Agalactosyl IgG in inflammatory bowel disease: correlation with C-reactive protein

R Dubé, G A W Rook, J Steele, R Brealey, R Dwek, T Rademacher, J Lennard-Jones

Abstract
The proportion of oligosaccharide chains on the Fc fragment of IgG which terminate with N-acetylgalactosamine (GlcNAc) rather than galactose is increased in rheumatoid arthritis and tuberculosis, and in sera from patients with Crohn’s disease, probably because of decreased activity of a galactosyltransferase in B lymphocytes. We have assayed the prevalence of agalactosyl oligosaccharides on IgG in sera from 67 patients with inflammatory bowel disease (32 ulcerative colitis and 35 Crohn’s disease). The prevalence of agalactosyl IgG significantly increases in the majority of Crohn’s patients (19/35 patients), and correlates with the level of C-reactive protein (r=0.79), and inversely with the concentration of serum albumin. Sera from ulcerative colitis patients show less frequent (nine of 32) and less marked rises in agalactosyl IgG, and sera with high C-reactive protein values can contain normal levels. Thus in ulcerative colitis no correlation was seen between the two assays. The diseases in which the percentage of agalactosyl IgG is raised (rheumatoid arthritis, tuberculosis, Crohn’s disease and some ulcerative colitis) are characterised by simultaneous T cell mediated granulomatous tissue damage, and acute phase responses. Levels are normal in less tissue damaging granulomatous conditions, including sarcoidosis, and leprosy (except during episodes of erythema nodosum leprosum). We suggest therefore that a raised percentage of agalactosyl IgG is a correlate of a particular type of T cell mediated pathology which may be relevant to the pathogenesis of inflammatory bowel disease.

The fact that immunoglobulins are glycoproteins is often ignored. It has become clear, however, that changes in the composition of the sugar components of immunoglobulins correlate with certain clinical states, and may have important functional significance. Thus there is a conserved N-glycosylation site on the Fc of IgG, at asparagine 297 on the CH2 domain. This site bears essentially non-sialylated (90%) branched (biantennary) oligosaccharides, a variable proportion of which bear terminal galactose on one or both of their outer arms. When galactose is not present the terminal sugar is usually N-acetylgalactosamine (GlcNAc). In sera from normal donors the proportion of oligosaccharide chains bearing no terminal galactose (G0) decreases from a level of about 30% in small children to a trough of approximately 20% at the age of 25 years. Then it increases steadily with age to about 40% by 70 years.1 In rheumatoid arthritis there is an increased prevalence of G0 (percent-
assay of agalactosyl IgG has been described elsewhere. Briefly, mice were immunised with group A streptococcal cell wall peptidoglycan/poly saccharide complex, (as the group A polysaccharide bears much terminal GlcNAc), and monoclonals were selected for binding to fetuin treated with enzymes to remove sialic acid and galactose, (asialo-, agalacto- fetuin), resulting in exposed terminal GlcNAc. The antibodies were then further screened on agalactosyl IgG.

STANDARDS FOR THE IMMUNOASSAY FOR AGALACTOSYL IgG

Serum and IgG samples of which the galactose values had been obtained by a previously published biochemical procedure were used as standards throughout this study. Briefly, the N-linked oligosaccharides were released from the IgG using anhydrous hydrazine, and purified. The percentage of these oligosaccharides bearing no terminal galactose residues was then determined by measuring the hydrodynamic volume after exposure to an exoglycosidase mixture. The set of standards used was derived from normal donors, tuberculosis, and rheumatoid arthritis patients.

IMMUNOASSAY FOR AGALACTOSYL IgG

Using a template and sharp scalpel, nitrocellulose sheets were shaped into ‘combs’ with 12 teeth, spaced so that each tooth could enter a microtitre well, while the backbone of the comb was supported on the sides of the wells. The entire combs were then incubated for two to three days in protein A (P-6031, Sigma) at 250 μg/ml in phosphate buffered saline. The protein A-coated nitrocellulose was then washed in 1% PBS/Tween for two hours at room temperature.

Serum samples were diluted once in 50 in a buffer consisting of 0·1M glycine and 0·16M NaCl adjusted to pH 7·0 with NaOH to reduce aggregation of IgG. Aliquots of 0·25 ml were placed in flat bottomed microtitre wells in triplicate. The protein A-coated combs of nitrocellulose were placed in these wells and incubated at room temperature for four hours with occasional agitation. Then the combs were removed from the wells, washed twice in 1% BSA, 0·05% Tween 20 in PBS (PBS/BSA/Tween), once in PBS, and fixed in 0·5% glutaraldehyde in PBS for 30 min at 0°C. Fixed combs were washed again in PBS at 4°C containing 0·1M lysine, then boiled for five minutes in PBS in a double waterbath to denature the IgG and expose the sugars.

The nitrocellulose was then incubated on a rocking plate at room temperature for three hours with biotinylated anti-GlcNAc GN7 at a dilution of 1/2000 in PBS/BSA/Tween. After careful washing the nitrocellulose was incubated for two hours at room temperature in an avidin/peroxidase complex (Amersham) diluted 1/500 in PBS/BSA/Tween.

Subsequently the binding of peroxidase was revealed with a conventional mixture of hydrogen peroxide and precipitating chromogen (4-chloronaphthol, Sigma) in 5 mM Tris/HCl buffer at pH 7·6 for 15 minutes. After drying the tests were read with a transmitted light photometer adapted from a simple ELISA reader described previously.

CALCULATION OF RESULTS

A curve fitting program (Dataplot, by S M Fraser, Strathclyde University, Glasgow, UK) was used to plot the absorbance values yielded by the standards in the immunoassay, against the percentage of agalactosyl IgG previously determined biochemically. A log linear correlation was found (r = 0·94). The same program was used to calculate correlations, and to interpolate values for the unknown sera from patients with ulcerative colitis and Crohn’s disease. Repeat assays of unknown sera gave a mean variation of (4·7)%.

RESULTS

CHARACTERISATION OF THE PATIENTS STUDIED

Details of the patients studied are shown in Table I. The Crohn’s disease and ulcerative colitis groups were comparable in terms of age, sex ratio, duration of disease and disease activity (Harvey-Bradshaw index for Crohn’s disease (mean (SD)) was 3·3 (1·7); for ulcerative colitis, 2·97 (1·7)).

AGALACTOSYL IgG

Agalactosyl IgG was measured as described. The results are plotted against the age of the donor in Figures 1a, b, because of the known age variation discussed in the introduction. The Figure also shows the mean values for normal donors of the same age and two standard deviations either side of the mean. These normal values were taken from Parekh et al.’ The percentage of agalactosyl IgG in sera from 19/35 patients with Crohn’s disease fell well above 2×SD, whereas only nine of 32 ulcerative colitis samples did so (and four of these nine patients were borderline, Figure 1b). The difference between the diseases was significant, (p < 0·009, Fisher’s exact test) although the distribution of disease activity was similar.

<table>
<thead>
<tr>
<th>Characterisation of the patients studied</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crohn’s disease (n = 35)</td>
</tr>
<tr>
<td>Age (Mean and range)</td>
</tr>
<tr>
<td>History (yr and range)</td>
</tr>
<tr>
<td>Site of disease</td>
</tr>
<tr>
<td>Whole colon</td>
</tr>
<tr>
<td>Rectum to midtransverse</td>
</tr>
<tr>
<td>Left colon</td>
</tr>
<tr>
<td>Rectosigmoid</td>
</tr>
<tr>
<td>Ileocolonic</td>
</tr>
<tr>
<td>Ileocaecal</td>
</tr>
<tr>
<td>Ileostomy</td>
</tr>
<tr>
<td>Ileocaecal amputation</td>
</tr>
<tr>
<td>Treatment (age corrected the percentage of agalactosyl)</td>
</tr>
<tr>
<td>Predisone only</td>
</tr>
<tr>
<td>5-ASA/sulphasalazine only</td>
</tr>
<tr>
<td>Pred + 5-ASA/sulphasal</td>
</tr>
<tr>
<td>Azathioprine + other drugs</td>
</tr>
<tr>
<td>No treatment</td>
</tr>
</tbody>
</table>

*Not applicable; †Colon resected; ‡Terminal ileum resected; §Mean increase in percent agalactosyl IgG above the normal values for donors of the same age, in each treatment group (SD).
Agalactosyl IgG in inflammatory bowel disease

Figure 1: The percentage agalactosyl IgG (GO) for 35 patients with Crohn’s disease (Fig 1a) and 32 patients with ulcerative colitis (Fig 1b) plotted against the age of the donor. The black line indicates the mean percentage of agalactosyl for normal donors of the same age. The hatched area shows two standard deviations either side of that mean. Nineteen of 35 patients with Crohn’s disease, and nine of 32 patients with ulcerative colitis fall outside the 2×SD.

CORRELATION BETWEEN PERCENTAGE AGALACTOSYL IgG AND OTHER PARAMETERS OF DISEASE

Before seeking correlations with other clinical and biochemical parameters, each value was corrected for the age of the donor by subtracting the percentage of agalactosyl IgG anticipated in serum from normal donors of the same age (shown in Figure 1). The correlations sought are shown in Table II. In sera from Crohn’s disease there were significant positive correlations between CRP (r=0.79; p<0.005) and alpha-1-acid glycoprotein (r=0.69; p<0.01), and a significant negative correlation with serum albumin (r=0.389; p<0.05). The latter is not secondary to liver damage as a separate study of 13 patients with cirrhosis severe enough to cause dysfibrinogenemia has shown no abnormality of agalactosyl IgG concentrations. There was no correlation with the Harvey-Bradshaw index in either disease. In ulcerative colitis, however, there was a weak correlation (r=0.42; p<0.05) between the rise in the percentage of agalactosyl IgG and the platelet count.

The correlation with C-reactive protein in Crohn’s disease is shown in Figure 2. It can be seen that in sera from patients with undetectable C-reactive protein (and inactive disease) the values for the percentage of agalactosyl IgG fall around zero as expected after subtraction of the relevant normal value. The mean for these patients (SD) is 4.9 (8.3)/%.

Our previous observations on the clinical correlates of raised agalactosyl IgG, suggest that more than 2×SD higher than the mean value found in sera from normal donors of the same age. In contrast, in ulcerative sera, the percentage of agalactosyl IgG showed no correlation with C-reactive protein (Fig 3), which was high in some patients with a normal percentage of agalactosyl IgG. This difference between the two diseases was still apparent when only those Crohn’s disease patients with colonic disease without ileal involvement were included in the calculation since these eight patients had a mean percentage of agalactosyl IgG 19% (16-872) above the normal value, and four of eight fell more than 2×SD above the normal value.

Discussion

The present data confirm that the percentage of agalactosyl IgG markedly rises in Crohn’s disease, and show that this correlates with the rise in C-reactive protein, and the fall in serum albumin in this disease. In contrast, there is no significant rise in the percentage of agalactosyl IgG in the majority of cases of ulcerative colitis, and when there is a rise it does not correlate with C-reactive protein. Thus there are ulcerative colitis patients with striking levels of C-reactive protein who have a normal percentage of agalactosyl IgG, a situation not seen in Crohn’s disease. The clinical associations of raised percentage of agalactosyl IgG discussed in the introduction suggest that this glycosylation change occurs when there is T cell-dependent granulomatous inflammation with tissue damage and an acute phase response. Thus it is significant that previous authors have noted that the nature of the acute phase response differs in the two diseases.14 During the acute phase response there are changes in the glycosylation of acute phase proteins secreted by the liver, and cytokines released from macrophages have been implicated in an in vitro system.15 We speculate therefore that a cytokine released from macrophages activated by T cells may also be responsible for the down regulation of galactosyltransferase in B cells which is responsible for the change in glycosylation.4

Our previous observations on the clinical correlates of raised agalactosyl IgG, suggest that

<table>
<thead>
<tr>
<th>Albumin</th>
<th>WBC</th>
<th>Platelets</th>
<th>ESR</th>
<th>Clin Sc*</th>
<th>CRP</th>
<th>α-1-AGP†</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD</td>
<td>0.389</td>
<td>0.106</td>
<td>0.25</td>
<td>0.335</td>
<td>0.199</td>
<td>0.795</td>
</tr>
<tr>
<td>UC</td>
<td>0.306</td>
<td>0.028</td>
<td>0.42</td>
<td>0.204</td>
<td>0.306</td>
<td>0.164</td>
</tr>
</tbody>
</table>

*Clinical score (Modified Harvey-Bradshaw); †Alpha-1-acid glycoprotein; ‡p<0.05; §p<0.005; ¶p<0.001.

Figure 2: Correlation (r=0.79; p<0.005) between percentage agalactosyl IgG (%GO) and C-reactive protein in the sera of patients with Crohn’s disease. In order to correct for variation in the percentage of GO due to age, the mean value of the percentage of GO for normal donors of the same age as each patient has been subtracted.
it rises when there is chronic T cell mediated inflammation leading to, or accompanied by, an acute phase response, but not when there is an acute phase response without such a T cell component. Thus the percentage of agalactosyl IgG rises in rheumatoid arthritis and tuberculosis, both of which are characterised by T cell-mediated tissue damage and cytokine release. In rheumatoid arthritis there is release of cytokines such as IL-1, IL-6 and tumour necrosis factor into the joint, and circulation. These mediators are involved either directly or indirectly through release of IL-6, in the induction of the acute phase response. Similarly tuberculosis is accompanied by fever, weight loss, and necrosis in the lesions. This syndrome is readily explained by cytokine release triggered by the mycobacterial lipoarabinomannan. In contrast the percentage of agalactosyl IgG does not rise in sarcoidosis or uncomplicated leprosy where chronic T cell mediated granulomata are not accompanied by T cell mediated tissue damage (though there is damage secondary to nerve compression in leprosy) or a striking acute phase response. When there is an acute episode of T cell mediated pathology and an acute phase response, as during erythema nodosum leprosum, the percentage of agalactosyl IgG rises even in leprosy.

Moreover, because the percentage of agalactosyl IgG was not raised in sera from several acute virus infections, and we have recently found that it is not raised during acute rheumatic fever, and falls during pregnancy (Rook, Whyte et al, in preparation) it is likely that an acute phase reaction without chronic T cell-mediated pathology does not lead to the formation of agalactosyl IgG. Thus the difference between Crohn’s disease and ulcerative colitis noted in the present study could be related to the known difference in the nature of the acute phase response in the two diseases, or to differences in the nature of T cell function which are suggested by the greater prevalence of granuloma formation in Crohn’s disease.

Finally, the occurrence of raised percentage of agalactosyl IgG in tuberculosis, rheumatoid arthritis, and Crohn’s disease is provocative as one is a known mycobacterial disease, and these organisms, or autoantigens cross reactive with them, are implicated by some authors in the others.

We are grateful to G D Searle Ltd for financial support.

Agalactosyl IgG in inflammatory bowel disease: correlation with C-reactive protein.

R Dubé, G A Rook, J Steele, R Brealey, R Dwek, T Rademacher and J Lennard-Jones

Gut 1990 31: 431-434
doi: 10.1136/gut.31.4.431

Updated information and services can be found at:
http://gut.bmj.com/content/31/4/431

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Topic Collections
Articles on similar topics can be found in the following collections

Ulcerative colitis (1113)
Crohn's disease (932)

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/