Enteric protein loss and intestinal permeability changes in children during acute shigellosis and after recovery: effect of zinc supplementation

A N Alam, S A Sarker, M A Wahed, M Khatun, M M Rahaman

Abstract
The effect of zinc supplementation on intestinal permeability changes and protein loss was studied in 32 children aged between 1 and 12 years during bouts of acute shigellosis and after recovery. An intestinal permeability test and then a 48 hour balance study were performed on all patients. They were then blindly assigned to receive vitamin B syrup either with or without zinc acetate (15 mg/kg per day) for a month. All patients received a five day course of nalidixic acid. The balance study was repeated during convalescence and follow up, but a permeability test was done only at follow up after one month. Intestinal permeability, expressed as a urinary lactulose:mannitol excretion ratio, improved significantly (p=0.001) along with a significant increase (p=0.005) in mannitol excretion in the zinc supplemented children, suggesting a resolution of small bowel mucosal damage. The latter was associated with a higher coefficient of nitrogen absorption (p=0.03), suggesting a possible role of zinc in the treatment of shigellosis. Enteric protein loss, as assessed by faecal α1 antitrypsin clearance, was not influenced by zinc supplementation.

(Gut 1994; 35: 1707-1711)

Shigellosis is still a major cause of morbidity and mortality, particularly among younger children in developing countries. In one recent study, there was an abnormal transmucosal protein loss in many of these patients. The pathological changes are confined to the colon and are characterised by erythematous and oedematous friable mucosa covered with an adherent mucopurulent layer. These changes result in loss of mucus and blood along with serum protein, and may continue even during the recovery period. Absorption of nitrogen compared to absorption of other nutrients was minimum in acute shigellosis. However, it could not be determined conclusively if the unabsorbed amount of nitrogen was related to protein loss other than failure of intestinal absorption. Intestinal permeability changes reflecting the integrity of the small intestinal mucosa have been reported in both acute and persistent diarrhoea. The extent and duration of enteric protein loss and its possible association with changes in intestinal permeability have, to our knowledge, not yet been studied in shigellosis. A metabolic balance study was therefore conducted together with determination of faecal clearance of α1 antitrypsin (an indirect measure of enteric protein loss) and excretion of lactulose and mannitol in children with shigellosis.

Zinc is a micronutrient that is commonly deficient in children in developing countries. Loss of zinc occurs in infants with acute diarrhoea. Furthermore, hypozincemia and low rectal mucosal zinc concentrations have been shown in cases of childhood chronic diarrhoea, although it was not clear whether the mucosal injury associated with these conditions resulted in such a loss. Zinc supplementation in patients with diarrhoea has been shown, however, to improve the mucosal integrity and is associated with significant reduction in diarrhoea attack rates, duration of diarrhoea, and stool volume, particularly when the patients are undernourished. The present study also aimed to explore the inter-relationship between gut permeability changes and enteric protein loss and to determine if zinc supplementation affects these changes.

Methods
Forty three boys and girls aged 1-12 years with a history of bloody mucoid diarrhoea of less than three days' duration were initially enrolled in the study. Microscopic examination of the stools of these patients suggested shigellosis. Children with obvious systemic illness – for example, pneumonia, meningitis, leukaemoid reaction, severe malnutrition (<65% weight for age according to the National Center for Health Statistics), otitis media, and distension of the abdomen – were excluded from the study. All the patients had a detailed clinical examination on admission to hospital. Rectal swabs and stool samples were obtained from each patient on admission and were plated on MacConkey agar, Salmonella-Shigella agar, and taurocholate-tellurite-gelatin agar. The plates were examined for Salmonella and Shigella by standard methods.

### Table 1 Composition of study diet (per 100g)

<table>
<thead>
<tr>
<th>Name of food</th>
<th>Protein (g/100g)</th>
<th>Zinc content (mg/100g)</th>
<th>Energy (kcal/100g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Milk suji</td>
<td>2.09</td>
<td>0.23</td>
<td>92.7</td>
</tr>
<tr>
<td>Bread</td>
<td>7.44</td>
<td>0.10</td>
<td>267.5</td>
</tr>
<tr>
<td>Egg (whole)</td>
<td>12.31</td>
<td>1.26</td>
<td>160.4</td>
</tr>
<tr>
<td>Egg yolk</td>
<td>14.25</td>
<td>4.00</td>
<td>436.5</td>
</tr>
<tr>
<td>Albumin (egg)</td>
<td>14.56</td>
<td>0.14</td>
<td>124.0</td>
</tr>
<tr>
<td>Banana</td>
<td>1.81</td>
<td>0.14</td>
<td>124.0</td>
</tr>
<tr>
<td>Rice</td>
<td>2.12</td>
<td>0.33</td>
<td>120.0</td>
</tr>
<tr>
<td>Chicken curry</td>
<td>14.26</td>
<td>0.72</td>
<td>296.7</td>
</tr>
<tr>
<td>Breast milk</td>
<td>1.0</td>
<td>0.03</td>
<td>56.2</td>
</tr>
</tbody>
</table>

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TABLE II  Characteristics of study patients at the time of admission to hospital with shigellosis (values mean (SD))

<table>
<thead>
<tr>
<th></th>
<th>Group I (zinc supplement: n=16)</th>
<th>Group II (no zinc supplement: n=16)</th>
<th>p value</th>
<th>Healthy control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (mth)</td>
<td>62 (34)</td>
<td>63 (31)</td>
<td>0.60</td>
<td>11/8 (2-4)</td>
</tr>
<tr>
<td>Boy/girl</td>
<td>13/3</td>
<td>12/4</td>
<td></td>
<td>n=30</td>
</tr>
<tr>
<td>Duration of diarrhoea (d)</td>
<td>6-0 (2-0)</td>
<td>5-0 (2-1)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Duration of fever (d)</td>
<td>6-0 (2-0)</td>
<td>5-0 (2-1)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Packed cell volume (%)</td>
<td>36-5 (6-3)</td>
<td>37-7 (4-9)</td>
<td>0.01</td>
<td></td>
</tr>
<tr>
<td>% Weight for age (NCHS)</td>
<td>76-2 (13-0)</td>
<td>73-7 (8-5)</td>
<td>0.48</td>
<td>n=150</td>
</tr>
<tr>
<td>% Weight for height (NCHS)</td>
<td>88-6 (10-5)</td>
<td>88-5 (6-0)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>% Height for age (NCHS)</td>
<td>91-6 (8-1)</td>
<td>91-6 (5-2)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Shigella isolates were then biochemically identified and serotyped by a slide agglutination test using commercially available antisera (Burroughs Wellcome Research, Triangle Park, North Carolina, USA). Written informed consent was obtained from parents or guardians of children included in the study. The study was approved by the Human Ethical Review Committee of the International Centre for Diarrhoeal Disease Research, Bangladesh.

An intestinal permeability test using a freshly prepared solution containing 5 g lactulose with 0.5 g lactose (7.5 ml of Duphalac, Duphar Labs, Southampton, UK) and 1 g mannitol made up to 20 ml with 1% chloroform water was performed using the method described by Behrens et al., starting soon after admission when rehydration had been accomplished. All patients were hydrated and were made to empty their bladder before the test dose was offered. Adhesive paediatric urine bags (Downs Ltd, London, UK) containing a drop of 20% (vol/vol) chlorhexidine gluconate to prevent bacterial degradation of the probes were applied to the clean perineum to collect all urine samples over the next five hour period for subsequent measurement of the markers by an automated enzyme assay using Cobas-bio (Switzerland).

After this test, two charcoal tablets (500 mg, homogenised in 15 ml water) were given orally. Once the charcoal marker had appeared in the stool and the patients were clinically settled after full rehydration, a 48 hour balance study (first balance study), as previously described, was begun with a diet of known compositions (Table I). Food was weighed to an accuracy of 0.1 g on a Toledo scale (Ohaus, Dial Oq) and was offered freely. The amounts offered and left over were measured and the difference was recorded as the amount consumed. None of our patients was breast fed. All patients received nalidixic acid (55 mg/kg per day in four divided doses for five days), as Shigella species isolated from their stools or rectal swabs, or both, were sensitive to nalidixic acid. In a randomised, double blind manner, patients were also assigned to receive a vitamin B preparation (B1, 1.2 mg, riboflavin 3.0 mg, nicotinamide 6.0 mg, B6 0.6 mg, and calcium D-pantothenate 6.0 mg in 5 ml) either with or without zinc acetate (15 mg/kg per day with an elemental zinc of 3.9 mg/kg per day) given in three divided doses. The preparations were provided in suspensions that were identified by code numbers in order to mask the administration. It was well known that the requirement for zinc changes with age and that the availability of zinc from diets varies widely. Recommendations for the daily requirement of zinc range from 5 to 22 mg daily. A predominantly cereal-legume-vegetable-based diet containing phytates and, possibly, other chelating substances in the study population, presence of undernutrition, and increased losses of zinc in the diarrhoea stool and sweat led us to use a larger amount of zinc to replace the perceived deficit in zinc status. The mean dietary zinc intakes were 0.3-3.0 mg/kg per day and 0.285 mg/kg per day in the supplemented and the un-supplemented groups respectively. A second 48 hour balance study was performed on day eight at the end of the nalidixic acid therapy. Patients were discharged from hospital and asked to continue the zinc/placebo suspension at home for a further period of three weeks and then return for follow up testing. A permeability test was then repeated and a third balance study was performed. Transit time was measured as the time interval between feeding the charcoal marker and its first appearance in the stool during the three balance studies.

On admission to hospital and on day 8, venous blood was obtained for a complete blood count and estimation of serum electrolytes, creatinine, total protein, albumin, alkaline phosphatase, serum zinc, and serum ζ1 antitrypsin levels. Another blood sample was taken on day 30 to repeat the measurements of serum zinc, alkaline phosphatase, serum ζ1 antitrypsin, total protein, and albumin.

Total protein was estimated as total solids using a refractometer (American Optics Co), albumin by a modified colourimetric method using a spectrophotometer, and alkaline phosphatase was estimated using a commercial (BDH) kit.

Serum zinc concentration was estimated
Enteric protein loss and intestinal probes

Lactulose:

<table>
<thead>
<tr>
<th>Permeability probes</th>
<th>Day 1</th>
<th>Day 30</th>
<th>Statistical significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lactulose:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No zinc supplement</td>
<td>0.41 (0.23-0.73)</td>
<td>0.27 (0.18-0.43)</td>
<td>NS</td>
</tr>
<tr>
<td>Zinc supplement</td>
<td>0.47 (0.32-0.70)</td>
<td>0.33 (0.19-0.58)</td>
<td>NS</td>
</tr>
<tr>
<td>Mannitol:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No zinc supplement</td>
<td>1.36 (0.85-2.20)</td>
<td>2.71 (1.33-4.8)</td>
<td>&lt;0.005</td>
</tr>
<tr>
<td>Zinc supplement</td>
<td>1.15 (0.71-1.89)</td>
<td>1.32 (0.52-3.31)</td>
<td>NS</td>
</tr>
<tr>
<td>L/M ratio:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Zinc supplement</td>
<td>0.30 (0.16-0.52)</td>
<td>0.09 (0.08-0.14)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>No zinc supplement</td>
<td>0.40 (0.24-0.68)</td>
<td>0.25 (0.14-0.46)</td>
<td>NS</td>
</tr>
</tbody>
</table>

with an atomic absorption spectrophotometer (Pye Unicam SP9) as described by Smith et al.\textsuperscript{16} and the serum and faecal concentrations of α\textsubscript{1} antitrypsin were estimated as described by Crossley and Elliott.\textsuperscript{17} Faecal nitrogen content was measured according to the Micro-Kjeldahl procedure.\textsuperscript{18} The coefficient of absorption of N\textsubscript{2} was calculated using the following formula:

\[
\text{Coefficient of } N_2 \text{ absorption } N_2 = \frac{N_2 \text{ intake} - N_2 \text{ output}}{\text{Intake of } N_2} \times 100
\]

Faecal clearance of α\textsubscript{1} antitrypsin was calculated\textsuperscript{3} using the formula shown below:

\[
C = \frac{F + W}{P} \text{ ml/d}
\]

Where C = faecal clearance of α\textsubscript{1} antitrypsin, expressed as ml serum/d; F = faecal concentration of α\textsubscript{1} antitrypsin (mg/g); W = daily faecal weight (g); and P = serum concentration of α\textsubscript{1} antitrypsin (mg/ml).

**STATISTICAL ANALYSIS**

The Student’s t test was used to compare the mean values of the two study groups when the distribution of data was normal. Data on intestinal permeability tests for both groups that were not normally distributed were log transformed before statistical analysis, and the anti-log was expressed as the geometric mean, with 95% confidence interval. The Student’s paired t test was done for comparison of two data points among the same group of subjects.

**Results**

Only 32 of the 43 patients were eligible for analysis, since the cultures of stool samples or rectal swabs were negative for *Shigella* species in the remaining 11 patients. Admission characteristics (Table II) of the study patients in the two groups were comparable. Table III presents the laboratory findings in the study patients. The serum total protein and albumin concentrations improved in both groups of patients with clinical recovery. The mean serum zinc concentration and alkaline phosphatase activity increased significantly in the zinc supplemented group on day 8 during the second balance study and these changes maintained a similar trend during recovery (third balance study). Alkaline phosphatase accurately reflected total body zinc status\textsuperscript{16} in the study patients; activity decreased with zinc deficiency and significantly increased with zinc supplementation. Serum α\textsubscript{1} antitrypsin was raised in both study groups in the acute stage, but decreased during clinical recovery. The mean duration of diarrhoea after hospital admission was shorter and the mean cumulative stool volume during hospitalisation was lower in the zinc supplemented group (137 ± 146 hours and 28 ± 31 gm/kg per day), but the differences were not significant. Children in both groups made similar gains in height and weight. The effect of zinc supplementation on intestinal mucosal permeability is presented in Table IV. On admission day, no differences in probe markers between the study groups were observed. Lactulose excretion decreased after four weeks in both the groups, being more marked in the supplemented group. In the latter group, there was significant increase in mannitol excretion (p=0.005) at follow up. As a result, the lactulose:mannitol ratio improved significantly (p=0.001) towards normal value(s) in this group (analysis of variance was done with two way interaction terms including syrup and day, which was found to be statistically significant at 10% level). On the contrary, no such improvement was noticed in the unsupplemented group. An increased faecal clearance of α\textsubscript{1} antitrypsin was observed (Table V) in both study groups during the first balance period (acute stage). These values decreased significantly in both groups during the early convalescent and after recovery but failed to attain normal values of less than 20 ml/d\textsuperscript{20} in either group. Between the two groups, no difference in the faecal clearance of α\textsubscript{1} antitrypsin was observed during the three balance periods. Total energy and nitrogen intake during the balance periods, as well as total nitrogen loss in stool, urine, and vomitus, were similar in the two study groups. The mean transit time varied from 8-2 to 9-3 hours during the acute stage, 10-6 to 13-2 hours during the convalescent period, and 16-7 to 19-9 hours after recovery in the zinc supplemented group. In the other study group, transit time varied from 9-3 to 11-4 hours, 11-7 to 16-5 hours, and 14-1 to 16-2 hours during the three stages of illness. The increase in transit time from acute to the recovery stage was significant (p=0.01) in both the study groups. No significant relationship was however, found between the transit time and coefficient of nitrogen absorption or between the transit time and urinary mannitol excretion during any stage of the illness. During the second balance period, nitrogen intake was marginally higher in the supplemented group (Table VI) with significant improvement in the

<table>
<thead>
<tr>
<th>48 h balance study</th>
<th>Group I (zinc supplement) (n=16)</th>
<th>Group II (no zinc supplement) (n=16)</th>
<th>Statistical significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Study 1 (day 1)</td>
<td>161-0 (4.0-753.3)</td>
<td>160-0 (9.2-604.3)</td>
<td>NS</td>
</tr>
<tr>
<td>Study 2 (day 5)</td>
<td>59-0 (6.0-160.3)</td>
<td>35-1 (9.2-130.4)</td>
<td>NS</td>
</tr>
<tr>
<td>Study 3 (day 30)</td>
<td>57-0 (10.0-109.0)</td>
<td>27-0 (3.2-170.1)</td>
<td>NS</td>
</tr>
</tbody>
</table>
apparent nitrogen absorption (p=0.04). The coefficients of nitrogen absorption were similar during the first (acute) balance period in both study groups, and it was significantly better (p=0.03) in the zinc supplemented group during the second balance period. This differential effect was not maintained after recovery, however, when nitrogen absorption returned towards normal levels in both groups. Overall, a significant negative correlation (r=−0.52, p=0.001) was observed (Figure) between nitrogen absorption and faecal clearance of α1 antitrypsin.

Discussion

The results of the study show that zinc plays an important part in intestinal mucosal regeneration and in the improvement of mucosal function. Higher lactulose and lower mannitol excretion with a higher lactulose:mannitol ratio in the study patients at hospital admission, compared with healthy Bangladeshi children suggested the presence of intestinal mucosal damage in shigellosis. It is well known that zinc repletion hastens regeneration of the damaged gut epithelium since zinc is essential for DNA, RNA, and protein synthesis. A significant increase in mannitol excretion with improvement in the lactulose:mannitol ratio after recovery, as was observed only in the supplemented group, suggests improved absorptive function after zinc supplementation. Lactulose excretion was, however, reduced in both study groups. Similar observations have been made in diarrhoea of other aetiologies. The improved absorptive mucosal function allowed significantly better nitrogen absorption in the zinc supplemented group during the convalescent period. Permeability tests done simultaneously would have further clarified the role of zinc in shigellosis. This was an obvious limitation of the study. Nalidixic acid therapy with resolution of symptoms would not explain the changes seen after zinc supplementation as specific treatment was offered to both groups of patients. Zinc supplementation failed to show any effect on enteric protein loss in shigellosis since a similar, but steadily decreasing, faecal clearance of α1 antitrypsin was observed in both groups. The changes were more marked, however, in those supplemented. The clearance values did not attain normal levels even after recovery at one month, suggesting that the mucosal damage caused by the Shigella group of organisms and the protein-losing enteropathy might continue even after apparent clinical recovery. There was a significant negative relationship between nitrogen absorption and faecal clearance of α1 antitrypsin, suggesting a decreased antitrypsin clearance with increased nitrogen absorption in the supplemented group. This further indicates that estimation of only one parameter cannot act as a proxy indicator for another.

The present study clearly show that independent of gut transit time, zinc improves the absorption of nitrogen early in the course of shigellosis when the patient remains anorectic and there is derangement of serum protein homeostasis. This effect of zinc may be highly beneficial to young children with the illness whose dietary protein intake is marginal or inadequate in the presence of an increased protein requirement and severe protein loss in the stool. Our study has failed to show any significant difference on height and weight gain in the supplemented children but the importance of long term elemental supplementation to attain such an effect cannot possibly be ruled out.

Watery diarrhoea frequently precedes the dysenteric symptoms in acute shigellosis. Shiga toxin has been shown to cause fluid production in rabbit ileal loops. Mobsassaleh et al have shown evidence for a glycolipid receptor for shiga toxin involved in the fluid secretory response in rabbit small intestines. Small bowel involvement in shigellosis has been best demonstrated in the rhesus monkey model where the organisms proliferate first in the proximal small bowel lumen and cause a local

Table VI

<table>
<thead>
<tr>
<th>Time</th>
<th>N2 intake (g/kg day)</th>
<th>Total N2 excretion (g/kg day)</th>
<th>N2 balance (g/kg day)</th>
<th>Coefficient N2 absorption (% intake)</th>
</tr>
</thead>
</table>
|                         | Group I | Group II | (faeces+urine+vomit)          | Group I | Group II | (faeces+urine+vomit) | Group I | Group II | (faeces+urine+vomit) |...
| Balance study (day 1)   | 0.381 | 0.432 | 0.319 | 0.369 | 0.062 | 0.063 | 0.199 | 0.259 | 0.432 | 0.578 | 0.618 | 0.777 | 0.412 | 0.354 | 0.206 | 0.223 | 0.493 | 0.457 | 0.978 | 0.972 | 1.00 | 1.00 |...
secretory diarrhoea without invading the bowel mucosa. Jejunal transport abnormalities observed in the experimental animals with the disease could not be attributed to the enterotoxin mediated process alone. Evidence for similar changes in human small bowel are not definitive. The present study also raises the possibility of small intestinal mucosal changes associated with enteropathy in shigellosis, although this is considered to be primarily a colonic disorder.

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9 Sachdev HPS, Mital NK, Yadav HS, Serum and rectal mucosal zinc levels in acute and chronic diarrhea. Indian Pediatr 1990; 27: 125–33.
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