Small intestinal absorption of amino acids and a dipeptide in pancreatic insufficiency

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SUMMARY In this study a perfusion technique has been used to investigate in vivo jejunal absorption and transmural potential difference evoked by the neutral amino acids phenylalanine (56 or 20 mmol/l) and glycine (20 mmol/l), the dibasic amino acid lysine (56 or 5 mmol/l), and a dipeptide glycyl-l-phenylalanine (20 mmol/l) in 11 children with pancreatic insufficiency due to cystic fibrosis and in three children with other causes of exocrine pancreatic insufficiency. Net absorption and potential difference evoked by phenylalanine in both cystic fibrosis and pancreatic insufficiency, and net absorption of glycine in cystic fibrosis were significantly reduced; but the absorption of lysine and glycyll-l-phenylalanine was normal. Absorption of the constituent amino acids from the dipeptide was normal or increased in cystic fibrosis. Thus, these studies show a defect in active absorption of neutral amino acids in cystic fibrosis with pancreatic insufficiency and exocrine pancreatic insufficiency. We speculate that pancreatic factors participate in neutral amino acid absorption.

It is well established that, despite adequate pancreatic replacement therapy, patients with pancreatic insufficiency due to cystic fibrosis excrete increased amounts of faecal nitrogen and fat. The increased faecal nitrogen is accounted for by both unhydrolysed proteins and amino acids of dietary origin. The faecal amino acids are derived in part from intraluminal bacterial degradation of intact proteins. Recent studies have shown defective in vitro uptake of neutral amino acids by jejunal biopsy material obtained from patients with cystic fibrosis, and impaired in vivo absorption of a synthetic neutral amino acid, suggesting that small intestinal malabsorption of certain amino acids may also contribute to the increased faecal nitrogen losses in cystic fibrosis.

We have utilised a steady-state perfusion technique to investigate the in vivo absorption of the neutral amino acids phenylalanine and glycine, the dibasic amino acid lysine, and a dipeptide glycyl-l-phenylalanine in children with cystic fibrosis, in those with other causes of exocrine pancreatic insufficiency, and in control subjects. The results have in part been reported in abstract form.

Methods

SUBJECTS

Eleven patients with pancreatic insufficiency due to cystic fibrosis with a mean age of 4·1 years (range 1·0-10·2 years), three patients with other forms of exocrine pancreatic insufficiency, two with Shwachman’s syndrome aged 2·2 and 4·3 years, one with acquired pancreatic insufficiency after chicken pox, and 16 control subjects with a mean age of 4·0 years (range 0·9-10·0 years) were investigated. The diagnosis of cystic fibrosis was based on raised sweat sodium concentrations (>70 mmol/l), typical clinical symptoms and signs, and pancreatic insufficiency. Shwachman’s syndrome was diagnosed on the basis of a combination of pancreatic insufficiency, impaired growth unrelated to dietary intake and/or malabsorption, intermittent neutropenia, defective neutrophil mobility, and skeletal anomalies. Pancreatic function was assessed by determining the activity of trypsin, amylase, and lipase in duodenal juice in response to a test meal as previously described. In all patients...
with pancreatic insufficiency there was a marked reduction in pancreatic enzyme activity compared with our established normal range. The control subjects comprised children with the irritable bowel syndrome who had a normal jejunal mucosa on light microscopy and no demonstrable evidence of malabsorption.

Growth was normal at the time of the study in both the cystic fibrosis and control subjects. Cystic fibrosis height velocity three months before and after study varied between the 50 and 90 centiles and height between 10–75 centiles; weight velocity between the 25 and 90 centiles, and weight between 10–50 centiles. Urinary hydroxy proline/creatinine ratio was 49–102 (normal range 30–254). At the time of study the control subjects’ height velocity varied between 25 and 75 centiles and height between 25–75 centiles; weight velocity between 10 and 75 centiles, weight 10–50 centiles, and urinary hydroxy proline ratio 45–89.

Neither the patients nor the control subjects received any drugs or pancreatic supplements for 72 hours before perfusion. The studies were approved by the Standing Committee on Ethical Practice of the Hospital for Sick Children, and informed written consent was obtained from parents.

The proximal 20 cm of the jejunum was perfused using a double-lumen tube at a rate of 8 ml/min as previously described. After a 30 minute equilibration period, effluent was collected on ice in 10 minute aliquots for a further 30 minutes. All the solutions perfused contained an amino acid or dipeptide, polyethylene glycol 4000 (3 g/l) (PEG, a water soluble non-absorbable volume marker), KCl (4 mmol/l), NaHCO₃ (25 mmol/l) their osmolalities were adjusted to 285–290 mosm/kg by the addition of NaCl; pH was adjusted to 7.0 by gassing with CO₂. After the 30 minute equilibration period PEG concentration in the three 10 minute aliquots of effluent was steady indicating that a steady state had been achieved. Amino acid and dipeptide concentrations were determined in each of the 10 minute collections as appropriate. The pH of the effluent did not differ significantly from the pH of the solutions perfused, and only trace amounts of peptidase activity could be detected in the effluent.

Transmural potential difference was monitored by a 23 gauge syringe needle placed in the inlet limb of the perfusion tube which served as a flowing intraluminal electrode. The reference electrode consisted of isotonic bicarbonate saline in a 23 gauge butterfly needle which was inserted under the skin of the leg. The intraluminal electrode and reference electrode were connected via isotonic bicarbonate saline bridges and disposable 5 ml syringes containing Ag/AgCl half cells to the input terminals of a battery powered voltmeter (Levell TM9B), the output was displayed on a chart recorder (Vitatron 2001). The asymmetry between the intraluminal and subcutaneous electrodes was measured before and at the end of the perfusion by placing the distal ends of the saline bridges in a solution of isotonic bicarbonate saline. The asymmetry potential was always less than 1 mV. Potential difference measurements are presented as the mean recorded during the steady state period, the mucosa being negative relative to the serosa. The ‘resting’ potential difference was recorded by perfusing normal saline before the first equilibration period.

Initially control and cystic fibrosis subjects were perfused with equimolar concentrations (56 mmol/l) of phenylalanine and lysine in random order. It was possible, however, that this concentration of the amino acids might saturate their transport systems and mask any more specific effects on their ‘carriers’ that might have been observed at concentrations approximating the apparent Kₘ of the amino acids concerned. No information could be found regarding the kinetics of phenylalanine absorption in vivo in man and thus a group of control subjects (n=4) were perfused with solutions containing 10, 20, 40, and 60 mmol/l of phenylalanine in ascending order of concentration. A 30 minute equilibration period was allowed between each concentration perfused and effluent collected for a further 30 minutes in 10 minute aliquots. The data revealed an apparent Kₘ and Vₘₐₓ of 24.4 mmol/l and 12.8 μmol/cm/min respectively. Data regarding the kinetics of lysine absorption in man in the literature were widely discrepant, however, Rey using a system similar to our own in young children found an apparent Kₘ of approximately 80 mmol/l, whereas a study in adults using different perfusion dynamics revealed an apparent Kₘ of approximately 5 mmol/l. Subjects were then perfused with lower concentrations of phenylalanine and lysine, 20 and 5 mmol/l respectively, which for phenylalanine was close to the apparent Kₘ in this system. As the concentration of lysine perfused in the first series of perfusions approximated the apparent Kₘ found by Rey in this second series of perfusions, a lower concentration (5 mmol/l) close to the apparent Kₘ found by Hellier et al was perfused. Plasma amino acid concentrations were determined in peripheral venous blood collected by venepuncture immediately before the perfusion, and at the end of the phenylalanine and lysine perfusion periods.

In the final part of the study each subject was perfused with solutions containing either phenylalanine, glycine, or glycyll-phenylalanine at 20 mmol/l, and as in all other subjects the order of perfusion of test solutions was randomised.
REAGENTS AND ANALYSIS
All reagents were of analytical grade and obtained from BDH Ltd, or Koch-Light Ltd. The amino acids and dipeptide were all found to be chromatographically homogenous.

Polyethylene glycol was assayed turbidometrically. Amino acids and dipeptide concentrations were estimated by ion exchange chromatography.

CALCULATIONS
Rates of absorption were calculated according to standard formulae. The disappearance rates of dipeptide and the absorption rates of the constituent amino acids were calculated by the following formulae:

\[
Dd = \frac{(Dp - De \times \frac{Peg p}{Peg e}) \times F}{S}
\]

\[
Aa = \frac{nDp - (nDe + Ca) \times \frac{Peg p}{Peg e} \times F}{S}
\]

where \(Dd\) = dipeptide disappearance rate in \(\mu\)mol/min/cm; \(Dp\) = concentration of dipeptide perfused; \(De\) = concentration of dipeptide in effluent; \(Peg p\) = concentration of polyethylene glycol perfused; \(Peg e\) = concentration of polyethylene glycol in effluent; \(F\) = rate of perfusion ml/min; \(S\) = segment length in cm; \(Aa\) = amino acid absorption rate in \(\mu\)mol/min/cm; \(n\) = number of the same amino acid residues in dipeptide; \(Ca\) = concentration of constituent amino acid in free form in effluent in \(\mu\)mol/ml.

Absorption and disappearance rates are assumed to be synonymous for free amino acids, whereas disappearance rates of dipeptide may include absorption of intact dipeptide as well as hydrolysis to constituent amino acids. For each subject, rate of absorption is expressed as the mean of the three 10 minute periods. The results for each group are expressed as the mean ± SEM and were analysed statistically by means of paired and unpaired Student's t test.

Results
The results of the initial part of the study are shown in Fig. 1. There was a marked reduction in phenylalanine absorption in cystic fibrosis patients at both 56 mmol/l (p<0.002) and 20 mmol/l (p<0.005) compared with the control values; phenylalanine (20 mmol/l) absorption was also depressed (p<0.05) in the patients with exocrine pancreatic insufficiency; in contrast, lysine absorption was not statistically different from control values.

The mean 'resting' potential difference was similar in the control cystic fibrosis and exocrine-pancreatic insufficiency groups (3.0±0.10, 2.8±0.15, 2.5±0.20 mV) respectively. The potential difference evoked by phenylalanine (56 mmol/l) was reduced in the cystic fibrosis patients (control 7.95±0.65; cystic fibrosis 4.06±0.62, p<0.002), as was the case at the lower concentration for both the cystic fibrosis (control 5.40±0.20; cystic fibrosis 3.9±0.21, p<0.005), and exocrine pancreatic insufficiency patients (control 5.40±0.20; exocrine pancreatic insufficiency 4.1±0.05, p<0.05). As reflected by the low potential difference values recorded, there was a significant reduction in water and Na+ absorption at the higher concentration of phenylalanine perfused and secretion of water and Na+ at the lower concentration of phenylalanine in the patients with cystic fibrosis and exocrine-pancreatic insufficiency as seen in Table 1. There was no difference in lysine evoked potential difference between the three groups at either concentration (56 mmol/l, control 8.90±0.50; cystic fibrosis 8.58±1.08; 5 mmol/l, control 5.2±0.19; cystic fibrosis 5.1±0.16; exocrine pancreatic insufficiency 5.13±0.12 mV). Lysine consistently provoked water and Na+ secretion to a similar degree, however, in both patients and control subjects (Table 1).
<table>
<thead>
<tr>
<th></th>
<th>Phenylalanine (56)</th>
<th>Lysine (56)</th>
<th>Phenylalanine (20)</th>
<th>Lysine (5)</th>
<th>Phenylalanine (20)</th>
<th>Glycine (20)</th>
<th>Glycyl-l-phenylalanine (20)</th>
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<td>Water Na⁺</td>
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<td></td>
<td>97·33 ± 10·78</td>
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<td>38·75 ± 3·14</td>
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<td>30·83 ± 3·60</td>
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<td>26·38 ± 13·10</td>
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<td>12·66 ± 16·92</td>
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<td>11·25 ± 0·66</td>
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<td>2·10 ± 0·92</td>
<td>0·66 ± 2·25</td>
<td>4·32 ± 0·54</td>
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<td>±2·5 ± 0·54</td>
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Figures in parentheses indicate concentrations of amino acid and dipeptide perfused (mmol/l). 
Net rate of water absorption in μl/min/cm. Net rate of sodium absorption in μmol/min/cm. 
—ve = secretion of water or sodium; values are mean ± SEM. 
Significance between test and control values: *<0·05; †<0·01; ‡<0·002; NS = not significant.
Table 2  Rise in plasma amino acid concentration after perfusion with phenylalanine (20 mmol/l) and lysine (5 mmol/l)

<table>
<thead>
<tr>
<th>Phenylalanine</th>
<th>Tyrosine</th>
<th>Lysine</th>
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<td>Tyrosine</td>
<td>Lysine</td>
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<tr>
<td>Controls (n=6)</td>
<td>570-65±30-42</td>
<td>67-60±11-49</td>
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<tr>
<td>†</td>
<td>†</td>
<td>NS</td>
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<tr>
<td>CF (n=6)</td>
<td>384-50±27-11</td>
<td>34-00±8-29</td>
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<tr>
<td>†</td>
<td>*</td>
<td>NS</td>
</tr>
<tr>
<td>EPI (n=3)</td>
<td>339-70±29-30</td>
<td>55-00±7-70</td>
</tr>
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</table>

Data expressed as mean ± SEM. Significance between test and control values:
* <0-05; †p<0-02; ††p<0-001.

Table 2 shows the increments in the concentration of plasma phenylalanine, and tyrosine after perfusion with phenylalanine, 20 mmol/l, and plasma lysine after perfusion with lysine, 5 mmol/l. As the rise in plasma phenylalanine during phenylalanine perfusion will be modified by significant metabolism of absorbed phenylalanine to tyrosine during its first pass through the liver, plasma tyrosine was also measured to give an indication of this. The rise in plasma phenylalanine and tyrosine concentrations was reduced in the cystic fibrosis and exocrine pancreatic insufficiency (p<0-001 and p<0-005 respectively) patients compared with controls, but there was no difference in lysine increments between the groups. The results of the final part of the study when phenylalanine, glycine, and glycy1-l-phenylalanine were perfused at concentrations of 20 mmol/l are shown in Fig. 2. Absorption of free phenylalanine was again found to be depressed in cystic fibrosis patients (p<0-02) as was absorption of the other neutral free amino acid glycine (p<0-01). The mean disappearance rate of glycy1-l-phenylalanine in cystic fibrosis patients was not significantly different from controls. During perfusion with phenylalanine and glycine, water and Na⁺ absorption was reversed to secretion in the patients with cystic fibrosis whereas during glycy1-l-phenylalanine perfusion, there was no significant difference in water and Na⁺ absorption between the cystic fibrosis patients and the control subjects (Table 1). The fact that absorption rates of phenylalanine and glycine from the dipeptide were normal or increased (glycine p<0-02) in cystic fibrosis supports the notion that intact dipeptide absorption was normal in the cystic fibrosis group.

Discussion

This study confirms and extends previous studies of impaired amino acid absorption in cystic fibrosis⁴ and represents the first in vivo perfusion study of amino acid absorption in cystic fibrosis. We have defined, for the first time to our knowledge, the kinetics of phenylalanine absorption in the jejunal of normal children in vivo and have shown that in cystic fibrosis there is reduced absorption of phenylalanine at concentrations which both saturate the transport system and are close to the apparent K_m of the study system used. These findings might suggest that in cystic fibrosis there are both reduced numbers of transport sites, and that they do not function as well as in control subjects. It would be necessary, however, to investigate phenylalanine absorption over a wider range of concentrations than has been studied here to confirm this suggestion. We have clearly shown impaired absorption of two structurally dissimilar neutral amino acids in cystic fibrosis, one polar-glycine, the other non-polar-phenylalanine; both are absorbed by the same group specific transport system. Previous studies have shown defective absorption of neutral amino acids leucine and cyclo-leucine in vitro⁵ and 3-methoxy-phenylalanine in vivo.⁶ It seems likely therefore that the neutral amino acid transport system is operating in a defective manner in patients with pancreatic insufficiency owing to cystic fibrosis. Neutral amino acid stimulated water
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and electrolyte absorption and its reversal to secretion together with the reduced transmural potential difference seen in the patients with cystic fibrosis and exocrine pancreatic insufficiency, can almost entirely be accounted for by the reduction in neutral amino acid absorption. The dibasic amino acid lysine, however, which is absorbed by a transport system quite distinct to that utilised by neutral amino acids, appears to be absorbed normally under the same conditions but to be quite certain a full kinetic study should have been carried out.

Perfusion with the dipeptide glycyl-l-phenylalanine showed that yet another transport system, which differs from both the dibasic and the neutral amino acid system, also appeared to be unaffected. The increased absorption of glycine from the dipeptide solution compared with the glycine solution in cystic fibrosis, and the abolition of the difference in absorption rates of both phenylalanine and glycine between cystic fibrosis and control subjects seen when the dipeptide was perfused provides further evidence that dipeptides are transported by separate systems from amino acids.

The nutritional significance of the defect in neutral amino acid transport is not clear. The products of luminal protein digestion are predominantly small peptides, in view of the normal dipeptide absorption it would seem unlikely that there would be much malabsorption of the products of digestion. It is known, however, that faecal nitrogen excretion is significantly increased in cystic fibrosis and that this is at least in part because of increased excretion of faecal amino acids.

The data cannot be explained in simple physical terms such as an alteration in the thickness of the unstirred water layer in pancreatic insufficiency states, as the defect in absorption seems to be restricted to neutral amino acids absorption of a dibasic amino acid, and a dipeptide being unaffected.

There are a number of possible explanations for the impaired neutral amino acid absorption in cystic fibrosis, including malnutrition, reduced pancreatic secretion or a defect specific to the disease itself. Malnutrition would appear to be an unlikely cause in our patients as growth was similar to control subjects. By altering the state of dissociation of the amino acids (and therefore their polarity), impaired pancreatic secretions of bicarbonate might theoretically be implicated as an explanation; for example, reduction of luminal pH decreases lysine absorption in the experimental animal. This is unlikely, however, to be the case, as in our patients the perfusates were buffered with bicarbonate and the pH of the solutions did not alter during perfusion. A previous in vitro study had suggested that jejunal malabsorption of neutral amino acids might be because of a defect intrinsic to cystic fibrosis. In our study the defect in phenylalanine absorption was also present in patients with other forms of pancreatic insufficiency suggesting perhaps that pancreatic insufficiency per se was involved. There is evidence that pancreatic proteolytic enzymes and mucosal lysosomal hydrolases play a role in the turnover of brush border proteins, and thus may modulate brush border function. It is tempting to speculate that the present data could be explained on the basis of a reduction in the number of brush border transport sites, which also functioned less efficiently.

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