labelled iron dextran, $^{51}$Cr-albumin, or $^{131}$I-PVP. The fractional catabolic rate of albumin was also increased in all, and the increase seemed to be correlated with the gastrointestinal $^{59}$Fe clearance although the correlation did not reach a statistically significant level.

Two patients were studied simultaneously with labelled albumin and transferrin and eight patients with labelled albumin and IgG. The increase in fractional catabolic rates of the three proteins above the normal mean was very similar in all cases (Fig. 3). This is strong evidence that the protein loss is a ‘bulk loss’ which affects a similar fraction of the intravascular masses of these three proteins. The observation is in contrast to the findings in the nephrotic syndrome where a selective protein loss occurs through the glomerular membrane resulting in a higher clearance of relatively low molecular proteins like albumin (molecular weight about 68 000) than of IgG (molecular weight about 160 000) (Jarnum, Jensen, and Bro-Jørgensen, 1966).

Six patients were studied with labelled albumin, IgG, and IgM. The relative increase in IgM catabolism was higher (on an average 264% of the normal mean) than that of albumin and IgG (194 and 262% of the normal mean, respectively). In absolute terms the average increase of IgM catabolism as a percentage of the intravascular mass per day above the normal mean was 18·1 as opposed to 8·2 and 11·0 for albumin and IgG, respectively. Thus
the increase in IgM catabolism was about twice as high as that of the other two proteins despite the fact that the molecular weight of IgM (about 920 000) is six and 13 times higher than that of IgG and albumin, respectively. It follows that the high catabolic rate of IgM in Ménétrier's disease cannot solely be due to abnormal unselective gastric protein loss. The mechanism of this hypercatabolism of IgM is unknown. In another condition with abnormal gastrointestinal protein loss, Crohn's disease, there is also an increased degradation rate of IgM. However, only in patients with an associated abscess did the increase in the catabolic rate of IgM exceed that of the simultaneously studied IgG catabolism (Jensen, Goltermann, Jarnum, Week, and Westergaard, 1970). In the present series there was no evidence of an associated abscess. All had a normal temperature and most of them had a normal erythrocyte sedimentation rate and a normal white cell count.

One may speculate whether the stomach is an organ which is normally a major degradation site of IgM. Probably no other organ in the body is exposed to so many antigens (species different proteins) every day. IgM might have an important function in the 'neutralization' of these antigens. In Ménétrier's disease there is a manifold increase in the mucosal surface area, which might mean that the expenditure of IgM is particularly high in this disease. The mucosa is heavily infiltrated with immunocytes (lymphocytes and plasma cells) (Kenney, Dockerty, and Waugh, 1954). Probably, it implicates local antibody synthesis on a large scale. It does not explain the rapid elimination of systemically administered homologous IgM molecules. However, both phenomena may have the same mechanism: a continuous antigen challenge across a much enlarged epithelial membrane with an abnormality high permeability to macromolecules. This would account for the local concentration of immunocytes and for a high 'consumption' of IgM antibodies during local neutralization of antigens. A high local IgM synthesis would result in a high local concentration of IgM as compared to other plasma proteins in the mucosal and submucosal 'pools' of plasma proteins. If so, a large amount of IgM lost in the stomach would be inevitable in the presence of a 'bulk loss' mechanism for protein loss. This mechanism could be further elucidated by means of systemically administered labelled IgM and the determination of specific activity of IgM in gastric juice from patients with Ménétrier's disease. Such studies were not undertaken in the present investigation.

Gastric protein clearance was determined to assess whether the directly observed protein loss in the stomach could account for the increased catabolism of albumin and IgG. Owing to the high proteolytic activity of gastric juice it was only possible to apply the observations made in two patients, one (case 8) in whom intragastric instillation of neutralizing buffer prevented proteolysis, and one (case 10) with achlorhydria. In both these cases the gastric protein clearance (of albumin and IgG, respectively) could fully account for the increased catabolism of albumin and IgG.

It is noteworthy that the administration of hexamethonium bromide markedly reduced the protein loss (case 8, Table IV). A similar observation after atropine injection was made by Overholt and Jeffries (1970).

Faecal radioiodine excretion after the injection of radioiodine-labelled proteins (albumin, transferrin, IgG) was normal in most cases. It confirms our previous report that increased gastrointestinal 59Fe clearance (after injection of 59Fe-iron dextran) in the presence of normal faecal radioiodine excretion (after the injection of radioiodine labelled proteins) permits a topographical diagnosis of abnormal protein loss in the stomach (Westergaard, Jarnum, Jensen, Seltoft, and Yssing 1968).

References


Plasma protein turnover in Ménétrié’s disease


The January 1972 Issue

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Stig Jarnum and Kurt Birger Jensen

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