Clinical detection of the hepatic lesion of pericentral sclerosis in chronic alcoholics

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SUMMARY It has been shown that a specific liver lesion—that is, pericentral sclerosis associated with pericellular fibrosis—is the precursor of alcoholic liver sclerosis. It is, however, difficult to diagnose this hepatic lesion in chronic alcoholics, using only clinical data without liver biopsy. To investigate the possibility of a clinical test reflecting the presence of this hepatic lesion, ethanol (0·75 g/kg body weight) was given orally to chronic alcoholics, and serum glycoprotein levels (prealbumin, αHSglycoprotein, haptoglobin, α₂macroglobulin) were measured before and six hours after. Chronic alcoholics were divided into three groups according to the histological findings in the liver at the time of study. Group I (alcoholic fatty liver or non-specific change) consisted of seven cases without pericentral sclerosis. Group II (alcoholic hepatic fibrosis or alcoholic hepatitis) consisted of five cases with pericentral sclerosis and pericellular fibrosis. Group III consisted of five cases with alcoholic liver cirrhosis. After the ethanol administration, serum glycoprotein levels decreased significantly in group I (p<0·05), whereas they increased in group II and group III. Their alternative ratios (see text) apparently differed (p<0·005) between group I and group II, and between group I and group III. These results indicate that the determination of serum glycoprotein levels before and after oral ethanol administration is a useful way of discriminating alcoholic patients with hepatic pericentral sclerosis and pericellular fibrosis from alcoholics without such lesions.

Alcoholic liver disease is increasing in proportion to the increase in alcohol consumption, and is becoming a serious health problem in Japan.1 It is therefore important to be able to diagnose and estimate the severity and prognosis of alcoholic liver disease.

Histological studies play an essential role in diagnosis and estimation of the severity of liver disease. It has been reported that a specific liver lesion—namely, pericentral sclerosis associated with pericellular fibrosis—is the precursor of alcoholic liver cirrhosis.2 It is, however, difficult to recognise this lesion in chronic alcoholics, if one uses only clinical data without liver biopsy. We therefore wanted to find some clinical marker indicating the presence of this hepatic lesion.

Hepatomegaly in alcoholics is reported to be attributable to protein deposition3 as well as fatty accumulation in the hepatocyte.4 It has also been suggested that this intracellular protein deposition is partly due to impaired glycoprotein secretion from the hepatocyte,5 6 and that it might play a role in the pathogenesis of alcoholic liver disease.

We examined serum glycoprotein level changes after the acute administration of ethanol, as the serum glycoprotein levels might reflect an altered protein metabolism in the hepatocyte.

Methods

SUBJECT Seventeen male patients were studied who had a history of heavy drinking (more than 100 g ethanol per day) for more than 10 years. Those who had a history of blood transfusion, a definite history of acute viral hepatitis, or a history of habitual taking of drugs were excluded from this study. None of them had HBs-Ag nor anti-HBs, as examined by radio-immunoassay, in their sera.

Under peritoneoscopic observation, liver biopsy, using a Vim Silverman needle, was performed. Liver specimens were interpreted independently by two observers without knowledge of clinical or laboratory data. Sections stained with haematoxylin and eosin,
and trichrome-stained, were classified into the following categories, based on the histological findings specific to alcoholic liver injury. They consisted of (1) non-specific change, (2) fatty liver, (3) alcoholic hepatitis, (4) hepatic fibrosis, and (5) liver cirrhosis. The histological features are shown in Table 1.

Table 1  Histological classification of alcoholic liver disease

<table>
<thead>
<tr>
<th>Classification</th>
<th>Histological features</th>
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<tbody>
<tr>
<td>Non-specific change of the liver</td>
<td>Normal liver histology or non-specific change of the liver</td>
</tr>
<tr>
<td>Fatty liver</td>
<td>Fatty metamorphosis is recognised in more than 25% of specimen with little other change</td>
</tr>
<tr>
<td>Alcoholic hepatitis</td>
<td>Existence of following three findings with few other findings: (1) hepatocyte necrosis with polymorphonuclear leucocyte infiltration with or without Mallory bodies, (2) fatty accumulation, (3) pericentral sclerosis with pericellular fibrosis</td>
</tr>
<tr>
<td>Hepatic fibrosis</td>
<td>Pericentral sclerosis associated with pericellular fibrosis without evidence of other change</td>
</tr>
<tr>
<td>Liver cirrhosis</td>
<td>Definite pseudonodular formation with micronodular appearance</td>
</tr>
</tbody>
</table>

The subjects were divided into three groups according to the histological classification described: group I consisted of seven cases with non-specific change or fatty liver. No pericentral sclerosis or pericellular fibrosis could be seen in this group. Group II consisted of five cases with alcoholic hepatic fibrosis and alcoholic hepatitis. In this group, pericentral sclerosis associated with pericellular fibrosis were the characteristic features. Group III consisted of five cases with alcoholic liver cirrhosis.

The mean age of group III (54.6 ± 2.9 years; mean ± SEM) was higher (p<0.05) than that of group II (42.5 ± 1.7 years). There was no significant difference, however, between the mean age of group I (44.6 ± 3.8 years) and group II. Also the difference between the mean age of group I and that of group III was not statistically significant. Duration of habitual drinking in group II (32.6 ± 2.4 years) was longer (p<0.05) than that of group I (21.4 ± 2.5 years) and group III (21.5 ± 4.4 years). In addition, total alcohol consumption of group III (1740 ± 160 kg) was greater (p<0.01) than that of group I (670 ± 140 kg) and group II (630 ± 20 kg). As to the mean abstinence period before ethanol administration, there were no significant differences among the three groups.

PROCEDURES
Before ethanol administration, a standard diet was given to each patient for five days. This represented a total heat value of 2200 kilocalories and contained 100 g protein. After overnight fasting for 15 hours, 20% ethanol (0.75 g/kg of body weight) was given orally for 60 minutes. Blood samples were collected from the antecubital vein before and six hours after ethanol administration. During the study, neither drinking nor eating was permitted (Table 2).

Table 2  Procedure of oral ethanol administration

1. Standard diet for five days (2200 Cal containing 100 g protein per day)
2. Overnight fasting (15 hours)
3. Blood sampling
4. Peroral administration of 20% ethanol for 60 minutes (0.75 g ethanol/kg body weight)
5. Blood sampling (after 6 hours)
6. Measurement of serum glycoprotein levels

TECHNIQUES
Serum concentrations of total protein and albumin were measured colorimetrically by an autoanalyzer (SMAC; Technicon Corp.) using biuret and bromcresol-green, respectively. Serum bilirubin concentrations were measured by the method of Jendrassik using SMAC. Serum aspartate aminotransferase (SGOT) activity was measured by the method of Brueton using SMAC. Serum gamma glutamyl transpeptidase (SGPT) activity was measured by a modification of the method of Orlowski et al.7 All these parameters were assayed within two days after the blood sampling. Serum glycoproteins (prealbumin, a1-HS-glycoprotein, haptoglobin, and a2-macroglobulin) were measured by single radial immunodiffusion using partigen plate (Hoechst Corp.).

ALTERNATIVE RATIOS
We calculated alternative ratios (AR), in order to take into account the degree of change in serum glycoprotein levels observed after ethanol administration as follows: AR = (Y - X)/X, where X is the serum glycoprotein level before ethanol administration and Y is the serum glycoprotein level six hours after administration.

STATISTICS
Results are expressed as the mean ± SEM. Significance between differences of means was assessed by Student's t test and Kruskal-Wallis's h test.

RESULTS
SERUM ENZYME ACTIVITIES
Before ethanol administration, the mean activity of sGOT in group III (67.0 ± 17.8 U/l) was higher than
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that of group I (29.0 ± 6.1) and group II (28.5 ± 6.1); there were, however, no significant differences between the three groups. The mean activity of sGGT was highest in group III (116.6 ± 34.5 U/l), compared with that in group I (62.1 ± 12.8) and group II (99.3 ± 37.6); there were, however, no significant differences between these three groups. Ethanol administration caused no change in sGOT and sGGT activity in any of the groups (Fig. 1).

![Graphs of enzyme activities](image)

Fig. 1  Serum enzyme activities before and six hours after ethanol administration. NS: not significant. N: number of subjects. Mean (± SEM) values for each group are shown by open and shaded bar.

SERUM PROTEIN, ALBUMIN, AND BILIRUBIN
Before ethanol administration, there were no significant differences among the three groups in serum concentrations of protein, albumin, and bilirubin. In addition, the ethanol caused no change in serum levels of these three parameters in any of the groups (Fig. 2).

![Graphs of protein, albumin, and bilirubin](image)

Fig. 2  Serum concentrations of protein, albumin, and bilirubin before and six hours after ethanol administration. NS: not significant. N: number of subjects. Mean (± SEM) values are shown. (SI conversion factor × 0.01.)

mg/100 ml), group II (0.241 ± 0.073g/1) (24.1 ± 7.3 mg/100 ml) and group III (0.185 ± 0.032g/1) (18.5 ± 3.2 mg/100 ml). After the ethanol was given, the serum prealbumin level decreased significantly (p<0.05) in group I (0.251 ± 0.031g/1) (25.1 ± 3.1 mg/100 ml). In contrast, it increased significantly (p<0.05) in group II (0.347 ± 0.085g/1) (34.7 ± 8.5 mg/100 ml) and group III (0.277 ± 0.056g/1) (27.7 ± 5.6 mg/100 ml) (Fig. 3a).

αHSglycoprotein level showed similar results to prealbumin; αHSglycoprotein level showed no differences among the three groups before ethanol administration. After the ethanol was given, it decreased significantly (p<0.05) only in group I (0.565 ± 0.073 → 0.436 ± 0.079g/1) (56.5 ± 7.3 → 43.6 ± 7.9 mg/100 ml), and increased significantly (p<0.05) in group III (0.459 ± 0.055 → 0.643 ± 0.121g/1) (45.9 ± 5.5 → 64.3 ± 12.1 mg/100 ml) (Fig. 3b).
Fig. 3 Serum glycoprotein levels before and six hours after ethanol administration. NS: not significant. Mean (± SEM) values are depicted. (SI conversion factor × 0.01.)

The haptoglobin level of group III (0.78 ± 0.095 g/l) (78.0 ± 9.5 mg/100 ml) was significantly lower than that of group I (1.794 ± 0.288 g/l) (179.4 ± 28.8 mg/100 ml) even before ethanol administration. After ethanol administration, it decreased significantly only in group I (1.794 → 1.314 ± 0.208 g/l) (179.4 → 131.4 ± 20.8 mg/100 ml). By contrast, it increased in group II (0.574 ± 0.076 → 0.789 ± 0.078 g/l) (57.4 ± 7.6 → 78.9 ± 7.8 mg/100 ml) and group III (0.780 → 1.432 ± 0.304 g/l) (78.0 → 143.2 ± 30.4 mg/100 ml), but the changes were not significant (Fig. 4a).

α2-Macroglobulin level showed similar results to haptoglobin; nevertheless, the serum levels of the three groups showed no significant difference from each other. Changes on ethanol administration were not significant, either (Fig. 4b).

**ALTERNATIVE RATIOS**

Mean alternative ratios are shown in Fig. 5. Each glycoprotein measured showed very similar alternative ratios; the mean alternative ratios of group I showed values lower than zero. In contrast, all of those in group II and group III were positive. We found apparent differences (p<0.005) between the mean alternative ratio of group I and those of group II and group III, respectively.

**Discussion**

It is difficult to detect the presence of pericentral sclerosis associated with pericellular fibrosis, using only clinical data, without liver biopsy. Indeed, the present study showed that there was no significant difference between group I and group II, before the administration of ethanol, in terms of serum enzyme activities and serum concentrations of protein, albumin, and bilirubin. In the present study, however, the findings that serum glycoprotein levels changed on ethanol administration and that the alternative ratios were different between group I and group II have particular import-
of liver disease. Several studies indicate a diminution of serum glycoprotein in acute hepatic failure, advanced liver cirrhosis, and other diseases. Other studies demonstrated that ethanol causes reduced glycoprotein synthesis and further impairment of secretion, and suggested that this change resulted in the intracellular accumulation of glycoprotein in the hepatocyte.

The present finding that the serum glycoprotein level of group I decreases after acute ethanol administration suggests impairment of glycoprotein secretion from the hepatocyte after acute ethanol intoxication. This interpretation is in agreement with recent studies suggesting impaired glycoprotein secretion during ethanol intoxication. On the other hand, we found that serum glycoprotein levels of group II and group III increased after ethanol administration. It may be suggested that this increase of serum glycoprotein level is partly due to the increased permeability of the hepatocyte membrane, after ethanol intoxication, in the liver with advanced damage, although it is not possible to define the exact mechanism of this change from the present study.

We suggest, on the strength of the present study, that the determination of serum glycoprotein levels is useful in the differentiation of alcoholic patients with hepatic fibrosis and alcoholic hepatitis from alcoholics with fatty liver, even when liver biopsy is not available.

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