Regulation of the absorption of dietary carbohydrate in man by two new glycosidase inhibitors

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SUMMARY Two new reversible inhibitors of intestinal α-glycosidases (BAY m1099 & o1248) have been derived from deoxynojirimycin. Their inhibitory substrate specificity has been investigated in man using test meals of the dietary carbohydrates, sucrose, maltose, and starch. Both inhibitors abolished the postprandial glycaemic rise after sucrose and m1099 50 mg did after maltose and starch, whereas o1248 20 mg had no effect after maltose and only a small effect after starch. Breath hydrogen evolution, as an indirect measure of malabsorption, showed that the reduced glycaemic responses, particularly after sucrose, were associated with considerable substrate malabsorption. Dose response studies showed that lower doses of both inhibitors could reduce postprandial glycaemia significantly without causing malabsorption. Both inhibitors were tolerated well. These two new enzyme inhibitors have different substrate specificity in man and can, in appropriate dose, regulate postprandial glycaemia by selective inhibition of brush border enzymes without causing malabsorption. In addition to their therapeutic importance, they provide a valuable experimental model of specific intestinal enzyme deficiency states.

The postprandial rise in blood glucose is determined by a number of dietary, gastrointestinal, hormonal, and metabolic factors. The amount of available carbohydrate, its physical form and complexity, the presence of dietary protein, fat and dietary fibre in the meal are all important dietary determinants of the rate of absorption. Abnormalities of the small intestinal mucosa and enzyme deficiencies can slow digestion and thus absorption. The rates of hepatic carbohydrate metabolism, insulin release, and peripheral glucose uptake also regulate blood glucose levels.

Awareness of the importance of close control of blood glucose concentrations in reducing the long term complications of diabetes stimulated research into new ways of stabilising postprandial glycaemia. One approach to this has been by the use of specific inhibitors of the intestinal α-glycosidase hydrolases, because all dietary carbohydrate, with the exception of lactose, is digested by these intestinal enzymes. There are many naturally occurring substances which inhibit these enzymes, but most are complex proteins or carbohydrates and are unstable, non-specific or irreversible.

The first pure synthetic glycoside hydrolase inhibitor was the pseudotetrasaccharide acarbose (Fig. 1a) which was isolated from culture of various genera of Actinomycetes. It is a stable compound and a powerful inhibitor of sucrase and maltase activity in vitro and in vivo in the rat and in man. It was the first inhibitor to be shown to have a reversible effect in the short term and it has no effect on monosaccharide transport mechanisms. Stable partial inhibition of sucrase activity can be obtained with this inhibitor in which the rate of onset and the steady state are dose related. In test meal studies in man with acarbose the glycaemic response to sucrose was greatly reduced, that to starch was moderately impaired, and to maltose was reduced slightly. The reduction in glycaemic response was associated with evidence of carbohydrate malabsorption.

Two new α-glycoside hydrolase inhibitors derived from 1-deoxynojirimycin have now been synthesised in microbial culture. Both nojirimycin (Fig 1b), synthesised by various species of Streptomyces and strains of the genus Bacillus, and its reduced form, 1-deoxynojirimycin (Fig 1c), are potent inhibitors.

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of mammalian intestinal brush border α-glycoside hydrolases in vitro, but not of α-amylase. Two derivatives of 1-deoxynojirimycin with modified side chains, designated BAY m1099 and o1248, have been produced (Figs 1d,e). Both are potent inhibitors of specific rat intestinal α-glycoside hydrolases in vitro. In studies of intestinal perfusion in the rat in vivo the inhibitors have been shown to have reversible effects and to have different patterns of substrate specificity.

In these studies the two new inhibitors have been used in man firstly to establish their substrate specificity and secondly to find dose levels at which postprandial blood glucose concentrations are controlled by slowing absorption, without causing malabsorption of dietary carbohydrate with its associated symptoms of flatulence, distension, and osmotic diarrhoea.

**Methods**

**Substrate Specificity Study**

Six healthy male subjects (mean age 30 years, range 27–33; 111%, range 84–130% desirable weight) took nine oral 50 g carbohydrate tolerance tests consisting of sucrose, maltose or starch with 400 ml water. In order to give a palatable and representative form of dietary starch, instant mashed potato was used (Winfield brand, 62 g containing 50 g starch, 4 g protein) made up with 200 ml hot water; with 200 ml cold water to drink. Each substrate was taken on three separate occasions with either m1099 50 mg, o1248 20 mg or a placebo of identical appearance at the start of the meal. These doses were chosen to be equipotent on the basis of the results of the animal studies. Tablets were given double blind in randomised order. The three tests for each substrate were done within a three week period with at least five days washout period between tests. Subjects followed the same routine of activity and meals on the day before each test, fasted overnight and took test meals at the same time on each occasion. Subjects were seated at rest for the first two hours of each test and were then allowed restricted activity over the next two hours. None of the subjects had any history of allergy or drug intolerance or had taken antibiotics in the previous month. No other medication was taken in the test period.

Finger prick blood samples were taken using Autolets (Owen Mumford Ltd, Woodstock, Oxon) into bottles containing 0.2 mg sodium fluoride and 0.6 mg potassium oxalate for measurement of blood glucose, using an enzymatic method detected by an oxygen electrode (Yellow Springs Instruments, Model 27). Finger prick samples were taken every 15 minutes from 0 to 60 minutes and then at 90 and 120 minutes. Venous blood was taken before and three hours after the start of each study for measurement of haematological indices and for biochemical measurements including urea, electro-
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lytes, calcium, phosphate, liver function tests, amylase, urate, cholesterol, and triglyceride.

End expiratory breath samples were taken using a modified Haldane-Priestley tube\textsuperscript{29} for the measurement of breath hydrogen concentration with a hydrogen electrode (Exhaled Hydrogen Monitor, GMI Medical Ltd, Renfrew). End expiratory samples were taken every 15 minutes from 0 to 150 minutes and then every 30 minutes to 240 minutes. A rise in breath hydrogen was taken as an indirect indication of malabsorption of carbohydrate.

A proportional relationship between the amount of carbohydrate malabsorbed and the hydrogen concentration in expired alveolar air has been reported.\textsuperscript{96-32} In an attempt to calibrate each subject’s malabsorption, a separate study was done in which end expiratory breath hydrogen concentration was measured after 25 g of the non-absorbable sugar lactulose. The ratio of the integrated breath hydrogen response (area under the curve) from the test meal to that from 25 g lactulose was used to estimate the amount of carbohydrate malabsorbed in the experimental situation for each individual.

Any symptoms or adverse reactions during the 24 hours after dosing were recorded by the subjects. The symptoms of distension, flatulence and loose motions which would be expected to be associated with malabsorption of carbohydrate were each recorded on a numerical scale of 0 (absent), 1 (mild), 2 (moderate) or 3 (severe).

**Dose response study**

In pilot studies sucrose absorption was found to be inhibited more than that of the other substrates in the presence of the inhibitors. In therapeutic use this would be the limiting substrate because it would be malabsorbed at lower inhibitor dose levels than other substrates. For this reason sucrose was used as the substrate in the dosage studies.

Six different healthy male subjects (mean age 28 years (23–30); 106% (98–121) desirable weight) took 50 g sucrose in 400 ml water on seven occasions. They took the following doses of inhibitors: m1099 12.5, 25 or 50 mg; o1248 5, 10 or 20 mg; or a placebo of identical appearance, double blind in randomised order. All other experimental details were as described above.

Statistical analysis was done using Student’s t test. Values given are means ± standard errors. The studies had the approval of the Ethics Committee of the Brent Health Authority.

**Results**

**Substrate specificity**

Both inhibitors abolished the postprandial glycaemic rise after sucrose 50 g and resulted in a considerable rise in breath hydrogen concentration from 45 minutes onwards. The results are shown in Figure 2 together with the mean symptom scores during the 24 hours after dosing for distension (D), flatulence (F) and loose motions (LM) for the two different inhibitors and placebo taken with sucrose. None of those with loose motions opened their bowels in the first two hours after dosing. The symptoms reported after o1248 were more severe than those after m1099. Breath hydrogen concentrations were also higher after o1248, though the differences between individual times or in the areas under the curves were not significantly different.

The effects seen after maltose were markedly different (Fig. 3). The postprandial glycaemic rise was reduced considerably by m1099 but o1248 had no significant effect. There was a small rise in breath hydrogen after maltose with m1099 but the symptoms reported were mild. Breath hydrogen levels and symptoms after o1248 and maltose were the same as after the placebo with maltose.

The postprandial glycaemic rise after starch was reduced moderately by o1248 and markedly by m1099 (Fig. 4). There was a small, late rise in breath

![Fig. 2 Blood glucose and breath hydrogen responses after sucrose 50 g with placebo, m1099 or o1248. Mean symptom scores for distension (D), flatulence (F) and loose motions (LM). (Glucose: 1 mmol/l =18 mg/100 ml).](http://gut.bmj.com/)
hydrogen after m1099 but not after o1248. The generally mild symptoms experienced were not significantly different between treatments and may well be a consequence of the bulk of the semisolid starch meal.

The amounts of the substrates which were malabsorbed in the presence of the two inhibitors, as calculated by the ratio of areas under the breath hydrogen curves, are shown in Table 1.

**DOSE RESPONSE**

The effects of the three different dose levels of m1099 on postprandial glycaemia after 50 g sucrose are shown in Figure 5 compared with the rise after

<table>
<thead>
<tr>
<th>Substrate</th>
<th>Sucrose 50 g</th>
<th>Maltose 50 g</th>
<th>Starch 50 g</th>
</tr>
</thead>
<tbody>
<tr>
<td>m1099 50 mg</td>
<td>38 ± 4</td>
<td>16 ± 9</td>
<td>5 ± 4</td>
</tr>
<tr>
<td>o1248 20 mg</td>
<td>49 ± 8</td>
<td>0 ± 1</td>
<td>3 ± 1</td>
</tr>
</tbody>
</table>
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sucrose with placebo. With increasing dose the postprandial rise is reduced and the peak delayed. The low dose (12.5 mg) caused no rise in breath hydrogen and no symptoms. The intermediate dose (25 mg) caused a slight rise in breath hydrogen but negligible symptoms. The high dose had similar effects in terms of breath hydrogen release and symptoms as had been found in the first study.

Even the lowest dose of o1248 (5 mg) caused a marked reduction in postprandial glycaemia, a moderate rise in breath hydrogen and some distension and flatulence. The 10 mg and 20 mg doses almost abolished the postprandial glycaemic rise, caused marked evolution of hydrogen and symptoms of similar severity at both dose levels (Fig. 6). The results at 20 mg were closely similar to those found with the first group of subjects in the substrate specificity study.

The amounts of the 50 g sucrose load which were malabsorbed in the dose response study, as calculated by the ratio of areas under the breath hydrogen curves, are given in Table 2. Both compounds were well tolerated by all subjects at all dose levels. The only symptoms of any severity which were reported were the predicted ones associated with the effects of fermentation of malabsorbed carbohydrate and osmotic diarrhoea. A few minor symptoms, such as headache and tiredness, were reported but these occurred as frequently in the placebo studies as after the inhibitors. There were no abnormalities of haematology or biochemistry found either before or after any of the tests in any of the volunteers, nor were there any consistent differences between the pre- and post-test values in any study.

Discussion

The results show that both these inhibitors are very effective in reducing postsucrose glycaemia. o1248 20 mg, however, has a relatively small effect on poststarch glycaemia and no effect on postmaltose glycaemia. The effects of o1248 are similar to those reported previously with acarbose 200 mg.11 In contrast, glycaemia after maltose and starch is reduced by m1099 50 mg very effectively without appreciable malabsorption being caused. The differences in substrate specificity suggest that there may well be differences in the inhibitors’ enzyme specificity in the intact gut. Both inhibitors are obviously effective against sucrase and m1099 appears to be effective against brush border maltases, other than sucrase, whereas o1248 seems to have negligible effect there. Inhibition of brush border maltase activity by m1099 would explain the reduction in glycaemia after both maltose and starch but the lack of effect of o1248 on maltose absorption suggests that its effect on starch absorption must be at a different level. These observations are in keeping with the findings of in vivo intestinal perfusion studies in the rat.23 25 but they contrast with the results of Lembcke et al22 who found that in vitro o1248 was as potent an inhibitor of isolated glucoamylase and maltase from mucosal homogenates as was m1099. Because maltose and maltopenta- and maltotetrosaccharides are end products of amylase digestion of starch, this suggests that the definite reduction of poststarch glycaemia by o1248 cannot be explained fully by a specific effect on brush border

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**Table 2** Amount of sucrose malabsorbed (g) from 50 g load at different inhibitor doses, calculated from hydrogen production with 25 g lactulose

<table>
<thead>
<tr>
<th>Dose (mg)</th>
<th>m1099</th>
<th>o1248</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>12.5±4</td>
<td>13±4</td>
</tr>
<tr>
<td>10</td>
<td>25±4</td>
<td>24±4</td>
</tr>
<tr>
<td>20</td>
<td>50±4</td>
<td>24±6</td>
</tr>
<tr>
<td>30</td>
<td>20±4</td>
<td>30±4</td>
</tr>
</tbody>
</table>

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**Fig. 6** Blood glucose and breath hydrogen responses after sucrose 50 g with placebo or o1248, 5 mg, 10 mg or 20 mg. Mean symptom scores for distension (D), flatulence (F) and loose motions (LM). (Glucose: 1 mmol/l =18 mg/100 ml)
maltases. It must therefore be due in part to an inhibitory effect on luminal amylase activity of whatever source. *In vitro* studies with purified α-amylase, however, have shown negligible inhibition of enzyme activity by m1099 or o1248 (Bayer AG, unpublished observations). The explanation of this paradox is not clear but may lie in the fact that both α-dextrinase and glucoamylase are active in hydrolysing malto-oligosaccharides with chain lengths up to nine or more glucose residues. These enzymes may contribute some ‘amylase-like’ activity to digestion of starch already partly broken down to shorter chain lengths in preparation. HPLC analysis of the potato, as eaten by the subjects, however, showed that less than 1% of the carbohydrate present consisted of chains of less than eight glucose residues.

The doses used in the substrate specificity studies were based on intestinal perfusion studies in the rat in which these doses were equipotent in inhibiting the disappearance of luminal sucrose. As the inhibitory effect of these doses appears to be almost total after sucrose 50 g, it is difficult to be sure whether this is a true equivalence in man. The dose response studies suggest that the high dose (20 mg) of o1248 was much more potent in man in inhibiting sucrase activity than was the high dose (50 mg) of m1099.

The potent sucrase inhibitory effect of both inhibitors has resulted in almost complete malabsorption of the carbohydrate load with its associated symptoms. The differences between the calculated amounts of sucrose malabsorbed in the presence of m1099 50 mg and o1248 20 mg in the two study groups are because of considerable individual variations in breath hydrogen evolution after lactulose and the inherently limited precision of this indirect test. These results must be treated with considerable caution, however, they have some value in giving an indication of the scale of malabsorption produced and the proportions for the two inhibitors are the same for both groups. There was some evidence of malabsorption of maltose with m1099 but not with o1248, though few symptoms were reported with either. There was no breath hydrogen evidence of starch malabsorption with either inhibitor but some symptoms were reported. Intestinal transit of the starch meal was probably much slower than that of the sugar solutions. Hydrogen evolution as an indication of malabsorption may thus have occurred later than the four hour collection period, as is hinted by the late rise in breath hydrogen after o1248. The volume of the meal and the dietary fibre present in the potato may both have contributed to symptoms reported over the 24 hour period. The amounts of hydrogen evolved after o1248 20 mg with sucrose were similar to those after acarbose 200 mg with sucrose. Like acarbose, o1248 did not cause malabsorption of maltose or starch. Despite the limitations of the technique, the dose response studies suggest that the amount of sucrose malabsorbed was broadly dose related. It seems that by using a lower dose of m1099 it is possible to regulate post-sucrose glycaemia without causing malabsorption, as was found with acarbose 50 mg, and it seems likely that an even lower dose of o1248 than 5 mg would have a similar effect. It is not only therapeutically undesirable to produce malabsorption of carbohydrate but in the regulation of diabetes it is essential to be confident that all carbohydrate that has been ingested is being absorbed and not lost to fermentation in the colon. Because sucrose is the most sensitive substrate for this, this is bound to be the limiting factor, unless it is totally excluded from the diet.

Maltose is not a commonly ingested dietary carbohydrate but is important in these studies because it is a major luminal digestion product of starch that is then hydrolysed at the brush border. The results suggest that the inhibitory effect of m1099 on poststarch glycaemia in man could be explained entirely by inhibition of maltase activity. In these studies, however, o1248 had negligible effect on maltose absorption but had a greater effect on starch absorption. This suggests that, despite the unpublished *in vitro* data, the compound appears to have some inhibitory effect on amylase activity *in vivo* in man.

The symptoms reported in these studies only occurred in the presence of the indirect breath hydrogen evidence of malabsorption of carbohydrate. In circumstances where carbohydrate uptake was slowed but complete, the only symptoms reported were minimal and occurred as frequently on placebo as they did in the presence of the inhibitors. It seems possible therefore to regulate the rate of carbohydrate uptake using these inhibitors and to reduce the postprandial glycaemic peak without causing undesirable malabsorption or symptoms. These compounds appear to have a therapeutic potential in the regulation of blood glucose in patients who have an impairment of carbohydrate metabolism and, in particular, in diabetes. The inhibitory properties of o1248 are similar to those of acarbose but it appears to be 10 times more potent than acarbose. The inhibitory profile of m1099 is appreciably different and its potency seems to lie between those of acarbose and o1248. The results suggest that m1099, with a broader spectrum of substrate specificity, may be a more effective therapeutic agent. They also suggest that, even if sucrose is excluded rigorously from the diet, a dose
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of m1099 25 mg at mealtimes may be sufficient to regulate postprandial glycaemia. In addition to their therapeutic potential, these derivatives of deoxynojirimycin have considerable scientific interest in that they provide a reversible experimental model of intestinal enzyme deficiency syndromes both in animals and man.

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