Suppression of rejection of *Nipponstrongylus brasiliensis* in iron and protein deficient rats: effect of syngeneic lymphocyte transfer


*From the University of New South Wales, Gastrointestinal Unit, Prince of Wales Hospital, and Department of Veterinary Pathology, University of Sydney, New South Wales, Australia*

SUMMARY Rejection of *Nipponstrongylus brasiliensis* is impaired in iron and protein deficient rats and this suggests that iron and protein deficiency directly or indirectly suppresses the immune response. The site of the immunological defect in deficient rats was investigated using the technique of cellular transfer of resistance. The functional activity of immune mesenteric lymph node cells obtained from iron and protein deficient donors was not depressed as measured by their capacity to cause parasite rejection in nutritionally sufficient recipients. In contrast, immune lymph node cells obtained from either sufficient or deficient donors did not result in parasite rejection in iron and protein deficient recipients. These results indicate that there is no permanent defect of lymphocyte function in iron and protein deficient rats and suggest that either some other component of the rejection mechanism is defective, or that lymphocyte function is blocked in an iron and protein deficient environment.

Helminthiasis remains a major problem in many parts of the world, particularly in developing countries where an association malnutrition is common. A variety of factors are involved in this association, including the ability of gut parasites to contribute to the diarrhoea/malnutrition cycle (Stanfield, 1966). Dietary or parasite induced malnutrition may potentiate helminth infections by reducing the capacity of the host to mount a protective immune response.

Evidence that malnutrition potentiates parasite survival through impairment of the host immune response has been provided by a series of experiments using the *Nipponstrongylus brasiliensis*—rat model. In this system, rats infected with *N. brasiliensis* normally develop an actively acquired immunity and thereafter are resistant to subsequent infection. However, in iron and protein deficient rats, worm expulsion following a primary infection is delayed (Bolin *et al.*, 1977) and acquired resistance to reinfection is substantially impaired (Duncombe *et al.*, 1978).

Recent studies have shown that the immune response which initiates expulsion of *N. brasiliensis* from the gut is at least a four-step process. In the first step, antibodies initiate biochemical and morphological damage to worms (Jones and Ogilvie, 1971) which then become susceptible to the second and third steps for which sensitised lymphocytes (Kelly and Dineen, 1972) and a bone marrow derived component (Dineen and Kelly, 1973) are required. In the fourth step, sensitised lymphocytes and the bone marrow derived component interact with various mediators of inflammation such as prosta-glandins (Kelly *et al.*, 1974) finally to effect worm expulsion. Although the precise sequence of events has not been determined, it is known that antibody triggered damage must occur before sensitised lymphocytes can initiate the immunologically non-specific phase of worm expulsion involving pharmacological mediators (Kelly *et al.*, 1974).

As worm expulsion is an immunological process, it is reasonable to assume that delayed expulsion of *N. brasiliensis* is due to a defect of one or more components of the immune response. In a previous study it was reported that antibody mediated damage occurred normally in iron and protein deficient rats even though rejection was impaired (Bolin *et al.*, 1977). This would suggest that failure of worm expulsion is due to a defect in either sensitised lymphocytes or immunologically non-specific components of the immune response. Thus the aim of
this study was to determine whether functional
lymphoid cells are produced by iron and protein
rats infected with *N. brasiliensis*. This was assessed
using the cell transfer technique described by Kelly
and Dineen (1972).

Methods

Animals

Donor and recipient animals were highly inbred DA
rats of both sexes (Dineen et al., 1973).

Parasite

Third stage larvae of *N. brasiliensis* were obtained
from charcoal cultures of faeces (Leigh, 1956). Methods of infection and estimation of total worm
counts have been described (Kelly and Dineen, 1972).

Preparation of Iron and Protein
Deficient Animals

Synthetic diets were prepared which were either
sufficient or deficient in iron and protein. A sufficient
diet contained 30% protein, 50% carbohydrate, and
9.6% fat with vitamin and mineral supplements. An
isocaloric iron and protein deficient diet contained
no iron supplement and only 10% protein (Bolin et
al., 1971). Blood was collected for haemoglobin
before inoculation with larvae and albumin estima-
tions were carried out at the time of necropsy for
total worm counts.

Immunisation of Cell Donors

Donors for cell transfer were infected twice with 1000
third stage larvae of *N. brasiliensis* at 21 and seven
days before removal of mesenteric lymph nodes.

Preparation and Transfer of
Syngeneic Lymph Node Cells

The method described by Kelly and Dineen (1972)
was followed. The peritoneal cavity was opened
using a sterile technique. Mesenteric lymph nodes
were removed from donors, dissected free of fat, and
placed in chilled Hanks solution (pH 7.3). The
lymph nodes were then diced with fine scissors, and
the diced fragments vigorously drawn up and down
with a wide-mouthed pipette. The fragments were
filtered through sterile gauze and the resulting cell
suspension centrifuged for six minutes at 200 g. The
supernatant was removed, the cells resuspended in
fresh Hank’s solution and centrifuged again. After
removal of the supernatant, washed cells were
resuspended in fresh chilled Hank’s solution and the
number of cells/ml estimated using a haemocyto-
meter. Viability was assessed by the capacity of cells
to exclude 2% Trypan blue. The volume of the cell
suspension was then adjusted so that 1 ml contained
125 × 10^6 viable cells and this was transferred to
recipients within two hours of killing donors by
intravenous injection into the lateral tail veins.

Statistics

Body weight, haemoglobin, and serum albumin
levels were analysed by Student’s *t* test. Analysis
of variance was carried out on all worm count data
after transformation to log_{10}(x + 1), and individual
comparisons made with Student’s *t* test.

Experimental design and results

Three week old DA rats were weaned onto synthetic
diets either sufficient or deficient in iron and protein.
Five weeks later these animals were infected twice
with *N. brasiliensis* and acted as either sufficient or
deficient cell donors (see Methods section).

Two identical experiments were carried out and the
results are pooled. There were five animals in
each group in each experiment. Iron and protein
sufficient (groups 1, 2, and 5) and deficient (groups
3, 4, and 6) rats were infected with 1000 third stage
larvae of *N. brasiliensis*. Groups 5 and 6 received no
further treatment and acted as infection controls.
At this time, rats in all groups were weighed and had
blood collected for haemoglobin levels. On the day of
infection, rats in groups 1 and 3 were each injected
intravenously with 125 × 10^6 cells from infected
donors sufficient in iron and protein, while those in
groups 2 and 4 were each injected intravenously with
125 × 10^6 cells from infected donors deficient in iron
and protein. Ten days later, rats in all groups were
killed for total worm counts and blood collected for
serum albumin levels. These results are presented in
Tables 1 and 2.

Table 1 Body weight, serum albumin, and haemoglobin
levels in iron and protein sufficient and deficient rats

<table>
<thead>
<tr>
<th>Dietary status</th>
<th>Iron and protein sufficient</th>
<th>Iron and protein deficient</th>
<th><em>p</em> value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight (g)</td>
<td>184 ± 9.1</td>
<td>104 ± 3.4</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Albumin (g/l)</td>
<td>32.0 ± 0.3</td>
<td>26.0 ± 0.2</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Haemoglobin (g/dl)</td>
<td>13.9 ± 0.2</td>
<td>8.0 ± 0.1</td>
<td>&lt;0.01</td>
</tr>
</tbody>
</table>

Body weight, Serum Albumin, and
Haemoglobin Levels

There was a statistically significant difference in body
weight, serum albumin and haemoglobin levels
between sufficient and deficient rats, indicating
iron and protein deficiency in animals on a deficient
diet.
Suppression of rejection of Nippostrongylus brasiiliensis in iron and protein deficient rats

LYMPHOCYTE YIELDS FROM DONOR RATS
The average total cell yields for mesenteric lymph nodes from deficient individual donors was $139 \times 10^6$, which was substantially lower than the average of $200 \times 10^6$ lymphocytes recovered from donors on a sufficient diet. This difference was significant at the 5% level. Although the lower recovery may suggest a diminished immune response to *N. brasiiliensis* in deficient donors, it may also be related to differences in body weights between the two groups.

TOTAL WORM COUNTS
The total worm counts performed 10 days after worm infection and cell transfer demonstrated no significant difference between non-recipient infection controls on a sufficient (group 5, mean count 275), or deficient diet (group 6, 320; $P > 0.05$). By comparison with these non-recipient controls, there was a highly significant reduction in total worm counts in rats on a sufficient diet given cells either from a sufficient (group 1, 5; $P < 0.01$) or deficient (group 2, 17; $P < 0.01$) donors. This showed that transferred cells are viable and effective as immune lymphocytes in initiating worm expulsion in normal recipients, regardless of the dietary status of the cell donors.

There was no significant difference between non-recipient infection controls on a deficient diet (group 6, 320) and iron and protein deficient rats receiving cells from sufficient (group 3, 269) or deficient (group 4, 298) immunised donors. This showed that the defect in *N. brasiiliensis* expulsion in deficient rats cannot be overcome by transfer of immune lymphocytes known to be fully functional (group 1, Table 2).

Although lymphocytes obtained from immunised nutritionally deficient donors were effective in initiating worm rejection when transferred to recipients on a sufficient diet (group 2), lymphocytes from nutritionally sufficient donors were not effective in deficient recipients (group 3). These results indicated that functional immune lymphocytes were produced by iron and protein deficient rats.

Discussion
Depression of the immune response has been observed in both iron (Nalder et al., 1972; MacDougall et al., 1975) and protein (Jose et al., 1973; Gebhardt and Newberne, 1974) deficiency and an immune defect has been implicated in the suppressed rejection of *N. brasiiliensis* from iron and protein deficient rats (Bolin et al., 1977). The purpose of this present study was to determine whether cell mediated immunity was suppressed in deficient animals using the technique of syngeneic lymphocyte transfer.

The results show that transfer of immune lymphocytes from donors of either dietary group significantly reduced worm counts in rats on a sufficient diet. Thus lymphocytes from deficient donors were able to function effectively in iron and protein sufficient rats, as determined by their capacity to expel parasites. In contrast, cells from either iron and protein sufficient or deficient animals did not reduce worm counts in deficient recipient rats. These results indicate that delayed worm expulsion in iron and protein deficiency is not due to a permanent defect in the immunologically specific component of the response involving lymphocyte function.

Thus delayed parasite expulsion in iron and protein deficiency could be due to some factor which reversibly blocks lymphocyte function, or to one or more defects in the rejection mechanism not involving sensitised lymphocytes. Reversible blockade of lymphocyte function has been reported previously in lactating rats, where inhibition of induced lymphoid cells into effector cells has been observed (Kelly and Dineen, 1973). Multiple defects in the immune response have also been reported in iron and protein deficient rats, involving impaired humoral immunity (Nalder et al., 1972; Good et al., 1976), cell mediated immunity (Jose et al., 1973; Gebhardt and Newberne, 1974), and bone marrow function (Aschkenasy, 1957; Sood et al., 1965). In addition the myeloid-amine response is altered with reduction in eosinophil.

Table 2  Effect of lymphocyte transfer on expulsion of Nippostrongylus brasiiliensis from iron and protein sufficient and deficient rats

<table>
<thead>
<tr>
<th>Dietary status of donor rats</th>
<th>Group no.</th>
<th>Dietary status of recipient rats</th>
<th>Total worm counts on day 10 after infection (mean ± SE)</th>
<th>$P$ value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sufficient</td>
<td>1</td>
<td>Sufficient</td>
<td>5 ± 1.1</td>
<td>GP 1 v 5 &amp; 6</td>
</tr>
<tr>
<td>Deficient</td>
<td>2</td>
<td>Sufficient</td>
<td>17 ± 7.9</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Sufficient</td>
<td>3</td>
<td>Deficient</td>
<td>269 ± 43.7</td>
<td>GP 2 v 5 &amp; 6</td>
</tr>
<tr>
<td>Sufficient</td>
<td>4</td>
<td>Deficient</td>
<td>298 ± 52.6</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Dietary status of infection controls</td>
<td>5</td>
<td>Sufficient</td>
<td>275 ± 44.4</td>
<td></td>
</tr>
<tr>
<td></td>
<td>6</td>
<td>Deficient</td>
<td>320 ± 50.0</td>
<td></td>
</tr>
</tbody>
</table>

Data transformed $\log_{10} (x + 1)$ for analysis of variance after which Student's $t$ test was employed for individual comparisons.
count and increased small bowel mucosal histamine in protein deficiency (Wells, 1962).

The complex interaction between nutrition, infection, and immunity clearly has important clinical implications for malnourished communities. The nutritional status of an individual alters his resistance to infection and may well influence the effectiveness of vaccination procedures and drug treatment regimes. Administration of transfer factor in malnourished man has met with little success in combating infection (Walker et al., 1975; Jose et al., 1976), and it is relevant to know more about the site of the immune defect in malnutrition. Studies are currently under way to assess the effect of bone marrow and serum transfer on the expulsion of *N. brasiliensis* in iron and protein deficient rats.

We would like to thank Miss Glenda Walsh, Miss Margaret Fagan, Mr Christopher Porter, and Mr D. Griffin for their excellent technical assistance. This study was supported by a grant from the National Health and Medical Research Council of Australia.

References


Jones, V. E., and Ogilvie, B. M. (1971). Protective immunity to *Nippostrongylus brasiliensis*: the sequence of events which expels worms from the rat intestine. *Immunology*, 20, 549-561.


Suppression of rejection of Nippostrongylus brasiliensis in iron and protein deficient rats: effect of syngeneic lymphocyte transfer.

A G Cummins, V M Duncombe, T D Bolin, A E Davis and J D Kelly

Gut 1978 19: 823-826
doi: 10.1136/gut.19.9.823