Elevated calcium and activation of trypsinogen in rat pancreatic acini

T W Frick, C Fernández-del-Castillo, D Bimmler, A L Warshaw

Abstract

Background—Acute pancreatitis associated with hypercalcaemia has been described in humans and experimental animals. It has been demonstrated that calcium dose dependently accelerates trypsinogen activation, and it is generally believed that ectopic activation of digestive enzymes is an early event in the pathophysiology of acute pancreatitis.

Aims and methods—Trypsinogen activation peptide (TAP) was measured in isolated rat pancreatic acini exposed to elevated extracellular calcium in order to investigate the association between calcium and trypsinogen activation in living cells. TAP was determined in the culture medium either before (extracellular compartment) or after (intracellular compartment) cell homogenisation.

Results—Neither secretory stimulation nor elevated calcium alone caused an increase in TAP levels. Maximal cerulein or carbachol stimulation superimposed on high medium calcium, however, significantly increased intracellular trypsinogen activation twofold. This increase was inhibited by either N\(^-\)monomethyl-L-arginine (L-NMMA) or verapamil. Acinar cell morphology and function remained intact as demonstrated by electron microscopy and secretagogue dose-response studies.

Conclusions—These results support the hypothesis that increased intracellular trypsinogen activation is an early step in the pathogenesis of hypercalcaemia induced pancreatitis. The model may therefore bear on other types of pancreatitis as elevated cytosolic calcium is thought to be an early event in the pathogenesis of acute pancreatitis in general.

Keywords: hypercalcaemia; pancreatitis pathogenesis; serine proteases; acute pancreatitis

There is increasing evidence that elevated calcium in pancreatic acinar cells is an important early step in the development of acute pancreatitis.\(^2\) We previously used hypercalcaemia as a model to study the effects of elevated calcium in the development of pancreatitis in vivo. In humans, acute pancreatitis was found to be associated with hypercalcaemic conditions, such as hyperparathyroidism or therapeutic calcium administration.\(^2\)\(^4\) In a rat model acute pancreatitis was induced by bolus injection of calcium.\(^3\) Other experimental protocols used low dose continuous infusions of calcium leading to a twofold increase in serum ionised calcium. Morphological changes of early acute pancreatitis were seen in several animal species.\(^6\)\(^7\) It was shown that hypercalcaemia induced a secretory block and accumulation of digestivezymogenswithinthePancreaticacinacell.\(^8\) Zymogen activation, in particular trypsinogen, in homogenates of pancreatic tissue after calcium injection, suggested that the combination of zymogen accumulation and increased calcium leads to increased intrapancreatic trypsinogen activation as a very early step in the pathogenesis of acute hypercalcaemia induced pancreatitis.\(^1\)\(^8\)

Despite this evidence it remained unclear whether the ectopic zymogen activation occurred as an initial step in the pathogenesis of pancreatitis, or whether it was the result of acinar cell injury. In the work reported in the present paper, the effect of elevated environmental calcium on trypsinogen activation was investigated more directly. We used an in vitro model of isolated pancreatic acini exposed to elevated medium calcium. As a marker for trypsinogen activation we measured trypsinogen activation peptide (TAP), the N-terminal of trypsinogen which is cleaved to active trypsin.\(^10\) The five amino acid carboxyl end of TAP is highly preserved among species, and the antibody against TAP used in the competitive ELISA is highly specific. Quantification of TAP is a direct measurement of the amount of activated trypsinogen as one TAP molecule is generated for each molecule of trypsinogen cleaved to trypsin.

Materials and Methods

ACINAR CELL SUSPENSIONS

For each experiment three male Wistar rats (80–100 g) fasted overnight were used. They were sacrificed in CO\(_2\), the pancreas was quickly excised in the cold room, and pancreatic acini were prepared by collagenase digestion.\(^11\) Collagenase (type CLS 4, 1000 U/ml) was purchased from Worthington Biochemical Corporation, Freehold, New Jersey, USA. After digestion the cells were incubated at 37°C in medium containing HEPES 12.5 mM, NaHCO\(_3\) 5.0 mM, NaCl 125 mM, KCl 5.0 mM, KH\(_2\)PO\(_4\) 1.2 mM, MgSO\(_4\) 1.2 mM, D-glucose 5.0 mM, aprotinin 0.01 mg/ml, soybean trypsin inhibitor 0.1 mg/ml, BSA 0.1%, pH adjusted to 7.40 with NaOH, and either CaCl\(_2\) 1.2 mM (physiological concentration) with additional NaCl 3.8 mM, or CaCl\(_2\) 5.0 mM (hypercalcaemia)
(all chemicals were from Sigma Chemical Company, St Louis, Missouri, USA, unless otherwise specified). We tested a calcium concentration of 5.0 mM because previous studies on pancreatic lobules indicated that this concentration maximally enhanced acinar secretion without tissue damage. Cell viability was tested with the trypan blue exclusion method immediately after preparation of acini by collagenase digestion. Preparations were accepted for study only if more than 95% of the cells excluded the dye.

**STUDIES OF AMYLASE SECRETION AND TRYPSINOGEN ACTIVATION**

The cells were incubated at 37°C in 24-well cell culture plates (Falcon 3047, Becton Dickinson Labware, Lincoln Park, New Jersey, USA) with 500 µl of cell suspension in each well, and gassed with 100% O₂. After 15 minutes of preincubation, cerulein, carbachol, N⁶-monomethyl-L-arginine (L-NMMA), and verapamil were added. All measurements were performed in duplicate and controls were run on the same plates. At the end of the one hour incubation period 100 µl of medium was removed, 400 µl of Tris (0.1 M, pH 8.9) and Triton X-100 (2%) was added for cell homogenisation, and the plate was incubated on a mini shaker for 30 minutes at room temperature.
**STUDIES OF TRYPSINOGEN ACTIVATION KINETICS**

To examine trypsinogen autoactivation 3 mg/ml bovine trypsinogen was dissolved in incubation medium, pH adjusted to 7.40, with either no calcium or 1.2 mM or 10.0 mM CaCl₂. The preparation was incubated at 37°C in 24-well cell culture plates (1000 µl per plate) and 10 µl aliquots were removed at 15 minute intervals. The aliquots were boiled immediately in 1000 µl medium containing 0.02 M EDTA before determination of TAP.

**DETERMINATION OF AMYLASE AND TRYPSINOGEN ACTIVATION PEPTIDE**

Amylase was measured in the medium and the homogenate with the Phadebas amylase test (Pharmacia Diagnostics AB, Uppsala, Sweden).

To determine free TAP a competitive ELISA immunoassay was used as described elsewhere with the following modifications: the antiTAP antiserum (R7103/4/5) was diluted 1/400 in assay buffer, and the alkaline phosphatase-extravidin conjugate was used in a 1/750 dilution. To ascertain the relationship of the observed TAP levels with the total trypsinogen pool, the homogenates were incubated with enterokinase (1 U/ml) for one hour at 37°C to convert all trypsinogen to equimolar quantities of trypsin and TAP as described previously. The total TAP after this processing represents a measure of the trypsinogen content of the cells. TAP levels can then be expressed as a function of total trypsinogen to indicate the degree of activation.

**ELECTRON MICROSCOPY**

After the incubation period, acini were immediately fixed in glutaraldehyde (3%), formaldehyde (2%), and sodium cacodylate (0.1 M) for one hour at 4°C, washed twice in 0.1 M sodium cacodylate, and embedded in Epon after dehydration in graded series of alcohol. Thin sections were stained on the grid with uranyl acetate and lead citrate and viewed with a JEOL 100-CX electron microscope.

**STATISTICAL ANALYSIS**

Statistical calculations were performed with InStat software (Graphpad, San Diego, California, USA) using the two tailed Student’s t test for paired data for interpretation of the results. Data are presented as mean (SEM) unless indicated otherwise.

**Results**

**STUDIES OF IN VITRO TRYPSINOGEN ACTIVATION KINETICS**

The effect of calcium on spontaneous cleavage of trypsinogen was evaluated by measuring the appearance of TAP in trypsinogen solutions (3 mg/ml medium) containing graded calcium concentrations. Figure 1 shows that there was acceleration of trypsinogen activation at higher calcium concentrations.

**STUDIES OF CELLULAR TRYPSINOGEN ACTIVATION**

Extracellular TAP was less than 0.5% of total TAP at both concentrations of calcium in the medium, and no difference between the preparations was found (data not shown). Neither secretory stimulation nor incubation in high calcium (5.0 mM) alone caused an increase in intracellular TAP levels with respect to levels found in physiological calcium (1.2 mM). However, elevated calcium superimposed on either maximal or supramaximal cerulein stimulation significantly increased intracellular TAP (fig 2). Carbachol at maximal stimulatory concentrations similarly induced a significant increase in intracellular TAP concentration in the high calcium medium, but not at physiological calcium levels (fig 3). Verapamil and L-NMMA significantly inhibited the TAP increase by maximal carbachol and cerulein stimulation (fig 3).

**MORPHOLOGY**

Electron microscopy of acini after incubation in 5.0 mM calcium and 10⁻⁶M cerulein revealed powerful stimulation as indicated by enlarged Golgi apparatus, increased number of condensing vacuoles, and decreased number of zymogen granules (fig 4). Acinar cell ultrastructure remained intact with normal mitochondria, cell nucleus, and cell membrane.

**EFFECT OF ELEVATED CALCIUM ON AMYLASE SECRETION**

The effect of elevated extracellular calcium on stimulated amylase secretion was investigated with carbachol and cerulein. The dose-response curves with carbachol and cerulein showed a significant increase in amylase output at maximal stimulation in elevated medium.
minia is a calcium triggered intrapancreatic activation of trypsin: (1) calcium accelerates generation of trypsin from trypsinogen in vitro in a dose dependent fashion as documented by direct determination of trypsin activity and in this study by measuring TAP. (2) Elevated extracellular calcium increases cytosolic calcium in isolated pancreatic acini in proportion to the extracellular calcium level.

To test the hypothesis, we exposed isolated pancreatic acini with or without stimulation to elevated extracellular calcium. Using Fura-2 as a marker for spectrophotometric measurement of intracellular calcium, we confirmed that the cytosolic calcium increases in response to increased extracellular calcium concentrations (data not shown). After one hour TAP was determined in the medium before (extracellular compartment) and after (intracellular compartment) lysis of all acinar cells. In the extracellular compartment, the concentrations of TAP were below 1% of the total and there were no differences seen in preparations stimulated with cerulein or exposed to high calcium. This indicates that intracellular TAP is not secreted into the medium in substantial amounts. It also indicates that there is no significant activation of secreted trypsinogen in the extracellular compartment in this experimental setting.

The mean TAP concentration in the intracellular compartment of unstimulated cells exposed to physiological calcium concentrations was close to 5%. Leach et al had demonstrated a comparable amount of active intracellular proteases in unstimulated pancreatic acini, although no active trypsin was found. This may represent spontaneous intracellular protease activation. It may however indicate that intracellular trypsin is immediately neutralised by local protease inhibitors, thereby maintaining a stable state. The advantage of TAP determination is that it demonstrates trypsinogen activation irrespective of whether the resulting trypsin is active or blocked by inhibitors. However, because the intracellular metabolism of TAP is unknown, the measured TAP level may also be the result of TAP accumulation while trypsinogen actually activates at a lower rate.

Elevation of extracellular calcium alone did not generate an increase in intracellular TAP, but when the acinar cells were stimulated with cerulein a rise in intracellular TAP was noted in cells exposed to high environmental calcium and the degree of TAP production was directly related to the cerulein level. The dose-response curve was biphasic with a peak at maximal cerulein stimulation. The increase was significant compared with both maximally stimulated acinar cells in 1.2 mM calcium and unstimulated cells in 5.0 mM calcium.

Another significant rise in intracellular TAP was seen when cells exposed to high extracellular calcium were maximally stimulated with carbachol. As both maximal secretory stimulation and elevated extracellular calcium increase cytosolic calcium, it is possible that a
concordant effect caused the intracellular calcium concentration to rise beyond a critical level above which trypsinogen autoactivation was accelerated. It is however unclear why the increase in TAP was less pronounced during supramaximal stimulation, although equally high calcium spikes have been demonstrated.20 A possible explanation is that, although the calcium spikes are equally high, the duration of the calcium spikes is shorter and thus the absolute calcium concentrations may be lower.20

To test whether the increase in TAP is related to the increase in intracellular calcium, we used the calcium blockers l-NMMA and verapamil. The nitric oxide synthetase inhibitor l-NMMA has been shown to reduce significantly 45Ca2+ uptake in pancreatic acini stimulated by carbachol and is thought to decrease cytosolic calcium concentration.21 Verapamil blocks calcium uptake through voltage-gated calcium channels.22 Although a direct effect on cells without voltage gated calcium channels, such as pancreatic acinar cells,23 has not been demonstrated, verapamil has an inhibitory effect on agonist stimulated amylase release which is partially reversible by increasing the extracellular calcium concentration.22 24 In our studies, both compounds inhibited the increase in intracellular TAP induced by maximal cerulein or carbachol stimulation. In order to determine whether trypsinogen activation is the result of acinar cell injury or an event that precedes acinar cell damage, we looked at acinar cell ultrastructure and acinar cell function. Electron microscopy of maximally stimulated cells in high extracellular calcium revealed signs of intense stimulation, such as increased numbers of condensing vacuoles and enlarged Golgi apparatus, but normal chromatin structure, mitochondria, and endoplasmic reticulum, indicating intact acinar cell structure. Study of acinar cell function as determined by amylase secretion in response to cerulein or carbachol showed that the cells responded to high calcium by enhanced amylase output with an intact dose-response curve.25 This demonstrates undisturbed acinar cell response to stimuli and intact acinar function.

In this study, we have demonstrated increased intracellular TAP production in response to elevated extracellular calcium and maximal stimulation in morphologically and functionally intact rat pancreatic acini. The results support the hypothesis that increased intracellular trypsinogen activation may be an initial step in the development of hypercalcemia induced pancreatic tissue damage. The model may also have a bearing on other types of pancreatitis: it is becoming increasingly apparent that an increase in cytosolic calcium might be a key event in the pathogenesis of acute pancreatitis in general.26

We would like to acknowledge the great help and effort of Dr Max A Spycher in the preparation of the electron microscopy samples, and of Uma Mandavilli and Regula Gassmann for their enormous help in preparing the acinar cells and measuring TAP. The work was supported in part by a grant from the Schweizerische Stiftung für Medizinisch Biologische Stipendien, the Kommission zur Förderung des Akademischen Nachwuchses der Universität Zürich, and the Swiss National Foundation.

Elevated calcium and activation of trypsinogen in rat pancreatic acini

T W Frick, C Fernández-del-Castillo, D Bimmler and A L Warshaw

Gut 1997 41: 339-343
doi: 10.1136/gut.41.3.339

Updated information and services can be found at:
http://gut.bmj.com/content/41/3/339

References
This article cites 23 articles, 4 of which you can access for free at:
http://gut.bmj.com/content/41/3/339#BIBL

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Topic Collections
Articles on similar topics can be found in the following collections
Pancreas and biliary tract (1949)
Pancreatitis (531)

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/