Low gluten diet in the treatment of adult coeliac disease: effect on jejunal morphology and serum anti-gluten antibodies

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SUMMARY Treatment of patients with coeliac disease with a low gluten containing diet (LGD) remains controversial. We have studied jejunal morphology and antigluten (AG) antibody titres by ELISA in patients on a LGD of 2.5-5 g/day for three to 14 months (median six months) and compared results with patients on a strict gluten free diet (GFD) for six to 27 months (median 13 months). We found no significant difference in villous height or crypt depth (eight LGD v 10 GFD patients) or serum AG-IgA, -IgG, and -IgM titres (13 LGD v 12 GFD patients). There was however, a significant increase (p<0.05) in intra-epithelial lymphocytes in those patients on a LGD. We conclude that adult coeliac patients can tolerate a LGD without gross morphological change and without initiating significant AG antibody responses.

After the discovery that gluten is involved in the pathogenesis of coeliac disease, the medical profession has maintained that a life long strict gluten free diet is mandatory for the health of all patients. Although there is no doubt that dietary gluten withdrawal results in improvement of clinical symptoms and jejunal morphology, the necessity for total dietary gluten exclusion has been challenged. Furthermore, despite physicians' recommendations, many patients are unable to maintain a restrictive GFD and continue to ingest small quantities of gluten. A proportion of these patients remain asymptomatic with normal haematological and biochemical parameters. In view of these findings we believe that a low gluten containing diet (LGD) may be a more realistic treatment for coeliac disease in some patients. This concept, however, remains controversial.

The aim of the present study was to determine the effects of a LGD on jejunal morphology and serum antibody levels to gluten in a group of coeliac patients we have maintained on a controlled LGD.

Methods

Patients

The following patients were included in the study: 13 (seven men) patients (median age, 40 years; range 17-74 years) with untreated coeliac disease, 12 of these patients (seven men) treated with a strict GFD, 13 (six men) coeliac patients (25 years; 18-70 years) on a LGD and 10 (four men) control patients (40 years; 17-74 years). The control patients consisted of patients with low serum folate concentrations caused by dietary deficiency, or patients with the irritable bowel syndrome with diarrhoea who required gastrointestinal investigation.

Treated coeliac patients had been on a 90-100% strict GFD for six to 27 months (median 13 months). Each of these patients assessed his or her own diet using a visual analogue scale and underwent formal dietary assessment by a dietician. Patients on a LGD received a list of foods, determined by a dietician, with each item containing 2.5 g portions of gluten. Low gluten diet patients had consumed 2.5 g or 5 g (three patients) of gluten per day for three to 14 months (median six months).

Diagnosis of coeliac disease was based on subtotal villous atrophy at first biopsy and subsequent
improvement of morphology on a repeat jejunal biopsy after a GFD. All patients had blood tests including a full blood count and serum and red cell folate.

**JEJUNAL BIOSPIES**

Jejunal mucosa from jejunum junction was obtained by Crosby capsule. Biopsies suitable for morphometric analysis were obtained from 12 of 13 of the untreated coeliac patients, from 10 of these patients during treatment, from eight patients on the LGD and from eight control patients. The control patients had jejunal biopsies to exclude coeliac disease because of a low red cell folate and gastrointestinal symptoms. In all cases intestinal morphology was found to be normal. Biopsies were fixed in 10% formal saline and then routinely processed and embedded for paraffin sections.

**JEJUNAL MORPHOMETRY**

Five micrometre sections of the jejunal biopsies were stained with haematoxylin and cosin. Villous height and crypt depth were measured using an eye piece micrometer. A minimum of 10 well orientated villi and crypts were measured per biopsy specimen. Intra-epithelial lymphocytes per 100 villous enterocytes were recorded, with a minimum of 500 enterocytes counted per patient. Slides were read blind.

**ANTIgluten ANTIBodies**

Serum antigluten IgG, IgM, and IgA were measured by microplate enzyme linked immunosorbent assay (ELISA). Crude gluten was dissolved in 70:30 solution of methanol: 0-1 M Tris buffered saline pH 7-6 to a final concentration of 40 mg/l. Fifty microlitres of this solution per well was used to sensitise 96-well polyvinyl plates at 4°C overnight. The eighth well of each column was sensitisised with 50 μl of 1% bovine serum albumin (BSA) as the control well. The plates were washed with PBS containing 0-1% Tween-20 three times and blocked with 1% BSA during a 60 minute incubation at 37°C.

Sera were diluted 1:50 in 0-1 M phosphate buffered saline (PBS) pH 7-6 containing 1% BSA and serial dilutions made in the plates. Each plate held a serum reference positive for antigluten antibodies. The control well contained the starting dilution of 1:50. After a 60 minute incubation, the plates were washed and 50 μl of 1:1000 dilution of antihuman -IgG, -IgA, or -IgM-peroxidase conjugate (Sigma, Poole, Dorset) in 1% BSA was added. Incubation was for 60 minutes followed by a further wash. 3,3',5,5'-tetramethylbenzidine (TMB) was used as peroxidase substrate. One hundred and fifty microlitres of 0-1 g/l solution of TMB in citrate acetate buffer (pH 6) containing 1-3 mmol H2O2 was added to each well, the plate incubated at room temperature for 30 minutes and the reaction stopped by the addition of 25 μl 2-35 M sulphuric acid. Optical density (OD) at 405 nm was read using an ELISA reader (Minireader II, Dynatech, Billinghurst, Sussex). A titre was taken as the reciprocal of the lowest dilution at which the OD reading was >0-05.

**STATISTICAL ANALYSIS**

Statistical evaluation of morphometric and ELISA data was carried out by means of a non-paired Wilcoxon’s test (non-parametric). Data obtained for patients before and after treatment on a gluten free diet were evaluated using a paired Wilcoxon’s test. This study had the approval of the district ethical committee.

**Results**

**MORPHOMETRY**

There was no difference in villous height or crypt depth between GFD and LGD patients. Villous height was lower (p<0.01) in untreated coeliac patients (median 57 μM; range 11-207 μM) compared with LGD patients (347; 171-430), GFD patients (295; 231-482) and control patients (422; 321-487). Villous height was higher (p<0.05) in control patients compared with GFD patients (Fig. 1). Some patients on a gluten free diet had low villous heights suggesting that they were in fact not adhering to a strict gluten free diet despite stating that they were.

Crypt depth was increased (p<0.01) in untreated patients (362; 222-482) compared with the LGD patients (146; 122-202) GFD patients (163; 121-220) and control patients (131; 93-170). Crypt depth was lower (p<0.05) in control patients compared with LGD and GFD patients.

There was an increase (p<0.05) in the IEL count of the LGD patients median 43/100 villous enterocytes; range 22-60) compared with GFD (30; 19-51) and control patients (25; 18-32). The IEL count was higher (p<0.01) in untreated patients (67; 54-86) compared with all other groups (Fig. 2).

**ANTIgluten ANTIBody TITRES**

AG-IgA titres in the control group were in the range 0-800 (median 200). Titres were raised in 11 of the 13 untreated coeliac patients (median titre, 1600; range 200-3200), but decreased after treatment and a strict GFD (200; 0-800; p<0.01). There was no difference in AG-IgA titres between GFD patients, LGD patients (200; 0-3200) and control patients (Fig. 3). One LGD patient had a persistently raised AG-IgA titre (3200), but subsequently admitted to ingesting more than 5 g gluten a day.
AG-IgG titres were higher (p<0.05) in the coeliac patients compared with the control group. There was, however, no difference in AG-IgG titres between the untreated coeliac patients LGD patients and GFD patients. There was no difference in AG-IgM titres between any of the study groups.

**Blood Tests**

In the coeliac group the serum folate concentrations in the untreated group rose from 1.9 μg/l (range 0.3 to 4.3) to 6 μg/l (range 0.6 to >18) after treatment. In only one patient did the serum folate remain below the normal range after treatment. The red cell folate rose from 134 μg/l (range 69 to 280) to 299 μg/l (range 64 to >640). In four patients the red cell folates remained below the normal range after treatment with a gluten free diet.

In the patients put on a low gluten diet the serum folate fell from 14 μg/l (range 2.5 to 14.5) to 6.3 μg/l (range 2 to 8.4). In only one patient did it remain below normal before and after the low gluten study started suggesting previous ingestion of gluten. The red cell folates were 303 μg/l (range 122 to 534) before and 262 μg/l (range 173 to 484) after the low gluten trial. There were no significant changes in haemoglobin concentration or serum B12 concentrations.

**Discussion**

Studies of humoral immune responses to gluten and its fractions in coeliac disease are numerous.\(^\text{12-18}\) Raised concentrations of serum antibody to gluten have been found in 85–100% of untreated patients,\(^\text{19,20}\) and this antibody is predominantly in the IgA and IgG classes.\(^\text{21,22}\) The introduction of a GFD results in a fall of antibody titres, but serum antigliadin-IgG may remain raised for many months.\(^\text{21,22}\) Serum AG-IgA, IgG, and IgM titres shown in this study are in accordance with these observations.

The similarity in AG antibody titres, between LGD and GFD patients suggests that patients can tolerate a regular gluten intake of 2.5–5 g/day without initiating humoral immune responses. Only one LGD patient had a raised AG-IgA titre (3200; Fig.
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3), and she later admitted to a gluten intake of more than 5 g/day. Our data further show that a LGD has no significant effect on gross mucosal architecture over the three to 14 month study period. While children and adolescents may take over 12 months to show morphological relapse in response to gluten challenge,23 Kumar et al24 have shown that adult patients relapse significantly more quickly after challenge with a minimum of 10 g gluten/day. Despite a lack of villous atrophy or crypt hyperplasia a small but significant increase in IELs was found in LGD patients. This observation supports previous work, in suggesting that an increase in IELs is a more sensitive indicator of gluten ingestion than partial villous atrophy.5 25

Although there is no doubt that a GFD is beneficial, we believe that the presence or absence of clinical abnormalities is dependent on the amount of gluten ingested, and that small amounts of gluten may be tolerated. McGrae et al26 followed 100 children with coeliac disease who had continued ingesting gluten for over a year after diagnosis. Within this population body weight, serum protein and magnesium were found to be normal and only two of these patients developed symptoms compelling enough to seek medical attention. In a recent study of 85 adult coeliac patients, we found that 27 patients were not maintaining a strict GFD despite the physician’s repeated recommendations. These patients were, however, found to be asymptomatic and had normal haematological and biochemical parameters.26 A parallel can be made with patients with dermatitis herpetiformis where many patients have jejunal mucosal abnormalities of varying severity and have no gastrointestinal symptoms.

In conclusion, we have shown that adult coeliac patients can tolerate a small regular intake of gluten without gross morphological change and without initiating significant AG antibody responses. There was, however, evidence of mild lymphocyte infiltration of the jejunal epithelium in LGD patients, the longterm clinical significance of which is uncertain.

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