Biochemical changes in the jejunal mucosa of dogs with naturally occurring exocrine pancreatic insufficiency

R. M. BATT¹, B. M. BUSH, AND T. J. PETERS

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SUMMARY The roles of extracellular and intracellular mechanisms in the degradation of brush border proteins have been investigated by studying the small intestinal mucosa of dogs with naturally occurring exocrine pancreatic insufficiency. Peroral jejunal biopsies were homogenised and the organelles separated by isopycnic centrifugation on continuous sucrose density gradients. The distributions of marker enzymes for the principal subcellular organelles were determined in the gradients and related to the specific activities in the homogenates. There were increased activities of the brush border carbohydases zinc-resistant α-glucosidase, maltase and sucrase in the pancreatic insufficient animals, but no change in lactase activity. The activity of γ-glutamyl transferase was also higher in the affected group; the activities of two other brush border enzymes, alkaline phosphatase and leucyl-β-naphthylamidase, however, were unaltered. These findings with an increase in the modal density of the brush border from 1·20 to 1·22 are consistent with an enhanced glycoprotein content of the microvillus membrane. There were also rises in the activities of lysosomal enzymes. N-Acetyl-β-glucosaminidase activity was increased in the soluble fractions and the percentage latent enzyme activity was reduced, findings indicative of an increased fragility of the lysosomal membrane. There were no marked alterations in the activities or density gradient distributions of marker enzymes for the other organelles, stressing the specificity of the changes in the brush borders and lysosomes. These findings are compatible with the degradation of certain exposed brush border proteins by pancreatic proteases and suggest that when this is defective, intracellular degradative mechanisms may be stimulated.

The enterocyte is a highly specialised cell involved in the terminal processing of nutrients and their transfer from the lumen of the small intestine. Although the life span of this cell is relatively short, the protein content of the enterocyte is not static, proteins turning over at heterogeneous rates by a continuous process of synthesis and degradation (Alpers and Kinzie, 1973). A balance between these processes maintains the steady state level of individual proteins. Free ribosomes are responsible for the synthesis of intracellular proteins, whereas those destined for export or insertion into membranes are synthesised on the rough endoplasmic reticulum (Dallner et al., 1966). Control of synthesis, by the regulation of genetic expression, may be achieved in a variety of ways at the transcriptional and post-transcriptional levels (Gelehrter, 1973) and may be influenced by exogenous stimuli such as corticosteroids (Baxter et al., 1972). Protein degradation and the controlling mechanisms, on the other hand, are not well understood, particularly in the enterocyte. The most specialised organelle, the brush border, is at the luminal surface and hence in a unique position potentially susceptible to both extracellular and intracellular degradative mechanisms. Pancreatic proteolytic enzymes (Alpers and Tedesco, 1975; Kwong et al., 1978) and lysosomal enzymes (Seetharam et al., 1976) are thought to play a part in the turnover of specific brush border proteins. This has been investigated in the present study by examining the biochemical changes in the jejunal mucosa at a subcellular level in dogs with naturally occurring exocrine pancreatic insufficiency. A pre-

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liminary report of this work has been published previously (Batt et al., 1979a).

Methods

Groups of Animals

Control animals were normal dogs with no functional or histological evidence of a jejunal abnormality. Exocrine pancreatic insufficiency was detected in animals presented for an investigation of diarrhoea and weight loss by the demonstration of severe steatorrhoea and negligible faecal proteolytic activity. The clinical details and results of the screening tests (Batt et al., 1979b) are shown in Table 1. The diagnosis was confirmed by the oral administration of N-benzoyl-L-tyrosyl-p-aminobenzoic acid, a substrate specific for chymotrypsin, and the subsequent estimation of plasma and urine p-aminobenzoic acid (PABA) (Batt et al., 1979b). The results (Table 1) show that at two hours normal animals achieve a plasma PABA concentration approximately 10 times greater than that of the animals with pancreatic insufficiency. The urinary excretion of PABA similarly distinguished between the two groups, most of the PABA being excreted in the control animals in a six hour period in contrast with the negligible amounts found in the urine of the affected animals.

Analytical Subcellular Fractionation

Peroral jejunal biopsies were obtained at the duodenal-jejunal flexure with a Watson paediatric capsule (Batt, 1979). Portions of the tissue were homogenised in sucrose (0·3 mol/l) containing EDTA (1 mmol/l) pH 7·4, and ethanol (22 mmol/l) (SVE medium) and a post-nuclear supernatant (PNS fraction) subjected to analytical subcellular fractionation in the Beaufay automatic zonal rotor as described previously (Batt and Peters, 1978a). After centrifugation, alkaline phosphatase was assayed fresh and the remainder of the gradient then stored at −20°C. Results are expressed in the form of frequency-density histograms, the pooling and averaging of distributions being performed by computer (Leighton et al., 1968).

Latent N-Acetyl-β-Glucosaminidase

Latent-N-acetyl-β-glucosaminidase activity was determined by assaying this enzyme in the PNS fraction in SVE medium containing buffered substrate with (total activity) and without (free activity) Triton X-100 (100 mg/100 ml) (Batt and Peters, 1976).

\[
\text{Latent activity (}% = \frac{\text{Total activity} - \text{Free activity}}{\text{Total activity}} \times 100
\]

Analytical Techniques

Marker enzymes for the principal subcellular organelles were assayed as described previously (Batt and Peters, 1978a). Acid phosphatase was assayed with 4-methylumbelliferyl phosphate (0·15 mmol/l) in sodium acetate buffer (0·1 mol/l), pH 4·0, containing Triton X-100 (100 mg/100 ml). Maltase, sucrase, and lactase were assayed (Peters et al., 1976) in portions of biopsies collected in deionised water and disrupted in a Dounce homogeniser (Kontes Glass Co., Vineland, New Jersey, USA) with 20 strokes of a tight-fitting pestle (type B). Protein was determined according to Schacterle and Pollack (1973) with bovine serum albumin (Armour Pharmaceutical Co., Chicago, USA) as a standard. The unpaired Student's t test was used to assess the significance of differences between the two groups.

Results

Enzyme Activities

The specific activities of brush border enzymes in biopsies from control and pancreatic insufficient animals are shown in Table 2. There are significant increases in the activities of zinc-resistant α-glucosidase, maltase, sucrase, and γ-glutamyl transferase in the affected animals, but no significant changes in

<table>
<thead>
<tr>
<th>Group and case no.</th>
<th>Age (yr)</th>
<th>Weight (kg)</th>
<th>Fat absorption (%)</th>
<th>Faecal fat (g/day)</th>
<th>Faecal proteolytic activity (azo-casein units/g)</th>
<th>2 h plasma PABA (μmol/l)</th>
<th>6 h urinary excretion of PABA (% of oral dose)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pancreatic insufficiency</td>
<td>1</td>
<td>3</td>
<td>30</td>
<td>59</td>
<td>16.0</td>
<td>2.5</td>
<td>1.2</td>
</tr>
<tr>
<td>2</td>
<td>2½</td>
<td>27</td>
<td>40</td>
<td>14.4</td>
<td>1.4</td>
<td>2.3</td>
<td>9.6</td>
</tr>
<tr>
<td>3</td>
<td>5</td>
<td>37</td>
<td>72</td>
<td>13.5</td>
<td>3.9</td>
<td>2.3</td>
<td>12.7</td>
</tr>
<tr>
<td>4</td>
<td>6</td>
<td>35</td>
<td>79</td>
<td>9.8</td>
<td>1.5</td>
<td>2.8</td>
<td>10.1</td>
</tr>
<tr>
<td>5</td>
<td>2½</td>
<td>35</td>
<td>55</td>
<td>21.2</td>
<td>1.6</td>
<td>---</td>
<td>---</td>
</tr>
<tr>
<td>Control</td>
<td>3.8 ± 0.6(11)</td>
<td>2.2 ± 0.4(11)</td>
<td>67.2 ± 12.5(11)</td>
<td>31.8 ± 3.4(4)</td>
<td>76.3 ± 3.6(4)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Control data, with number of observations between parentheses, are presented as mean ± SEM.
Exocrine pancreatic insufficiency in the dog

Table 2  Activities (mUnits/mg protein) of brush border enzymes in biopsies from control and pancreatic insufficient animals

<table>
<thead>
<tr>
<th>Group</th>
<th>Zinc-resistant α-glucosidase</th>
<th>Malate</th>
<th>Sucrase</th>
<th>Lactase</th>
<th>Alkaline phosphatase</th>
<th>Leucyl-β-naphthylamidase</th>
<th>γ-Glutamyl transferase</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>6-1 ± 0-3</td>
<td>336 ± 24</td>
<td>62-6 ± 5</td>
<td>31-4 ± 3-0</td>
<td>138 ± 11</td>
<td>101 ± 9-6</td>
<td>10-9 ± 0-6</td>
</tr>
<tr>
<td>Pancreatic insufficiency</td>
<td>8-1 ± 0-8</td>
<td>535 ± 76</td>
<td>136 ± 28</td>
<td>45-7 ± 9-7</td>
<td>101 ± 14</td>
<td>111 ± 13</td>
<td>15-5 ± 1-7</td>
</tr>
<tr>
<td>Statistical significance (P)</td>
<td>0-01</td>
<td>0-01</td>
<td>0-003</td>
<td>0-2</td>
<td>0-1</td>
<td>0-6</td>
<td>0-004</td>
</tr>
</tbody>
</table>

Data are expressed as mean ± SEM. The figures between parentheses indicate the number of observations.

The activities of lactase, alkaline phosphatase, or leucyl-β-naphthylamidase. The specific activities of marker enzymes for the other subcellular organelles are shown in Table 3. The activity of 5'-nucleotidase, a basal-lateral membrane enzyme, is unaltered. There are, however, large increases in the activities of the acid hydrolases N-acetyl-β-glucosaminidase and acid phosphatase in the pancreatic insufficient animals, although α-mannosidase activity is unchanged. Malate dehydrogenase, with both soluble and mitochondrial components, tris-resistant α-glucosidase, an endoplasmic reticular enzyme, and catalase, a peroxisomal enzyme, have similar activities in the two groups. Latent N-acetyl-β-glucosaminidase activity, a measure of lysosomal integrity, is significantly reduced in the animals with pancreatic insufficiency compared with the control.

SUBCELLULAR FRACTIONATION STUDIES

The relative specific activities and density gradient distributions of the principal marker enzymes in jejunal biopsies from control and pancreatic insufficient animals are compared in the Figure. The distributions of these enzymes in the normal dog have been reported previously (Batt and Peters, 1978a).

The increase in zinc-resistant α-glucosidase activity in the affected animals is almost entirely associated with the brush border. This organelle shows a distinct shift in modal density from 1-20 in the control to 1-22 in the pancreatic insufficient group, a change also reflected in the distributions of the other three brush border enzymes alkaline phosphatase, leucyl-β-naphthylamidase and γ-glutamyl transferase. Although the activities of alkaline phosphatase and leucyl-β-naphthylamidase are not altered, the enhanced activity of γ-glutamyl transferase in the affected animals is clearly associated with the brush border.

The distributions of 5'-nucleotidase (basal-lateral membrane) are not markedly different comparing the two groups but there is a small shift in modal density from a value of 1-11 in the control to 1-12 in the pancreatic insufficient animals. The subcellular distributions of N-acetyl-β-glucosaminidase, in contrast, are strikingly different between the two groups. Although the particulate activities associated with lysosomes are almost identical, there is a dramatic increase in soluble enzyme activity in the pancreatic insufficient group. The activities and distributions of the other marker enzymes malate dehydrogenase (mitochondria-cytoplasm) and tris-resistant α-glucosidase (endoplasmic reticulum) and of catalase (peroxisomes, not shown) revealed negligible differences between the two groups.

Discussion

The role of pancreatic enzymes in the turnover of brush border proteins has been investigated by studying the small intestinal mucosa of dogs with naturally occurring exocrine pancreatic insufficiency.

Table 3  Activities (mUnits/mg protein) of marker enzymes and percent latent N-acetyl-β-glucosaminidase activity in biopsies from control and pancreatic insufficient animals

<table>
<thead>
<tr>
<th>Group</th>
<th>5'-Nucleotidase</th>
<th>N-acetyl-β-glucosaminidase</th>
<th>Acid phosphatase</th>
<th>α-Mannosidase</th>
<th>Malate dehydrogenase</th>
<th>Tris-resistant α-glucosidase</th>
<th>Catalase</th>
<th>Latent N-acetyl-β-glucosaminidase</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>2-97 ± 0-28</td>
<td>3-57 ± 0-22</td>
<td>3-75 ± 0-35</td>
<td>1-17 ± 0-10</td>
<td>2464 ± 280</td>
<td>0-70 ± 0-04</td>
<td>8-6 ± 0-76</td>
<td>70-1 ± 1-2</td>
</tr>
<tr>
<td>Pancreatic insufficiency</td>
<td>2-41 ± 0-64</td>
<td>5-26 ± 0-91</td>
<td>6-53 ± 0-95</td>
<td>1-15 ± 0-16</td>
<td>2561 ± 520</td>
<td>0-75 ± 0-1</td>
<td>8-0 ± 1-7</td>
<td>55-3 ± 5-8</td>
</tr>
<tr>
<td>Statistical significance (P)</td>
<td>0-4</td>
<td>0-02</td>
<td>0-005</td>
<td>0-9</td>
<td>0-9</td>
<td>0-6</td>
<td>0-7</td>
<td>0-003</td>
</tr>
</tbody>
</table>

Data are expressed as mean ± SEM. The figures between parentheses indicate the number of observations.
Figure  Isopycnic centrifugation of PNS fraction from jejunal biopsies of control (solid line) and pancreatic insufficient animals (stippled). Graphs, of averaged data from at least three experiments, show the relative frequency-density distributions of eight marker enzymes. For each enzyme the areas of the distributions comparing the two groups are proportional to enzyme specific activity. Frequency is defined as that portion of total recovered activity present in an individual fraction divided by the density span covered by that fraction. Relative frequency (mean±SEM) was derived by multiplying the frequency data for the pancreatic insufficient animals by the relative specific activity (mUnits/mg protein) of the pancreatic compared with the control animals. The percentage recoveries for the control and pancreatic insufficient animals, respectively, with the relative specific activities between parentheses, are: zinc-resistant α-glucosidase 81, 83 (1.3); alkaline phosphatase 58, 77 (0.7); leucyl-β-naphthylamidase 79, 76 (1.1); γ-glutamyl transferase 84, 87 (1.4); 5'-nucleotidase 79, 129 (0.8); N-acetyl-β-glucosaminidase 68, 68(1.5); malate dehydrogenase 87, 60 (1.0); tris-resistant α-glucosidase 107, 95 (1.1).
Biochemical characterisation of the enterocyte at a subcellular level has permitted an examination of the consequences of pancreatic insufficiency not only on the microvillus membrane but also on the other organelles of this cell.

Pancreatic degenerative atrophy in the dog is a well-recognised naturally occurring condition with a predisposition for the Alsatian or German Shepherd Dog (Hill et al., 1971). Histologically, there is a close similarity to the exocrine pancreatic insufficiency reported in CBA/J mice (Eppig and Leiter, 1977) where there is a degeneration of acinar tissue without a severe inflammatory involvement and a retention of the islets of Langerhans, major ducts, vessels, and nerves. The present study emphasises how little functional acinar tissue remains in the dogs with exocrine insufficiency, the urine recovery of PABA after oral peptide being almost as low as in dogs with the pancreatic duct ligated (Imondì et al., 1972). In common with the mouse model, the lack of distinct endocrine abnormalities limits any effects on the small intestine which may be due to diabetes (Olsen and Korso, 1975).

The dogs with pancreatic insufficiency were found to have marked and quite specific changes in the brush borders of the jejunal enterocytes. The modal density of this organelle was increased and accompanied by enhanced activities of the carboxydrases zinc-resistant α-glucosidase, maltase, and sucrase. The activity of γ-glutamyl transferase was also higher in the affected animals than in the control; the activities of both alkaline phosphatase and leucyl-β-naphthylamidase, however, were essentially unaltered. These findings are consistent with previous studies both in man (Arvanitakis and Olsen, 1974) and the experimental animal (Alpers and Tedesco, 1975; Kwong et al., 1978) implicating pancreatic enzymes in the normal turnover of the microvillus membrane. Low intraluminal concentrations of pancreatic enzymes could result in a decreased degradation—and thus increased brush border content—of enzymes normally susceptible to this attack. This susceptibility, in turn, must depend on the accessibility of the appropriate peptide bonds to these proteolytic enzymes and their importance in determining either the attachment of the brush border enzyme to the microvillus membrane or its active three-dimensional conformation.

Biochemical studies (Eichholz, 1968; Critchley et al., 1975; Louvard et al., 1975) indicate that, with the exception of trehalase, the carbohydrases are located at the external surface of the microvillus membrane, a position most vulnerable to the activity of pancreatic enzymes, and that they may be released from the membrane readily by pancreatic proteases (Alpers and Tedesco, 1975). Alkaline phosphatase (Batt and Peters, 1978b) and leucyl-β-naphthylamidase (Louvard et al., 1976), in contrast, appear to be intrinsic membrane proteins which are more embedded in the microvillus membrane and consequently much less susceptible to release either by the action of papain or detergent (Eichholz, 1968; Critchley et al., 1975; Louvard et al., 1975). Indeed, pancreatic enzymes have been found to release negligible amounts of alkaline phosphatase activity from the brush border after incubation in vitro. Thus, in the present study, the enhanced activities of zinc-resistant α-glucosidase, maltase, and sucrase but unaltered activities of alkaline phosphatase and leucyl-β-naphthylamidase in the pancreatic insufficient animals may be explained by the differences in the localisation and orientation of these enzymes in the brush border membrane. The spatial arrangement of γ-glutamyl transferase in the microvillus membrane is less well understood; this study, however, suggests a susceptibility to pancreatic enzymes and therefore a significant exposed component in the brush border of the dog.

There was little alteration in either the enzyme content or density of the basal-lateral membranes in the pancreatic insufficient animals, thus emphasising the specificity of the changes at the luminal surface of the plasma membrane of the enterocyte. There were, however, distinct effects on the acid hydrolases. The activities of both N-acetyl-β-glucosaminidase and acid phosphatase were raised, although α-mannosidase activity was unchanged. For N-acetyl-β-glucosaminidase this increase was almost entirely associated with the soluble fractions of the density gradient, a finding confirmed by demonstrating a reduction in the percentage latent enzyme activity. This suggests either that a soluble non-lysosomal component of this enzyme activity is specifically raised, or, a more likely interpretation, that the lysosomes have an increased fragility.

The possible functions of lysosomes, particularly the roles of heterophagy and autophagy, in relation to cellular economy have been discussed by de Duve and Wattiaux (1966). There is considerable evidence for the fusion of endocytic vesicles with lysosomes (Miller and Palade, 1964; de Duve and Wattiaux, 1966) and it has been suggested that this may be associated with the recycling of constituents of the plasma membrane (Silverstein et al., 1977; Werb and Cohn, 1972). The specific part lysosomes play in the turnover of the brush border membrane is not known; there is, however, good evidence for endocytosis by the enterocyte and the subsequent fusion of vesicle and lysosomal membranes (Walker and Isselbacher, 1974). The ability of lysosomal enzymes to degrade microvillus membrane proteins appears to be limited (Seetharam et al., 1975), so that an increase
in the protein content of the brush border, associated with decreased extracellular proteolysis, could result in the excessive accumulation of material within the lysosomes. In common with other conditions where this occurs, an enhancement in the fragility of the lysosomal membrane may result that is associated with an increase in the activities of specific lysosomal enzymes (Seymour and Peters, 1978).

An excessive uptake of macromolecules from the lumen of the small intestine could have the same consequences. A reduction in the concentration of pancreatic enzymes will increase the number of macromolecules, including antigen and antigen-antibody complexes (Walker et al., 1975), at the surface of the enterocyte. This provides the opportunity for an enhanced absorption of these molecules (Walker and Isselbacher, 1974) and could result in an accumulation in the lysosomes and hence damage to the surrounding membrane.

There was little change in either the activities or density gradient distributions of marker enzymes for the mitochondria or peroxisomes. This was also true for particulate tris-resistant $\alpha$-glucosidase activity, suggesting that there was no major change in the endoplasmic reticulum and being consistent with the hypothesis that a decreased rate of degradation is responsible for the increased activities of specific brush border enzymes. This is in contrast with the increased activities in the prednisolone-treated animal, where a proliferation of the rough endoplasmic reticulum suggests an enhanced rate of synthesis of brush border enzymes and carrier proteins (Batt and Peters, 1976; Batt et al., 1978).

A study of exocrine pancreatic insufficiency in the dog has revealed highly specific changes in the brush border membrane and evidence of altered intracellular degradative mechanisms in response to decreased intraluminal concentrations of proteolytic enzymes. Further investigation of this condition should provide additional information on the biochemical and physiological interactions between the pancreas and the small intestine.

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