Increased leucocyte adhesiveness/aggregation is a most useful indicator of disease activity in patients with inflammatory bowel disease


Abstract
The aim of the study was to determine the comparative usefulness of inflammatory markers, in evaluating disease activity in patients with inflammatory bowel disease. Disease activity was assessed by the Mayo Clinic score for ulcerative colitis, and Harvey-Bradshaw score for Crohn's disease. Five hundred normal blood donors who had no underlying inflammatory condition served as controls. The erythrocyte sedimentation rate, platelet and white blood cell count, C reactive protein, and the leucocyte adhesiveness/aggregation test (LAAT) were determined in each patient. One hundred and twenty four patients with inflammatory bowel disease were tested while in remission and 128 in relapse. Their mean (SD) per cent of aggregated white blood cells in the peripheral blood was 8 (5) and 17 (10) respectively compared with controls 6 (4) (p<0.0001). Moreover, the LAAT could effectively discriminate between various grades of disease activity, the values in patients with active disease being 13 (6)% in mild, 17 (10)% in moderate, and 26 (10)% in severe disease (p<0.0001). Other acute phase reactants including the erythrocyte sedimentation rate, the white blood cell count, the platelet count, neutrophil count, as well as, the C reactive protein concentration did not differentiate as well between the various groups. Using logistic regression analysis to differentiate between inflammatory bowel disease patients in remission or relapse, the LAAT was the single best indicator. The addition of any other test did not contribute to the discrimination. Among the different laboratory variables tested only the LAAT significantly discriminated between the five different subgroups of controls, remission and mild, moderate or severe disease activity.

Methods

Patients
Two hundred and ninety nine blood samples were examined in 160 patients with IBD (84 men and 76 women). Ninety eight patients had CD and 62 had UC. The mean (SD) age was 33.2 (17.2) and 38.5 (18.2) respectively (range 8–76). Disease activity was assessed by the Mayo Clinic score for UC, and the Harvey-Bradshaw score (Mayo-Clamp modification) for CD. Five hundred normal age and sex matched volunteers, who had no underlying inflammatory condition served as controls. All clinical scores and biochemical indices for both Crohn's disease (CD) and ulcerative colitis (UC) have been developed; along with the physician's and patient's assessment. These scores, however, were found to be unsatisfactory; partly because of their complexity and partly because of their heavy dependence on subjective symptoms that may be changed by mechanical factors (for example, strictures, resection, etc) rather than by disease activity. The various laboratory indices were also found to be imperfect.

While the extremes in the clinical condition (that is, remission or severe disease) are easily recognised in both UC and CD, minor changes of disease severity are more difficult to detect or quantify. The large number of proposed scores and indices suggest that none is ideal. In this study we have evaluated prospectively, in a large cohort of IBD patients and controls, the usefulness of several common biochemical markers in estimating disease activity.

Keywords: disease activity, inflammatory bowel disease, leucocyte adhesiveness/aggregation test.

Knowledge of the severity of inflammation in inflammatory bowel disease (IBD) is important for evaluation, treatment, and prediction of prognosis. To quantify disease severity several

Aggregated leucocytes in the peripheral blood of a patient with severe Crohn's disease.
TABLE I  Laboratory values obtained in patients with IBD and controls

<table>
<thead>
<tr>
<th></th>
<th>ESR (mm/h)</th>
<th>CRP (mg%)</th>
<th>WBCC (10^3/mm³)</th>
<th>PMN (%)</th>
<th>PLT (10^9/mm³)</th>
<th>LAAT (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls (n=500)</td>
<td>16 (15)</td>
<td>0-5 (0-3)</td>
<td>6-7 (0-8)</td>
<td>57 (15)</td>
<td>232 (88)</td>
<td>6 (4)</td>
</tr>
<tr>
<td>Relapse (n=128)</td>
<td>28 (20)</td>
<td>1-4 (1-9)</td>
<td>9-1 (8-0)</td>
<td>72 (39)</td>
<td>282 (90)</td>
<td>8 (5)</td>
</tr>
<tr>
<td>p Value (ANOVA)</td>
<td>0-0001</td>
<td>0-0001</td>
<td>0-05</td>
<td>NS</td>
<td>0-0001</td>
<td>0-0001</td>
</tr>
</tbody>
</table>

Data shown as mean (SD). ESR (mm/h) = erythrocyte sedimentation rate, PLT (10^9/mm³) = platelet count, WBCC (10^3/mm³) = white blood cell count, PMN = polymorphonuclears, CRP (mg%) = C reactive protein, LAAT (%) = leucocyte adherence/aggregation test, ANOVA = one way analysis of variance.

TABLE II  Laboratory values obtained in patients with CD

<table>
<thead>
<tr>
<th></th>
<th>ESR (mm/h)</th>
<th>CRP (mg%)</th>
<th>WBCC (10^3/mm³)</th>
<th>PMN (%)</th>
<th>PLT (10^9/mm³)</th>
<th>LAAT (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls (n=500)</td>
<td>16 (15)</td>
<td>0-5 (0-3)</td>
<td>6-7 (0-8)</td>
<td>57 (15)</td>
<td>232 (88)</td>
<td>6 (4)</td>
</tr>
<tr>
<td>Remission (n=73)</td>
<td>32 (21)</td>
<td>1-8 (2-1)</td>
<td>10-5 (10-4)</td>
<td>79 (51)</td>
<td>299 (83)</td>
<td>8 (5)</td>
</tr>
<tr>
<td>Relapse (n=79)</td>
<td>45 (27)</td>
<td>4-4 (4-1)</td>
<td>12-2 (8-0)</td>
<td>73 (32)</td>
<td>376 (143)</td>
<td>18 (10)</td>
</tr>
<tr>
<td>p Value (ANOVA)</td>
<td>0-001</td>
<td>0-0001</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>0-0001</td>
</tr>
</tbody>
</table>

Data shown as mean (SD). Abbreviations as in Table I.

Laboratory methods

The erythrocyte sedimentation rate was determined according to the method of Westergren, the platelet, white blood cell count, and differential count were performed using a Technicon H1 autoanalyzer, while the quantitative C reactive protein assay was performed in a turbidimeter using a commercial kit. The LAAT was performed using a direct slide test. In brief, blood is drawn into a syringe that contains sodium citrate (one volume of 3-8% citrate and three volumes of whole blood). Several large drops of blood are placed on a slide that is held for two to three seconds at an angle of 45 degrees, so that the blood can slip down by gravity, leaving a fine film of blood. The slides are then dried while in a horizontal position in an incubator or at room temperature. The dried slides are placed at −18°C for 10 minutes to start haemolysis. Fixation is done with absolute methanol and staining with haematoxylin. For microscopic examination a drop of immersion oil is placed on the stained slide and covered with a cover glass. The ×1000 ocular is not used for the examination. Because of the comparative thickness of the blood film not all the leucocytes are present on the same level and the focus has to be adjusted often during the examination. The percentage of aggregated leucocytes on the slides is determined by counting 300 white blood cells at random (for example, if 30 of 300 are in aggregates, whether one aggregate of 30 cells or two aggregates of 15, the percentage of aggregated leucocytes is 10%). Cells are considered aggregated (Figure) when three or more nuclei are seen less than one cell diameter apart. From each patient, two slides are prepared, and the final result is a simple mean of both. Variations between duplicate samples does not generally exceed 20%, although from time to time larger differences are seen because of non-homogenous distribution of the aggregates.

This is also the variation noted when two independent observers examine different slides from the same patient. When the same slide is tested and retested, differences do not exceed 15%. The intra-assay coefficient of variation was calculated from 15 slides, which were simultaneously prepared from the same donor, and the interassay coefficient of variation was calculated when the same normal donor was examined on successive days for 15 consecutive days, the respective values being 0.2 and 0.3. The composition of the aggregates is generally similar to the differential white blood cell count - that is, it is formed mainly of neutrophils in cases of neutrophilia and of lymphocytes in cases of lymphocytosis. Mixed aggregates of polymorphonuclear and mononuclear leucocytes are often seen. If incubation is used for drying, results of the test can be given within 30 minutes. The statistical analysis was performed by using the SPSS package.

Results

Table I shows the results of all the laboratory tests in patients with IBD in remission and relapse, as well as the controls. All variables (erythrocyte sedimentation rate, C reactive protein, white blood cell count, platelet count, LAAT) reached statistical significance after one way analysis of variance except for the per cent of polymorphonuclears; showing that the mean values were different at least between two of the three groups (controls, remission, and relapse). Moreover, the unpaired t test with the Bonferroni correction showed significant differences between all the three inner differential groups (controls, remission, relapse).

In CD patients (Table II) statistically significant differences between each of the three groups (controls, remission, relapse) were found only for LAAT, C reactive protein, and erythrocyte sedimentation rate in that order. In UC patients (Table III) only the LAAT and erythrocyte sedimentation rate were significantly different between the three groups.

Analysis of the three subgroups of patients with IBD in relapse: with mild, moderate or severe disease (Table IV) showed that only the LAAT and the erythrocyte sedimentation rate showed statistically significant differences. The unpaired t test with the Bonferroni correction showed that the LAAT was significantly differ-
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### TABLE IV Laboratory values obtained in patients with active IBD

<table>
<thead>
<tr>
<th></th>
<th>ESR</th>
<th>CRP</th>
<th>WBCC</th>
<th>%PMN</th>
<th>PLT</th>
<th>LAAT (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mild (n=55)</td>
<td>34 (21)</td>
<td>2.8 (3.4)</td>
<td>10.1 (3.7)</td>
<td>69 (10)</td>
<td>334 (129)</td>
<td>13 (6)</td>
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<tr>
<td>Moderate (n=44)</td>
<td>48 (28)</td>
<td>3.5 (3.4)</td>
<td>11.9 (4.9)</td>
<td>73 (38)</td>
<td>390 (133)</td>
<td>17 (10)</td>
</tr>
<tr>
<td>Severe (n=29)</td>
<td>55 (30)</td>
<td>3.9 (4.7)</td>
<td>11.0 (4.4)</td>
<td>70 (11)</td>
<td>373 (157)</td>
<td>26 (10)</td>
</tr>
<tr>
<td>p Value (ANOVA)</td>
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<td>NS</td>
<td>NS</td>
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</table>

Data shown as mean (SD). Abbreviations as in Table I.

### TABLE V Laboratory values obtained in patients with active CD

<table>
<thead>
<tr>
<th></th>
<th>ESR</th>
<th>CRP</th>
<th>WBCC</th>
<th>%PMN</th>
<th>PLT</th>
<th>LAAT (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mild (n=52)</td>
<td>34 (21)</td>
<td>4.0 (3.8)</td>
<td>10.9 (4.3)</td>
<td>72 (11)</td>
<td>348 (136)</td>
<td>14 (7)</td>
</tr>
<tr>
<td>Moderate (n=30)</td>
<td>52 (27)</td>
<td>4.2 (3.8)</td>
<td>13.4 (10.9)</td>
<td>76 (46)</td>
<td>384 (134)</td>
<td>18 (10)</td>
</tr>
<tr>
<td>Severe (n=17)</td>
<td>54 (29)</td>
<td>5.1 (5.3)</td>
<td>11.0 (4.7)</td>
<td>70 (10)</td>
<td>311 (166)</td>
<td>26 (10)</td>
</tr>
<tr>
<td>p Value (ANOVA)</td>
<td>0.001</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
<td>0.0001</td>
</tr>
</tbody>
</table>

Data shown as mean (SD). Abbreviations as in Table I.

### Discussion

Inflammation confers upon the white blood cells increased adhesive properties. This adhesiveness has a known biological role regarding leucocyte – endothelial cell interactions. It has been repeatedly shown in the past that the increased adhesiveness of these cells can be used for diagnostic purposes including patients with IBD. In this study we have shown, in a comparatively large number of patients and controls that the LAAT is an excellent morphological parameter for determining disease activity and quantifying inflammation. In this regard it is clearly superior to other commonly used acute phase reactants including the erythrocyte sedimentation rate, the white blood cell count, the platelet count, the neutrophil count as well as quantitative C reactive protein measurements. It can be seen that by using the LAAT, that not only can the presence or absence of inflammation in patients with IBD be assessed but also the disease activity; a finding that might have clinical implications for both diagnostic and monitoring purposes.

One of the problems in IBD is determining the intensity of the disease, namely the inflammatory activity. Various clinical and laboratory tests have been used to solve this problem with only partial success. The problem whether a physician intends to measure disease activity or quantify inflammation, has been one of the problems in the use of clinical indices. Singleton, in his attempts to define a disease activity measurement, used inflammatory parameters and biochemical marker of disease activity in IBD should reflect the disease extent and the severity of inflammation. A substantial number of acute phase reactants and other parameters of inflammation are changed in relation to active disease, however, the types of biochemical parameters that should preferentially be used has not been established.

Using the consensus of all variables measured as a gold standard, the platelet count, followed by the erythrocyte sedimentation rate and C reactive protein correlated best with the final consensus in CD. Orosomucoid has been shown by others, to correlate best with clinical severity. Nevertheless we still lack a truly reliable laboratory indicator of the activity of CD. In this study the LAAT was the most useful test. It is possible that the leucocytes ‘sense’ the inflammatory mediators during their sojourn in the mesenteric capillaries of the inflamed intestinal segments and then present their stickiness in the peripheral circulation as shown in the adhesiveness test. It is tempting...
to assume that a correlation exists between the degree of intestinal inflammation and degree of adhesiveness of the cells. This study, which clearly shows a correlation between disease activity and degree of leucocyte adhesiveness, strongly supports this notion. In fact, we have already shown in the past that the LAAT correlates with inflammatory disease activity and that this adhesiveness is correlated to tissue leucostasis. 24-30 A stasis like this could contribute to unfavourable local rheology. The role of this increased LAA could be important in explaining the etiology of the various atherosclerotic or cerebrovascular disease. 8

We conclude that the LAAT is superior to other acute phase reactants currently used in daily practice to detect the presence of inflammation as well as for the assessment of its severity. In addition, our findings might have biological relevance to the disease process in patients with IBD and its potential manipulation by anti-adhesive agents.

This study was partially supported by The Shmuel Shalit Foundation, Tel Aviv University. Part of this material is included in the MD thesis of Gd Botan, Tel-Aviv University.


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N Arber, S Berliner, A Hallak, Y Bujanover, I Dotan, E Liberman, M Santo, M Moshkowitz, J Ratan and G Dotan

Gut 1995 37: 77-80
doi: 10.1136/gut.37.1.77

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