Liver, biliary and pancreas

Effect of glucocorticoid on liver regeneration after partial hepatectomy in the rat

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SUMMARY  The increase in activities of hepatic thymidylate synthetase (EC 2.1.1.45) and thymidine kinase (EC 2.7.1.21), which catalyse the formation of thymidylate through the de novo and salvage pathways, respectively, were significantly suppressed during liver regeneration in rats which were given glucocorticoids (hydrocortisone and dexamethasone) or indomethacin. These drugs also prevented the augment of hepatic DNA content in 24 h regenerating liver.

Liver regeneration after two-thirds partial hepatectomy (PH) has been used as an ideal model to investigate the regulatory mechanism of mammalian cell proliferation in vivo. Previous studies have established that various humoral factors regulate DNA synthesis in the regenerating liver.1 As for the effect of glucocorticoids, Einhorn et al2 reported that cortisone diminished the DNA content in the regenerating liver and a similar conclusion was deduced by other workers.3,4 On the contrary, Rixon and Whitfield5 reported that hydrocortisone did not prevent DNA synthesis in the regenerating liver.

We have recently showed that the activities of thymidylate synthetase (TS: EC 2.1.1.45) and thymidine kinase (TK: EC 2.7.1.21), which catalyse the formation of thymidylate through the de novo and salvage pathways, respectively, reflected closely the total amount of DNA synthesis and that these enzymes were rate determining in DNA synthesis.5-10 The present experiments were designated to determine the effect of glucocorticoids on the basis of the activities of TS and TK in liver regeneration. This approach allowed us direct comparison of these enzymatic activities with the amount of DNA synthesis as our previous reports.11,12 The results show that the administration of glucocorticoids inhibits the increase of activities of TS and TK, resulting in the suppression of DNA synthesis in regenerating rat liver.

Methods

MATERIALS
(±)-L-Tetrahydrofolic acid was prepared by catalytic hydrogenation of folic acid according to the method of Huenekeens et al.13 [5-3H]-Deoxyuridine monophosphate (10-6 Ci/mmol) and [6-3H]thymidine (21.0 Ci/mmol) were purchased from Amersham (England). Dexamethasone and indomethacin were obtained from Sigma (USA). Hydrocortisone was from Nakarai Chemical Co (Japan). All other reagents were of analytical grade.

ANIMALS AND TREATMENTS
Male Wister rats (200–230 g) were used in this study. The animals were permitted free access to food (MF, Oriental Yeast Co, Osaka, Japan) and water at all times. Partial hepatectomy (PH) was performed under ether anaesthesia according to the procedure of Higgins and Anderson.14 Hydrocortisone in saline (100 mg/kg ip) or dexamethasone in ethanol (2 mg/kg ip) was injected one hour before PH or eight hours after PH. Indomethacin in ethanol (10 mg/kg ip) was administered 30 minutes before PH or eight hours after PH. Control animals received the same quantity of saline as did the experimental animals because no difference was observed in the effects of the vehicles.

ANALYTICAL METHOD
The rats were killed under ether anaesthesia at 24 hours after PH. Blood was withdrawn from the abdominal aorta and the liver was perfused in situ with saline. The calcium concentration in the plasma
of these blood samples was measured utilising commercially available diagnostic kits (Calcium test Wako, Japan). The excised liver was homogenised with 4 volumes of 50 mM Tris-HCl buffer, pH 7.3, containing 0.25 M sucrose, 10 mM β-mercaptoethanol, 1 mM phenylmethylsulphonyl fluoride and 1 mM EDTA. The DNA content of liver was measured by the diphenylamine reaction after extraction with trichloroacetic acid according to the procedure of Schneider. Protein was determined by the method of Lowry et al using bovine serum albumin as standard. The activities of TS and TK were determined as described previously and expressed as pmols of product formed/min/mg protein at 37°C. Statistical analyses of data were by Student's t test.

Results

Effects of glucocorticoids on 24 h regenerating rat liver

After PH the activities of TS and TK started increasing at about 12th hour and peaked at 24-48 h after PH. In the normal liver (taken just after PH and resting in G0 state), the activities of these enzymes were very low (Table) as in sham operated liver. When hydrocortisone (100 mg/kg ip) was injected one hour before PH, the TS activity in 24 h regenerating rat liver was increased considerably compared with the normal value but decreased to 42-3% of the control (hepatectomised) as shown in the Table. The TK activity of these rats also increased 2.56 times of the normal but decreased to 21.6% of the control (hepatectomised) (Table). Dexamethasone (2 mg/kg ip) given one hour before PH suppressed TS activity at 24 h after PH to about one third of the control (hepatectomised). Thymidine kinase activity diminished to 27-3% of the control (hepatectomised) value by the injection of dexamethasone (Table). These TS and TK activities were again higher than the corresponding level of the normal values. The hepatic DNA content at 24 h after PH in the control animals increased significantly compared with that just after PH (Table). In the rat pretreated with hydrocortisone or dexamethasone, the liver DNA content was significantly lower than that of the control (hepatectomised). The decrease of DNA content was in concert with the low enzymatic activities of TS and TK (Table).

The plasma calcium concentration of rats treated with the glucocorticoids was lower than that of the control at 24 h after PH (Table).

When dexamethasone was administered to rats at eight hours after PH similar effects were observed on the activities of TS and TK as well as liver DNA content. Plasma calcium concentration was also depressed by the treatment.

Effects of indomethacin on 24 h regenerating rat liver

Rats were treated with indomethacin (10 mg/kg ip) 30 minutes before PH and the regenerative responses were evaluated in 24 h regenerating liver. Activities of TS and TK raised significantly compared with those of the normal but considerably lower than those of the control rats. These were comparable with the values of the dexamethasone treated group. The quantity of liver DNA was diminished by the administration of indomethacin compared with the control group. Indomethacin was found to decrease plasma calcium level as hydrocortisone and dexamethasone. This was consistent with the literature.

When indomethacin was administered at eight hours after PH, similar effects on DNA synthetic responses and plasma calcium concentration were observed as shown in the Table.

Table Effect of hydrocortisone, dexamethasone and indomethacin on the activities of TS and TK, DNA content, and plasma calcium level at 24 h after partial hepatectomy (PH)

<table>
<thead>
<tr>
<th>Treatment</th>
<th>[injected time: h after PH]</th>
<th>Rats (n)</th>
<th>TS (pmol/mg protein/min)</th>
<th>TK (pmol/mg protein/min)</th>
<th>Liver DNA (mg)</th>
<th>Plasma calcium (mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (PH only)</td>
<td>17</td>
<td>67-53 (6-10)</td>
<td>220-94 (26-22)</td>
<td>6-99 (0-24)</td>
<td>9-45 (0-22)</td>
<td></td>
</tr>
<tr>
<td>Hydrocortisone [−1]</td>
<td>6</td>
<td>28-57 (5-47)†</td>
<td>47-83 (8-43)†</td>
<td>6-02 (0-31)*</td>
<td>7-75 (0-69)*</td>
<td></td>
</tr>
<tr>
<td>Dexamethasone [−1]</td>
<td>10</td>
<td>22-33 (3-40)†</td>
<td>60-25 (8-45)†</td>
<td>5-94 (0-39)*</td>
<td>7-84 (0-36)*</td>
<td></td>
</tr>
<tr>
<td>Dexamethasone [8]</td>
<td>6</td>
<td>19-61 (1-09)†</td>
<td>77-11 (4-08)†</td>
<td>5-78 (0-55)*</td>
<td>8-42 (0-34)*</td>
<td></td>
</tr>
<tr>
<td>Indomethacin [−0.5]</td>
<td>13</td>
<td>27-51 (2-91)†</td>
<td>68-29 (13-90)†</td>
<td>6-03 (0-27)*</td>
<td>7-73 (0-22)*</td>
<td></td>
</tr>
<tr>
<td>Indomethacin [8]</td>
<td>4</td>
<td>21-14 (3-66)†</td>
<td>73-38 (24-67)†</td>
<td>6-06 (0-09)*</td>
<td>8-37 (0-35)*</td>
<td></td>
</tr>
<tr>
<td>Normal (just after PH)</td>
<td>20</td>
<td>9-91 (0-99)†</td>
<td>18-69 (1-38)†</td>
<td>5-46 (0-31)†</td>
<td>10-49 (0-31)</td>
<td></td>
</tr>
</tbody>
</table>

Hydrocortisone (100 mg/kg ip) was injected one hour before PH. Dexamethasone (2 mg/kg ip) was injected one hour before or eight hours after PH. Indomethacin (10 mg/kg ip) was administered 30 minutes before or eight hours after PH. At 24 h after PH, the activities of TS and TK, DNA content and plasma calcium level were determined as described in the Methods section. Values are expressed as mean (SE). † † indicate significant differences from the control (* p<0.05; † p<0.01).
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Discussion

The results of this study showed that hydrocortisone and dexamethasone prevented the rise in the activities of TS and TK which are rate determining enzymes in DNA synthesis,6-10 with a concomitant decline in DNA content in the regenerating rat liver. These observations are compatible with the reports that showed diminished DNA content by cortisone,11,12 and with the results that described the decreased incorporation of labelled thymidine into DNA by glucocorticoids.13,14

The fact that the glucocorticoids inhibited DNA synthesis during liver regeneration suggests that one or more of the arachidionate derivatives are involved in the early process of regulation of TS and TK. Hydrocortisone and dexamethasone are known to prevent the release of arachidonic acid from phospholipids by promoting the synthesis of a peptide inhibitor of phospholipase.15 16 The involvement of the product(s) of arachidionate cascade system in the regulation of TS and TK may be supported by our observation that indomethacin, a potent inhibitor of fatty acid cyclooxygenase,17 showed similar effects to the glucocorticoids as shown in the Table. The ability of indomethacin to block DNA synthesis estimated by incorporation of [3H] thymidine is shown in the literature18-24 and these reports coincide with our results. MacManus and Braceland25 suggested prostaglandin E2 and F2α, while Kanzaki et al26 proposed thromboxane (not prostaglandin F2α) as a regulator of liver regeneration. It remains to be explored what arachidionate derivatives control liver regeneration.

In liver regeneration after PH the first transition of the hepatic cell from G0 to G1 phase seems to take place within four hours, during which c-fos27 and c-myc28 oncopogenes are expressed.29 The second signalling form G1 to S phase is assumed to occur at 10-12 hours.1 This is supported by the time dependence of α1-blockers3 to prevent the induction of TS and TK, which are S-phase specific enzymes.30 Glucocorticoids and indomethacin prevented the induction of TS and TK when applied before PH as shown above. When these drugs were administered at as late as eight hours after PH, quantitatively similar effects were observed on 24 h regenerating liver (Table). Therefore, these drugs may interfere with the events involved in S phase and/or the second transition of hepatic cells into S phase, just like α1-blockers.3 Further studies are needed to elucidate the relationship among signals caused by catecholamine,3 calcium10 11 and arachidionate derivatives in the regulation of DNA synthesis in liver regeneration.

Hydrocortisone, dexamethasone and indomethacin lowered plasma calcium level in partially hepa-

tectomised rats as shown in the Table. The effect of indomethacin was reported by MacManus and Braceland.30 Prolonged hypocalcaemia (more than 48 h) induced by thyroparathyroidectomy prevented the rise in TS and TK activities in 24 h regenerating rat liver but hypocalcaemic condition for 24 h did not affect the induction of TK.31 Because the duration and the degree of decrease of plasma calcium in the present experiment are less than those caused by thyroparathyroidectomy, the inhibition of TS and TK by the drugs cannot be ascribed only to decreased calcium ion. Rather, the direct effect on the hepatic functions – for example, prostaglandin synthesis, calcium mobilisation and calcium entry, may explain the inhibitory action of these chemicals on DNA synthesis in rat liver regeneration.

Finally it is worthwhile to note that care must be taken in the administration of glucocorticoids and indomethacin to patients with a liver disease, because these drugs possibly interfere with liver regeneration and are potentially hepatotoxic.

References

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