Does gastric aminopyrine clearance reflect gastric mucosal blood flow or parietal cell function?

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SUMMARY Gastric aminopyrine clearance was measured in human volunteers and dogs with untreated basal secretion, in human volunteers and dogs treated with secretory inhibitors, in dogs treated with histamine, and in patients with pernicious anaemia. When aminopyrine was given as a bolus to man or dog, aminopyrine clearance and the ratio aminopyrine concentration in gastric juice/aminopyrine concentration in plasma showed an initial peak two to three times over steady state levels. When aminopyrine was given with histamine, the peaks were even higher. No peaks occurred when an aminopyrine bolus was given to patients with pernicious anaemia or to healthy volunteers treated with secretory inhibitors. The height of the peaks paralleled the acid secretory rate. The peaks may best be explained by aminopyrine accumulation in the parietal cells and washing out of aminopyrine by volume flow. The steady state levels might reflect both parietal cell function and gastric mucosal blood flow.

It is widely accepted that gastric aminopyrine clearance reflects gastric mucosal blood flow. It is assumed that the gastric mucosa functions like a dialysis membrane between the neutral plasma and the acidic gastric juice. Thus, the only rate limiting factor for aminopyrine clearance would be gastric mucosal blood flow. We have, however, in isolated dispersed gastric parietal cells observed considerable trapping of aminopyrine in the intracellular canaliculus. Thus, the gastric mucosa would not be a simple dialysis membrane but would also store—and eventually release—aminopyrine. The storage capacity within the parietal cell would depend on intracellular acidity and thus parietal cell function. In addition, transient aminopyrine trapping could also occur in an unstirred layer covering the mucosa, including, among other structures, the glandular lumen. In order to test the assumption that aminopyrine clearance solely reflects mucosal blood flow, a series of experiments was done in man and dogs under basal conditions, with gastric secretion stimulated with histamine and restrained with anti-secretory agents. As the observations cannot be explained by blood flow alone and as aminopyrine clearance parallels acid secretion rate, aminopyrine clearance appears to reflect in part parietal cell function.

Methods
STUDIES IN MAN
After an overnight fast, a double lumen gastric tube was positioned under fluoroscopic control with its tip in the antrum. Gastric juice was collected by intermittent suction using an Egnell pump model EMP 171 687. Fasting gastric contents were removed during 15 minutes. Then, the stomach was perfused at a rate of 1.5 ml min using a Harvard pump model 975. The perfusion fluid contained 100 mM HCl and 2% polyethylene glycol 4000 (Siegfried, Switzerland). In order to insure patency of the aspiration port manual aspiration was performed every five minutes and a small bolus of air was given at that time. The gastric aspirates were pooled in 15 minute fractions.

(Dimethyl-14C)-aminopyrine was obtained from New England Nuclear. Its specific activity was 100 mCi/mM. At time zero, after aspiration of the fasting gastric juice, an intravenous 14C-aminopyrine bolus (8μCi) was given. This was followed by an intravenous infusion for three hours at a rate of 2 μCi h. For this purpose, aminopyrine was dissolved in normal saline (2 μCi 100 ml of saline, infusion rate 150 ml h). A winged needle with a heparin lock was positioned in a cubital vein. Blood samples were obtained as shown in Fig. 1. The blood was collected in tubes containing EDTA.
STUDIES IN DOGS
In nine dogs, Komarov type oesophagostomies were constructed under general anaesthesia. The Komarov oesophagostomies allowed easy insertion of gastric tubes but did not interfere with food and fluid intake. The experiments were started four weeks after completion of surgery. The experimental procedure resembled that in man (Fig. 1).

ASSAYS
Aminopyrine was extracted from plasma and gastric juice by dichloromethane as described by Tague and Jacobson. Samples were counted for $^{14}$C-radioactivity in a liquid scintillation counter. Appropriate corrections were made for the calculation of dpm/ml. The polyethylene glycol concentration of the gastric aspirates was measured by photometry after protein precipitation by cold acetone as described by Buxton. The acid concentration in the gastric juice was determined by titration with NaOH in 0.01 molar concentration. For the calculation of the amount of acid secreted the appropriate amount of infused acid was substracted. The aminopyrine clearance in ml min is given by $RV$ where $R$ = aminopyrine concentration of the gastric juice/ aminopyrine concentration of the plasma and $V$ is the net secretion volume per minute.

STATISTICS
The values of aminopyrine clearance and $R$, respectively, were compared by Student's t test. Paired tests were used when appropriate. Changes in the concentration of $^{14}$C in plasma and gastric juice, respectively, were compared with zero (= no change) by paired t tests. The relation between mean acid output and maximum of aminopyrine clearance was examined by linear regression analysis.

SUBJECTS STUDIED
The study protocol was approved by the local ethical committee.

Healthy human volunteers
a. Basal secretion Eight volunteers were studied.
b. Treatment with secretory inhibitors In six subjects, 200 mg cimetidine and 25 mg pirenzipine were given eight, four, and two hours before the beginning of the experiment. In addition, an intravenous infusion of cimetidine was given during the experiment at a dose of 66.7 mg/h.

Patients with pernicious anaemia
In four otherwise healthy subjects with pernicious anaemia (age 68, 72, 79, and 91 years) informed consent was obtained to perform the experiments as outlined above. No cimetidine or pirenzipine was administered.

Dogs
a. Basal secretion Each dog was studied once (= nine experiments) without additional treatment.
b. Treatment with histamine In four experiments dogs received histamine dihydrochloride (60 µg/kg/h intravenously) 60 minutes before gastric juice aspiration was started and the infusion lasted for the duration of the experiment.
c. Treatment with secretory inhibitors Three dogs were treated with cimetidine given orally (400 mg eight hours and 200 mg two hours before each experiment) and then by intravenous infusion (66.7 mg/h during the study).

Results

\[ y = 12.7 + 34.6x, \quad r = 0.82, \quad n = 16, \quad p < 0.01 \]

Fig. 2. \(^{14}\text{C}\) radioactivity in plasma (●—●) and gastric juice (□——□). Top: volunteers under basal conditions. Bottom: dogs under basal conditions.

Aminopyrine clearance values in man and dog
Marked initial peak are noted in human controls and in histamine treated dogs (Fig. 3). In histamine treated dogs the peak is higher and occurs earlier than the peak in untreated dogs (p < 0.05) where peaks are also noted. In volunteers treated with cimetidine and pirenzipine, in patients with pernicious anaemia, and in dogs treated with cimetidine no peaks occur. Similar observations were obtained for the R values (Table).

Relation between aminopyrine clearance and acid secretion
When the mean acid output is compared with the maximal value of the aminopyrine clearance, a linear correlation is obtained in man (Fig. 4) and dog \((y = 12.7 + 34.6x, \quad r = 0.82, \quad n = 16, \quad p < 0.01)\). Similar observations were made for mean acid output and the steady state values of the aminopyrine clearance.
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Table  Values for ratio aminopyrine concentration in gastric juice/aminopyrine concentration in plasma in man and dog (means ± SEM)

<table>
<thead>
<tr>
<th>Group</th>
<th>Maximum</th>
<th>Steady state</th>
</tr>
</thead>
<tbody>
<tr>
<td>Man</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Basal (control)</td>
<td>33 ± 5§</td>
<td>15 ± 5</td>
</tr>
<tr>
<td>Secretory inhibition</td>
<td>14 ± 1</td>
<td>4 ± 2*</td>
</tr>
<tr>
<td>Pernicious anaemia</td>
<td>14 ± 1</td>
<td>4 ± 1*</td>
</tr>
<tr>
<td>Dog</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Basal (control)</td>
<td>15 ± 4§</td>
<td>5 ± 1</td>
</tr>
<tr>
<td>Histamine stimulation</td>
<td>23 ± 9§</td>
<td>6 ± 1</td>
</tr>
<tr>
<td>Secretory inhibition</td>
<td>9 ± 1</td>
<td>6 ± 1</td>
</tr>
</tbody>
</table>

*P < 0.05, † P < 0.01 when compared with basal, § significantly higher than steady state level.

Discussion

Aminopyrine, like neutral red and other substances cleared by an acid partition mechanism, is widely used for determination of gastric mucosal

Fig. 3  Aminopyrine clearance values. Top: man. ○ volunteers under basal conditions; ● volunteers treated with secretory inhibitors; □ patients with pernicious anaemia. Bottom: dogs. ○ volunteers under basal conditions; ● treated with secretory inhibitors; □ treated with histamine. Means ± SEM.

* P < 0.05, † P < 0.01, ‡ P < 0.001. Encircled values are significantly higher than steady state levels (P < 0.05).

Fig. 4  Linear regression analysis comparing mean acid output with the maximal value (top) and with the steady state value (bottom), respectively, of aminopyrine clearance in man.

pyrine clearance (Fig. 4 and y = 11·1 · 5·0x, r = 0·84, n = 16, P < 0·01, respectively). Thus, aminopyrine clearance and acid output (in ml/mmol H+1) are related in both analyses but the ratio is higher during maximal clearance than during the steady state (803 · 152 vs. 304 ± 65, P < 0·01, in man, and 452 ± 142 vs. 76 ± 14, P < 0·01, in dogs). Thus, the ratio is not a measure for the blood flow required to produce a millimole of H+.
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blood flow. Its main advantage over other methods, such as 12K-clearance, 82Rb-clearance, and mucosal accumulation of radiolabelled microspheres, is the ease of in vivo measurements in animals and man. In the present study we investigated whether gastric mucosal blood flow is the only determinant of gastric aminopyrine clearance. As shown by Figs. 2 and 3 and the Table this is clearly not the case. Peaks of R and of aminopyrine clearance are observed during the first 60 to 120 minutes, both in healthy man and dogs. Interestingly, they were high in untreated man who is known to have a relatively high basal secretion. They were lower in untreated dogs where basal secretion is low but very high in dogs stimulated with histamine. They were absent in man and dogs treated with secretory inhibitors and also in achlorhydric patients with pernicious anaemia. Thus, the time course of gastric aminopyrine clearance cannot be explained by blood flow. A good explanation for the observed peaks is aminopyrine trapping in parietal cells and washing out by volume flow across the parietal cells. This explanation appears likely because a steady state of 14C-aminopyrine concentration occurs later in the gastric juice than in plasma while the stomach secretes (Fig. 2). The finding that the steady state levels depend on acid secretion (Fig. 4) is compatible with the classical model that aminopyrine clearance reflects gastric mucosal blood flow. As, however, peaks occur only in the secreting stomach, they cannot be explained by blood flow. Thus, the peaks might reflect parietal cell function and the steady state levels might reflect both parietal cell function and gastric mucosal blood flow.

Other authors have already given evidence for shortcomings of the aminopyrine method. Archibald et al. compared aminopyrine clearance with blood flow measurements obtained by the microsphere technique. They reported that, in the nonstimulated canine stomach, aminopyrine clearance reflects only a small fraction of the gastric mucosal blood flow, whereas during histamine stimulation the two methods give similar results. Moody compared aminopyrine clearance with 12K-clearance. During thiocyanate inhibition aminopyrine clearance paralleled acid secretion but did not reflect the blood flow changes measured by the 12K-clearance. Delaney et al. measured the effect of cimetidine on gastric mucosal blood flow by a microsphere technique. They did not observe a decrease of the gastric mucosal blood flow, in contrast with Konturek who observed a reduction of the aminopyrine clearance under similar experimental conditions. No reduction was obtained by a neutral red clearance technique. It has also been reported that the clearance values with aminopyrine do not reach steady state in the first hour of an experiment, but these findings, so far, have not received further consideration. Finally, Dugas et al. have given evidence that the aminopyrine clearance is not only limited by blood flow, but also by diffusion.

In conclusion, aminopyrine clearance appears to reflect both gastric mucosal blood flow and parietal cell function. As in many instances there is a parallel change of gastric mucosal blood flow and of parietal cell function, changes in aminopyrine clearance has the following limitations: (1) it is only a relative measure of gastric mucosal blood flow; (2) steady state conditions in plasma and gastric juice are required; and (3) the acid secretion rate should remain constant during the experiment.

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