Increased ethane exhalation, an in vivo index of lipid peroxidation, in alcohol-abusers

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Abstract
Ethane exhalation was measured in 42 control subjects, 52 patients with various non-alcoholic liver diseases, and 89 alcohol abusers who had been admitted to hospital for alcohol withdrawal and assessment of liver disease (six with normal liver tests, 10 with steatosis with or without fibrosis, six with alcoholic hepatitis, 29 with cirrhosis, 34 with both cirrhosis and alcoholic hepatitis, and four with both cirrhosis and a hepatocellular carcinoma). Ethane exhalation was similar in control subjects and in patients with non-alcoholic liver diseases, but was five times higher in alcohol abusers. Ethane exhalation in alcohol abusers was significantly, but very weakly, correlated with the daily ethanol intake before hospital admission, and the historical score for steatosis, but not with the inflammation or alcoholic hepatitis scores. Ethane exhalation was inversely correlated with the duration of abstinence before the test. In nine alcoholic patients, the exhalation of ethane was measured repeatedly, and showed slow improvement during abstinence. Ethane exhalation was significantly but weakly correlated with the Pugh’s score in patients with alcoholic cirrhosis. It is concluded that the mean ethane exhalation is increased in alcohol abusers. One of the possible mechanisms may be the presence of oxidizable fat in the liver. The weak correlation with the Pugh’s score is consistent with the contribution of many other factors in the progression to severe liver disease.

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Experimental data support a role of free radical mechanisms, including lipid peroxidation, in ethanol induced cellular injury in animals. The question of whether lipid peroxidation occurs in alcoholic patients, however, and if it does of its possible mechanism(s) and importance in the development of liver lesions, remains largely unanswered at present. Hepatic levels of conjugated dienes (an early product of lipid peroxidation) were found to be higher in 16 patients with alcoholic liver disease than in eight patients with non-alcoholic liver disease. Hepatic thiobarbituric acid reactants (malondialdehyde and other products of lipid peroxidation) were higher in 13 heavy drinkers with fatty livers (studied after about seven days of abstinence) than in five non-drinkers with histologically normal livers. In another study, serum and hepatic thiobarbituric acid reactive substances were found to be slightly increased in patients with chronic liver diseases, but did not show any difference between patients with chronic alcoholic liver disease and patients with chronic non-alcoholic liver disease.

Peroxidation may also be assessed non-invasively by determining the exhalation of volatile hydrocarbons (pentane and ethane) in breath. Pentane and ethane are formed during the process of lipid peroxidation and are exhaled in breath (see Discussion). A difficulty with pentane is that it is extensively oxidised in the liver. In patients with liver disease, therefore, an increased exhalation of pentane may mean either an increased production rate (peroxidation), or a decreased hepatic metabolism. In contrast, ethane undergoes little hepatic metabolism, so that its exhalation mainly reflects its formation rate. An increased exhalation of pentane, but not of ethane, was found in 15 patients with alcoholic cirrhosis compared with 21 healthy controls. The duration of abstinence before breath sampling, however, was not mentioned. Yet another method of assessing in vivo lipid peroxidation is the measurement of serum vitamin E concentrations (vitamin E is consumed during peroxidation). Serum vitamin E concentrations, however depend on several other factors, including diet, absorption of this fat soluble vitamin, and serum lipid values. A decreased serum vitamin E concentration in chronic alcoholic patients has been found in two studies. In another study, however, the serum vitamin E/serum cholesterol ratio was found to be normal in 21 of 25 patients with chronic alcoholic liver disease.

The reasons for the apparent discrepancies in tests of lipid peroxidation in alcoholic patients in the published reports are not completely understood. They may include the inconsistent occurrence of lipid peroxidation in alcoholics, the small numbers of patients studied, differences in the liver diseases studied, and failure to take account of the duration of abstinence before the various measurements.

In view of these conflicting data, we decided to reinvestigate this question by a non-invasive, reliable test (the exhalation of ethane) in a large series of patients admitted to hospital for both alcohol withdrawal (thus enabling correct appreciation of the duration of abstinence) and assessment of the liver disease. Results were compared with a large group of controls, and with a large series of patients with various non-alcoholic liver diseases. In an effort to understand better the possible mechanism of increased lipid peroxidation in alcoholic patients and its significance in the development of liver lesions, correlations were looked for between the exhalation of ethane and several clinical and histological parameters.
**Methods**

**PATIENTS**

Three groups of subjects were included in this study, which took place between July 1989 and February 1991. The first group included 42 healthy volunteers recruited mainly from the laboratory team or clinical staff. This group included 22 men and 20 women, aged 22 to 67 years, with a mean (SD) age of 40 (8) years.

The second group consisted of 52 patients with non-alcoholic liver diseases. These were 29 men and 23 women, aged 21 to 78 years with a mean (SD) age of 49 (16) years. This group included two patients with a transplanted liver; six patients with acute hepatitis (two viral, four drug induced); 11 patients with chronic hepatitis (five caused by hepatitis B virus, four hepatitis C virus, and two autoimmune); 17 patients with viral cirrhosis (six caused by hepatitis B virus, and 11 by hepatitis C virus); one patient with polyadenomatosis of the liver; five patients with non-alcoholic hepatocellular carcinoma; two patients with liver metastases; five patients with sclerosing cholangitis; one with primary biliary cirrhosis; and two with extrahepatic bile duct obstruction.

The third group included 89 alcohol abusers admitted to hospital for both alcohol withdrawal and assessment of the liver disease (45 men, 44 women), aged 23 to 78 years, with a mean age (SD) of 49 (12) years. This group included six patients with normal hepatic tests, 10 patients with hepatic steatosis associated or not with hepatic fibrosis, six patients with alcoholic hepatitis, 29 patients with cirrhosis but without alcoholic hepatitis, 34 patients with both hepatic cirrhosis and alcoholic hepatitis, and four patients with alcoholic cirrhosis and hepatocellular carcinoma.

A liver biopsy specimen was obtained during the current hospital stay in the 83 alcohol abusers with abnormal liver tests. The time interval between ethane measurement and liver biopsy was less than 1 week in 75 patients. The liver histology was subsequently reviewed by one pathologist for semiquantitative grading of the following lesions: necrosis, hyaline necrosis, Mallory's bodies, microvacuolar steatosis, microvesicular steatosis, intralobular inflammation, inflammation in portal tract or in fibrosis, portal and perportal fibrosis, centrilobular fibrosis, hepatic vein lesions, iron content, cholestasis, and hepatic architecture. A global inflammation score was also calculated as the mean of the scores for intralobular inflammation and inflammation in portal tracts or fibrosis. A global hepatic alcoholic score was calculated as the mean of the various individual scores pertaining to necrosis, hyaline necrosis, Mallory's bodies, and inflammation. The following parameters were also recorded: age, sex, acknowledged daily ethanol intake before hospital admission, body temperature, ascites, encephalopathy, jaundice, oedema, mean corpuscular volume, polymorphonuclear white blood cells, serum bilirubin, serum alanine (ALT) and aspartate (AST) aminotransferase activities, alkaline phosphatase and γ-glutamyl transferase activities, serum albumin, serum iG, plasma prothrombin level. Biological parameters were measured within 4 days of the measurement of ethane exhalation. The Pugh's score was calculated in cirrhotic patients.

**MEASUREMENT OF ETHANE EXHALATION**

Smokers were asked to abstain from smoking for 3 hours before the test. After explanation and acceptance of the test, the subjects were asked to first exhale normally, and then to exhale forcefully into an empty 2:5 litre gas sampling bag, thus providing alveolar air. Another bag was filled with ambient air from the room to measure the concentration of ethane in the air inhaled by the subject.

The concentration of ethane in air was measured as previously reported by Letertón et al., by gas-liquid chromatography apparatus (Packard model 437 A), equipped with a flame ionisation detector. Peak areas were integrated and calculated on a Packard recorder (model 604) with a range set at 1 and an attenuation factor set at 1 (no attenuation). Briefly, samples of air were injected through a six-way valve and a 5 ml sample loop, in a 100 × 0:16 cm id stainless steel column packed with Carbosieve G (60/80 mesh). The column and detector temperatures were set at 170°C and 200°C, respectively. The flow rate of nitrogen was set at 20 ml/minute. Under these conditions, the retention time of ethane was about 3 minutes. Standard dilutions of pure ethane (Air Liquide, France) in air gave linear responses over a range of concentrations that exceeded both ways the range of concentrations observed in the test samples. For each test, a standard calibration was made with a dilution of ethane (8500 pmol/l air). The ethane exhaled by the subjects was calculated as the difference between the concentration of ethane in the air exhaled by the subject and that in the air inhaled (means of three injections from each gas sampling bag). The intraassay coefficient of variation was less than 10%.

**STATISTICS**

Mean ethane exhalation in the three different groups of subjects – that is, healthy subjects, non-alcoholic patients, and alcohol abusers – was compared by one way analysis of variance followed by Dunnett's t test. Univariate regression analysis was performed to assess possible relationships between ethane exhalation and the various parameters examined.

**Results**

Ethane exhalation was not different in the 42 healthy subjects and the 52 patients with non-alcoholic liver diseases (Fig 1). Ethane exhalation did not differ by analysis of variance among the various subgroups composing the non-alcoholic group (Table 1). In contrast, the exhalation of ethane was significantly increased in the 89 alcohol abusers (Fig 1), the mean value in this group being fivefold that found in healthy subjects. The exhalation of ethane in the 89 alcohol abusers was significantly but very weakly correlated with the acknowledged daily intake of...
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Figure 1: Exhalation of ethane in 42 control subjects, 52 patients with various non-alcoholic liver diseases, and 89 alcohol abusers. The exhalation of ethane, mean (SEM), was assessed as the difference between the concentration of ethane in alveolar air and that in ambient air. *p<0.0001, significantly different from that in controls.

Cholestasis

TABLE I  Ethane exhalation in 52 patients with various non-alcoholic liver diseases

<table>
<thead>
<tr>
<th>Disease</th>
<th>Ethane exhalation (pmol/l air)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Transplanted liver (n=2)</td>
<td>366, 25</td>
</tr>
<tr>
<td>Acute hepatitis (2 viral, 4 drug induced)</td>
<td>116 (20)*</td>
</tr>
<tr>
<td>Chronic hepatitis (9 viral, 2 autoimmune)</td>
<td>70 (15)*</td>
</tr>
<tr>
<td>Viral cirrhosis (n=17)</td>
<td>232 (49)*</td>
</tr>
<tr>
<td>Polycystic disease (n=1)</td>
<td>149</td>
</tr>
<tr>
<td>Liver carcinoma (5 hepatocellular carcinoma, 2 liver metastasis)</td>
<td>95 (85)*</td>
</tr>
<tr>
<td>Cholestasis (5 sclerosing cholangitis, 1 primary biliary cirrhosis, 2 extrahepatic biliary obstruction)</td>
<td>159 (32)*</td>
</tr>
</tbody>
</table>

* Values are mean (SEM).

ethanol (Fig 2). It was not significantly correlated with the body temperature, the presence of ascites, encephalopathy, jaundice, or oedema, the serum bilirubin concentrations, serum albumin, or plasma prothrombin, the mean corpuscular volume, or the polymorphonuclear white blood cell counts, the activities of alkaline phosphatase or γ-glutamyl transferase, the individual values for AST or ALT activities, or the AST/ALT ratio. Ethane exhalation was significantly but very weakly correlated with the Pugh's score in the 67 alcoholic patients with liver cirrhosis (with or without alcoholic hepatitis or hepatocellular carcinoma) (Fig 2).

Analysis of variance failed to show any significant difference in the ethane exhalation among the various histological subgroups detailed in Table II. Ethane exhalation was significantly but very weakly correlated with the histological score for macrovacuolar steatosis (Fig 2) but was not correlated with any of the other liver lesions directly quantitated by the pathologist (see Methods) nor with the global inflammatory score or the global alcoholic hepatitis score.

Ethane exhalation was inversely correlated with the duration of abstinence, as assessed by the time interval between the onset of hospital admission and the first measurement of ethane exhalation (Fig 3). In nine patients, the first ethane exhalation measurement (reported in Table II) was followed by repeated measurements (Fig 3). In all of these patients, the exhalation of ethane decreased with the duration of hospital stay, and thus abstinence (Fig 3). This decrease was usually slow, however, occurring over a period of several weeks (Fig 3).

Discussion

The initial step in lipid peroxidation is the abstraction, by a drug free radical or an oxygen radical (R·), of a hydrogen atom from an unsaturated lipid (lipid-H), forming RH and a lipid radical (lipid-·). This radical quickly adds molecular oxygen to form a peroxy radical (lipid-O2·). In the presence of Fe2+, the peroxy radical may form an alkoxyl radical (R1-CHO·R2), which may split into an alkyl radical (R1·), such as the pentyl or ethyl radicals, and an aldehyde (CHO–R2). A hydrogen atom is abstracted by the pentyl or ethyl radical to form pentane or ethane, respectively. Pentane either is hydroxylated by hepatic cytochrome P–450, or is exhaled in breath.
related to relatively long lived morphological and/or biochemical lesions induced in the liver by alcohol. Because non-alcoholic liver lesions were not associated with increased ethane exhalation (Fig 1), possible causes should be sought in lesions more specifically associated with alcohol ingestion.

A major abnormality characteristically (if not exclusively) associated with alcoholism is the presence of polymorphonuclear cells in the inflammatory infiltrate. Polymorphonuclear cells are activated at sites of inflammation and release superoxide anion. However, they also release the iron binding protein, lactoferrin, which limits the extracellular formation of the hydroxyl radical. Stimulated human polymorphonuclear leukocytes do not cause peroxidation, unless iron salts are added to the medium. Instead, their toxicity to hepatocytes seems related to the release of proteases. In the present study, there was no relationship between the exhalation of ethane and the semi-quantitative assessment of the inflammatory infiltrate.

A second lesion, which is rather characteristic of alcoholic liver disease is steatosis. Fat present in the liver may serve as a substrate for lipid peroxidation. Continuing experimental studies in our laboratory clearly show that hepatic steatosis produced by ethanol and a variety of unrelated compounds (aminopterin, tetracycline, various tetracycline derivatives, valproic acid, and pirprofen) is associated in each case with increased lipid peroxidation. In the present study, a significant relationship was observed between the exhalation of ethane and the semiquantitative estimation of macrovascular steatosis (Fig 2). Taken together, these experimental and human data suggest that the presence of fat deposits in liver cells may be one of the mechanisms leading to lipid peroxidation in alcoholic abusers. The correlation with steatosis was very weak, albeit statistically significant (Fig 2), however, suggesting that other factors may be involved as well.

Indeed, two biochemical modifications not assessed in our study may contribute to lipid peroxidation during the early phases of abstinence:

(a) Normally 2% of the oxygen consumed by mitochondria is transformed into superoxide anion and other reactive oxygen intermediates.

(b) Alcohol consumption induces cytochrome P-450 2E1. Increased activity of this isoform in alcoholics may persist for several days to weeks after stopping alcohol abuse. Cytochrome P-450 2E1 is peculiar by its marked ability to spontaneously form the hydroxyl radical. Increased production of oxygen radicals as a consequence of mitochondrial changes and the induction of cytochrome P-450 2E1 may have contributed to increased ethane exhalation in alcohol abusers with normal liver tests (Table II). However, because a liver biopsy was not performed in these patients, the

Figure 2: Exhalation of ethane as a function of, respectively, the acknowledged daily ethanol intake in the 89 alcohol abusers, the macrovascular steatosis score in the 83 alcohol abusers with abnormal liver tests and a histological examination of the liver, and the Pugh's score in the 67 patients with alcoholic cirrhosis. (Without or with alcoholic hepatitis or hepatocarcinoma). r=correlation coefficient; p=probability.

Figure 3: Inverse relationship between the exhalation of ethane, and the duration of hospital stay in the 89 alcohol abusers (r=correlation coefficient; p=probability), and evolution of ethane exhalation in the nine alcohol abusers who were studied repeatedly.
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<table>
<thead>
<tr>
<th>TABLE II Ethane exhalation in 89 alcohol abusers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ethane exhalation (pmol/l)</td>
</tr>
<tr>
<td>Normal liver tests (n=6)</td>
</tr>
<tr>
<td>Seatosis, with or without fibrosis (n=10)</td>
</tr>
<tr>
<td>Alcoholic hepatitis (n=6)</td>
</tr>
<tr>
<td>Cirrhosis (n=29)</td>
</tr>
<tr>
<td>Cirrhosis and alcoholic hepatitis (n=34)</td>
</tr>
<tr>
<td>Cirrhosis and hepatocellular carcinoma (n=4)</td>
</tr>
</tbody>
</table>

* Values are mean (SEM).

The presence of minor liver lesions, such as steatosis, cannot be excluded.

Although the exhalation of ethane was on average increased fivefold in alcohol abusers (Fig 1), there was wide variability in this group (Table II, Fig 2). Indeed in a few alcohol abusers, there was no detectable production of ethane (that is, the ethane concentration in expired air was similar to that in inspired air).

This variability may reflect both variability in alcohol consumption, in liver lesions, and in the time of measurement, as well as variability in protective mechanisms. Lipid peroxidation is efficiently prevented by vitamin E (α-tocopherol), which transforms lipid peroxyl radicals into hydroperoxides, and the latter are then transformed into lipid alcohols by glutathione peroxidase. In the process, however, α-tocopherol is converted to the α-tocopheroxyl radical, α-tocopheryl quinone, and other products, while reduced glutathione (GSH) is oxidised to the disulphide. As a consequence of lipid peroxidation, and other factors, hepatic vitamin E \(^{11,12}\) and GSH levels \(^{3}\) are low in some alcoholics. Depletion of these protective factors may, in turn, permit the unrestrained development of lipid peroxidation in some patients.

Experimentally, lipid peroxidation has several deleterious effects on the liver. It damages membranes, including the plasma membrane, leading to increased entry of calcium into the cell, \(^{13,14}\) decreased sequestration of calcium in endoplasmic reticulum, and mitochondria, \(^{15,16}\) and liberation of lysosomal enzymes. \(^{17}\) Lipid peroxidation activates phospholipase A\(_2\), with the release of toxic lysophospholipids, free fatty acids, leukotrienes, and other eicosanoid metabolites. \(^{18} \) It forms reactive aldehydes (4-hydroxyalkenals and malondialdehyde), which react with hepatic proteins. \(^{19}\) In the present study, however, the exhalation of ethane was similar in patients with cirrhosis and those with other liver lesions (Table II). Admittedly, progression to cirrhosis should depend not only on the instantaneous degree of a presumed pathogenic event (in this case, lipid peroxidation), but even more on the duration of this process, so that a similar instantaneous value might be seen in patients with incipient lesions, but not with full blown cirrhosis. Even in the group of cirrhotic patients, however, ethane exhalation was only weakly correlated with the Pugh’s score (Fig 2). This observation is consistent with the view that many other factors participate in the development of severe liver disease in these patients, including immunological mechanisms, nutritional deficiencies, mitochondrial injury, altered redox status, impaired protein processing, cyto-

skeletal abnormalities, activation of Ito cells, and circulatory disturbances. \(^{20,21}\)

In summary, the average exhalation of ethane was increased fivefold in a large hospital series of alcohol abusers, but was normal in a group of patients with various non-alcoholic liver diseases. Although the increased exhalation of ethane in alcohol abusers slowly improved after alcohol withdrawal, it was still high during the first week(s) of abstinence, suggesting that it was not related, at that time, to the production of free radicals during the metabolism of ethanol. One of the possible mechanisms may be the presence of increased amounts of oxidisable fat in the liver. The weak correlation with the Pugh’s Score in alcoholic patients with cirrhosis is consistent with the contribution of many other factors in the development of severe liver disease in such patients.


26 Spach PI, Cunningham CC. Control of state 3 respiration in liver mitochondria from rats subjected to chronic ethanol consumption. *Biochim Biophys Acta* 1987; 894: 460–7.


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