Treatment of ulcerative colitis in the cottontop tamarin using antibody to tumour necrosis factor alpha

P E Watkins, B F Warren, S Stephens, P Ward, R Foulkes

Abstract

Background—The aetiology and pathophysiology of ulcerative colitis remains unclear; however, there is increasing recognition of the critical role of inflammatory cytokines in the pathogenesis of this disease. Among these, tumour necrosis factor alpha (TNFα) seems to play an important role.

Aim—To study the effects of an engineered human monoclonal antibody to TNFα (CDP571) in the treatment of idiopathic ulcerative colitis in the cottontop tamarin.

Methods—Six cottontop tamarins with confirmed ulcerative colitis received repeated doses of CDP571. Progression of disease was assessed by measuring both body weight and rectal biopsy pathology.

Results—All animals showed a rapid improvement in clinical condition and rectal biopsy pathology that was maintained following completion of the therapy.

Conclusion—These studies indicate the efficacy of selective antibody therapy to TNFα for the treatment of ulcerative colitis in a primate and suggest that similar therapy in humans could be of value.

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Keywords: ulcerative colitis, cottontop tamarin, antibody to tumour necrosis factor α.

The term inflammatory bowel disease (IBD) encompasses two chronic disorders affecting all or part of the gastrointestinal tract. These are Crohn’s disease and ulcerative colitis. For both diseases the causative agent is unclear. Proposed aetiologies include infectious agents, response to measles vaccination, vascular disruptions, and disorders of the immune system. Furthermore, there is increasing evidence of a genetic susceptibility to these diseases, which may act alone or perhaps in association with environmental factors.

Ulcerative colitis and Crohn’s disease both demonstrate characteristic clinicopathological features. A central feature is the local inflammatory response in the gut wall which is characterised by an influx of T-lymphocytes and other mononuclear cells, especially macrophages. In recent years, numerous cytokines have been implicated as being crucial to the disease processes, including tumour necrosis factor alpha (TNFα), interleukin (IL) 6, IL-1β, platelet activating factor (PAF), and the significance of individual cytokines has often been difficult to elucidate from clinical studies. At times, investigators have failed to show a consistent alteration in cytokine production or release in IBD. Furthermore, in patients receiving treatment, certain treatments – for example, 5-aminosalicylic acid or corticosteroids, or both, will cause a reduction in the concentrations of numerous cytokines in the colonic mucosa. More recently, the development of specific antibodies to cytokines such as TNFα has provided the opportunity to assess the possible roles of individual cytokines in the disease state both in animal models and in humans.

TNFα is a cytokine released by activated mononuclear cells and T cells. It seems to have a clinically important role in septic shock and in rheumatoid arthritis. In the latter condition selective blockade of TNFα by monoclonal antibodies will ameliorate the disease process. In addition, TNFα has been implicated in the pathogenesis of IBD. Raised concentrations of TNFα have been noted in the serum and increased TNFα immunoreactivity has been shown in the lamina propria of patients with active Crohn’s disease or ulcerative colitis. Furthermore, it has been shown that there are significant increases in faecal TNFα concentrations in patients with active IBD.

A large number of animal models of IBD have been described and many have been reviewed previously. In most cases the models are solely of colonic inflammation, not of IBD. Inflammation is often induced by the local application of an irritant, such as acetic acid, to the colon, in some cases with prior sensitisation of the animal to the irritant material. Although these approaches can be used to study various aspects of colonic inflammation, the resulting pathology shows few, if any, similarities to that seen in human IBD. Moreover, the pathological changes invariably improve once the insult has been removed, unlike the situation in humans. None the less, in a rodent model of colonic inflammation a beneficial response was noted after administration of a selective monoclonal antibody directed against TNFα.

The cottontop tamarin, a small, new world primate, is unique among animal models of IBD in that it develops a spontaneous form of colitis which shows many similarities to the condition of ulcerative colitis in humans. Animals present clinically with chronic diarrhoea and weight loss and may die from the...
condition if untreated. Both the pathology and the response to therapy with 5-aminosalicylic acid compounds, in some cases combined with corticosteroids, show close similarities to the condition in humans. 18, 19 A unique feature of the disease in the cottontop tamarin is the development of secondary complications, namely colonic adenocarcinoma 20 and sclerosing cholangitis, 21 which are also seen in human ulcerative colitis. Also, we have recently demonstrated a significant increase in faecal TNFα concentrations in cottontop tamarins with active colitis, 22 illustrating a further similarity to the condition in humans.

The cottontop tamarin has been used before to evaluate potential therapeutic regimens for treating ulcerative colitis. These trials have included use of murine monoclonal antibodies against the adhesion molecules E-selectin and α4β1 integrin. 23

In this study the effect of an engineered monoclonal antibody against human TNFα (CDP571) was assessed in spontaneous ulcerative colitis in the cottontop tamarin. The study was divided into two sections: first, the pharmacokinetics of CDP571 were evaluated in two normal animals; and, second, a trial using CDP571 for treatment of ulcerative colitis in the cottontop tamarin was undertaken. This study was planned and executed in a similar way to an “open” clinical trial. This was because of the comparative scarcity of the animals (which are listed as endangered under the Conference on International Trade of Endangered Species (CITES)). No animals received placebo as an historical evaluation of colony health records showed that animals with confirmed ulcerative colitis which did not receive effective therapy continued to lose weight and showed further deterioration in their rectal biopsy pathology.

Methods

ANIMALS

All animals used in this study were captive bred at the University of Bristol colony. They were housed under standard environmental conditions as described previously. 24, 25 Animals were fed a complete pelleted diet (New World Primate diet, SDS Diets) and this was supplemented with fresh fruit each day. In addition, animals received regular supplementation with vitamin D3 administered orally.

Two animals were recruited to the pharmacokinetic study. Both appeared clinically normal and they were confirmed to be free of rectal pathology by histological examination of a rectal biopsy specimen taken before recruitment. Six cottontop tamarins were recruited to the second phase of the study (Table I). All animals had confirmed ulcerative colitis, based on their clinical history of diarrhoea and weight loss, endoscopic examination of the colon along with histopathological examination of a rectal biopsy specimen taken at that time. Faecal samples were taken from the animals for culture to eliminate any known faecal pathogens. None of the six animals recruited to the study had received previous treatment for colitis.

MONOCLONAL ANTIBODY

Generation of the antibody

The antibody used in this study (CDP571) is derived from a murine monoclonal antibody to recombinant human TNFα which has been engineered to contain human g4 and κ light chain constant regions with Eu frameworks as described previously. 26

Pharmacokinetics

Two animals received a single dose of CDP571 at 20 mg/kg by intramuscular injection. To administer the antibody, animals were sedated by an intramuscular injection of ketamine hydrochloride (Vetalar, Parke Davies; 20–25 mg/kg). Blood samples (maximum volume 0.5 ml into EDTA tubes) were taken from these animals before injection and at eight and 24 hours, and two, three, five, seven, 10, 14, 21, and 28 days after administration of CDP571. Samples were centrifuged and plasma stored at −70°C pending analysis for CDP571 and antibodies to CDP571.

Therapeutic protocol

Animals recruited to the study received CDP571 at a dose of 20 mg/kg every six days for a total of six doses. CDP571 was injected, by deep intramuscular injection, into the quadriceps femoris muscle under anaesthesia as described earlier (Table II).

EVALUATION OF RESPONSE TO TREATMENT

Standard indicators of disease progression in the cottontop tamarin were used to evaluate response to treatment. These were clinical signs, body weight and histopathological examination of rectal biopsy specimens taken on

<table>
<thead>
<tr>
<th>TABLE I</th>
<th>Details of animals recruited to second part of the study</th>
</tr>
</thead>
<tbody>
<tr>
<td>Animal</td>
<td>Sex</td>
</tr>
<tr>
<td>B221</td>
<td>Male</td>
</tr>
<tr>
<td>B213</td>
<td>Male</td>
</tr>
<tr>
<td>B201</td>
<td>Male</td>
</tr>
<tr>
<td>B111</td>
<td>Male</td>
</tr>
<tr>
<td>R194</td>
<td>Female</td>
</tr>
<tr>
<td>R192</td>
<td>Female</td>
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</tbody>
</table>

<table>
<thead>
<tr>
<th>TABLE II</th>
<th>Outline of protocol</th>
</tr>
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<tbody>
<tr>
<td>Day</td>
<td>CDP571</td>
</tr>
<tr>
<td>0</td>
<td>0 dose</td>
</tr>
<tr>
<td>12</td>
<td>12 dose</td>
</tr>
<tr>
<td>24</td>
<td>24 dose</td>
</tr>
<tr>
<td>25</td>
<td>25 dose</td>
</tr>
<tr>
<td>30</td>
<td>30 dose</td>
</tr>
<tr>
<td>49</td>
<td>49 dose</td>
</tr>
<tr>
<td>63</td>
<td>63 dose</td>
</tr>
</tbody>
</table>
days 12, 32, 49, and 63. Biopsy specimens were evaluated by a pathologist using an objective scoring system developed from one used previously for assessing human rectal specimens (Table III). Specimens were graded from 0 (normal) to 6 (severe active disease) on the basis of inflammatory cell infiltrate, crypt architecture and mucosal disruption (Fig 1).

Biopsy scores and body weights before and after CDP571 treatment were compared using the Friedman non-parametric repeated measures test. Differences were considered significant when p<0.05.

Blood samples were also taken when the animals were sedated for other procedures – that is, on days 12, 25, 32, 49, and 63, to permit measurement of plasma concentrations of CDP571 and of host antibodies to CDP571.

ELISAS for CDP571 and Antibodies to CDP571
ELISA techniques were used as described previously. Briefly, for the pharmacokinetic assay, diluted plasma samples were added to microtitre plates coated with rhTNFα and bound CDP571 was revealed with mouse anti-human IgG4 (Serotec) followed by horseradish peroxidase (HRP) conjugated goat anti-rabbit IgG (Jacksons Laboratories). Bound HRP was revealed using TMB substrate and colour development as proportional to the amount of CDP571 in the sample.

Antibodies to CDP571 were detected using a double-antigen sandwich ELISA system. Plasma samples (diluted 1 in 10 in phosphate buffered saline)/1% bovine serum albumin (BSA) were added to CDP571 coated plates and the sandwich was completed with CDP571 conjugated to HRP, followed by TMB substrate. An affinity purified rabbit hyperimmune serum was used as a standard and results expressed as units/ml (1 unit being equivalent to 1 mg/ml of the rabbit standard).

Results

PHARMACOKINETIC STUDY
Figure 2 shows the plasma elimination profiles for the two animals. Analyses demonstrated that CDP571 was cleared from normal tamarins with a half-life of around six days (5.8 and 7.0 days). By day 28, there was little host antibody response to CDP571. Therefore, the drug was administered every six days in the therapeutic protocol.

RESPONSE TO TREATMENT
None of the animals showed any adverse effects to repeated dosing with CDP571, either locally or systemically.

Analysis of plasma samples confirmed that during the treatment period, CDP571 concentrations continued to reach circulating levels in excess of 100 μg/ml (range 51.4–227.8 μg/ml; Table IV) and remained in circulation for several weeks (geometric mean of 0.5 μg/ml and range of 0.05–6.5 μg/ml at day 49). No immune response to the antibody was detected during the treatment period, although low levels of antibodies to CDP571 (3–9 units/ml) were seen in four of the six animals when CDP571 was cleared from the circulation after the final dose.

All animals showed a clinical response to treatment and in all cases there was an improvement in faecal quality after starting treatment. Figure 3 shows changes in body weight. Mean body weight rose following the first dose of antibody and remained significantly elevated during the treatment period and during the following month (+33.3 (10) g (mean (SEM)) at day 63, p<0.05). This represents an increase of around 6% compared with pre-entry body weight and was maintained throughout the study.

Examination of rectal biopsy specimens showed a rapid fall in mean rectal biopsy score after starting treatment (Table V), indicating a
value of 4.5 (0-2) to 3 (0-4) (p<0.05). This improvement was maintained until day 49.

**Discussion**

Recent advances in the understanding of the mechanisms and control of inflammation have focused on the importance of numerous cytokines in this process. TNFα seems to play an important role in septic shock, rheumatoid arthritis and IBD. This has been demonstrated by the raised concentrations of TNFα both in plasma and locally in inflamed tissues in these conditions and has led to the development of monoclonal antibodies as potential therapies in these conditions. A murine antibody to TNFα has been used in early phase trials in patients with septic shock but was associated with a significant host response to the antibody. However, antibody engineering involving either chimerisation or CDR grafting of murine monoclonal antibodies to TNFα to reduce immunogenicity has permitted certain conditions – for example, rheumatoid arthritis, to be treated. From this background, we evaluated the use of an antibody to TNFα in the treatment of ulcerative colitis in an animal model.

Previous work using chemically induced colonic inflammation in rats has clearly demonstrated the ability of anti-murine TNFα to modulate the inflammatory response; however, the initial dose was given prior to the induction of inflammation. In ulcerative colitis in humans, treatment is required once disease is diagnosed and often the disease process may be longstanding before treatment is instituted.

Spontaneous ulcerative colitis in the cottontop tamarin provides a good model for evaluating new treatments for established disease in circumstances more related to the clinical situation. All animals recruited to this study had not received any previous treatment and the anti-TNFα was the sole treatment used. Based on the knowledge of pharmacokinetics of CDP571 and the lack of a deleterious immune response, we administered the antibody safely on a total of six occasions over six weeks. By comparison, Podolsky et al. using a murine monoclonal antibody to α4 integrin to treat colitis in the cottontop tamarin, was restricted to therapy at two day intervals over 10 days because of the host immune response to the antibody. In this study, all animals demonstrated a rapid improvement after administration of anti-TNFα with an increase in body weight, an improvement in the consistency of stools and in rectal

![Figure 2: Plasma clearance profile of CDP571 in two cottontop tamarins after a single intramuscular injection of 20 mg/kg. Values are individual plasma concentrations.](image)

![Figure 3: Changes in body weight for individual animals and the mean (SEM) (solid circles) value in cottontop tamarins given a course of CDP571 treatment (6×20 mg/kg).](image)

**Table IV** Plasma concentrations of CDP571 (μg/ml)

<table>
<thead>
<tr>
<th>Time after first dose (days)</th>
<th>Dose no./days after administration of CDP571</th>
<th>Animal number</th>
<th>Geometric mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>-3</td>
<td>Before</td>
<td>R192</td>
<td>0.05</td>
</tr>
<tr>
<td>12</td>
<td>2/6</td>
<td>R194</td>
<td>0.05</td>
</tr>
<tr>
<td>25</td>
<td>5/1</td>
<td>B111</td>
<td>0.05</td>
</tr>
<tr>
<td>32</td>
<td>6/2</td>
<td>B201</td>
<td>0.05</td>
</tr>
<tr>
<td>49</td>
<td>6/19</td>
<td>B213</td>
<td>0.05</td>
</tr>
<tr>
<td>63</td>
<td>6/33</td>
<td>B221</td>
<td>0.05</td>
</tr>
</tbody>
</table>

*Limit of detection 0.05 μg/ml.
NT=not tested.

**Table V** Colonic biopsy scores

<table>
<thead>
<tr>
<th>Time after first dose (days)</th>
<th>Biopsy score (mean (SEM))</th>
</tr>
</thead>
<tbody>
<tr>
<td>-3</td>
<td>4.5 (0-2)</td>
</tr>
<tr>
<td>12</td>
<td>2.7 (0-8)*</td>
</tr>
<tr>
<td>32</td>
<td>3.0 (0-8)*</td>
</tr>
<tr>
<td>49</td>
<td>2.3 (0-7)*</td>
</tr>
<tr>
<td>63</td>
<td>3.6 (0-2)</td>
</tr>
</tbody>
</table>

Values are mean (SEM) for six animals.
* p<0.05 compared with pretreatment value (Friedman test).
biopsy score. The response, which occurred as early as the first injection, was far more rapid than that seen to an established therapeutic regime based on the 5-aminosalicylic acid compound olsalazine. 5-Aminosalicylic acid based treatment is standard for ulcerative colitis in humans and has been used successfully to treat ulcerative colitis in the tamarin. However, when starting treatment there must be a gradual increase in dose levels. Improvement in rectal biopsy pathology and in clinical condition may not be seen for eight to 10 weeks in tamarins.

There were no placebo treated control animals in this study. The cottontop tamarin is listed under CITES as an endangered species and as such it was felt unethical to withhold therapy from animals with ulcerative colitis. Furthermore, it is known from previous studies in the colony that if animals are not treated the disease progresses unabated with progressive weight loss and deterioration of clinical condition leading either to death or euthanasia. Reference to colony records for six animals with confirmed ulcerative colitis that were not receiving therapy were monitored over a four week period and revealed an increase in mean rectal biopsy score from 1.3 to 2.6 over this period, indicating a deterioration in pathology.

Of the six animals that were recruited to the study, the majority (4/6) have not required any further treatment 18 months after the last dose. Two animals did, however, relapse and were given maintenance therapy with olsalazine. This occurred between eight months (R192) and one year (B213) after ceasing therapy. Prolonged benefit after a single course of therapy provides further encouragement for the use of anti-TNFα in the treatment of ulcerative colitis. The exact mechanism of action of anti-TNFα is uncertain in these animals, but it seems most likely that it inhibits the action of TNFα in the colon wall and so attenuates the inflammatory process.

Repeated therapy with CDP571 has been used in a small number of tamarins with ulcerative colitis which was refractory to standard therapy of olsalazine and prednisolone. Again, no adverse effects were recorded and although the animals showed some improvement in their condition, the clinical response was not as notable as that reported in this study of native animals (unpublished observations). Human patients with rheumatoid arthritis have received repeated doses of chimeric anti-TNFα (cA2) and although apparently safe it was noted that the interval between doses fell as the number of doses administered rose. By comparison, CDP571, at a dose of 10 mg/kg, has been given at 2 to 3 month intervals for a total of four doses in the treatment of rheumatoid arthritis. No adverse effects were noted and pharmacokinetic studies indicated no reduction in the half-life in patients. These data provide encouragement for repeated dosing in human.

The potential role of anti-TNFα in the treatment of human ulcerative colitis is now under investigation. Engineering of antibodies can prolong the half-life in humans from a few hours to one to two weeks. Encouraging results have already been obtained with chimeric antibodies to TNFα in the treatment of Crohn’s disease. Studies with CDP571 are now continuing in acute and chronic inflammatory conditions such as IBD, and indeed preliminary studies with a single dose of antibody in an open study have proved encouraging.

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