Major virulence factors, VacA and CagA, are commonly positive in *Helicobacter pylori* isolates in Japan

S Maeda, K Ogura, H Yoshida, F Kanai, T Ikenoue, N Kato, Y Shiratori, M Omata

Abstract

**Background**—VacA and CagA proteins have been reported to be major virulence factors of *Helicobacter pylori*. However, antibodies against these proteins are frequently found in the sera of Japanese patients regardless of their gastroduodenal status.

**Aim**—To evaluate the expression of VacA and CagA proteins by *H pylori* strains isolated in Japan.

**Methods**—By using specific antibodies raised against recombinant VacA and CagA proteins, the expression of VacA and CagA was evaluated in 68 *H pylori* strains isolated from Japanese patients; a vacculating assay and genotyping of the vacA gene were also used in the evaluation. The results were analysed in relation to the gastroduodenal diseases of the hosts.

**Results**—VacA and CagA proteins were expressed in 59/68 (87%) and in 61/68 (90%) isolates respectively. The vacculating assay was positive in 57/68 (84%) isolates, indicating that most immunologically VacA positive strains produced active cytotoxin. The prevalence of infection with strains expressing CagA and positive for vacculating activity (Type I) was very high, 54/68 (79%), irrespective of the gastroduodenal status of the host.

**Conclusion**—Most *H pylori* isolates in Japan are positive for vacculating cytotoxin and CagA, and thus these virulence factors cannot be used as markers to discern the risk of developing serious gastroduodenal pathologies in the hosts. However, the high prevalence of infection with strains positive for vacculating cytotoxin and CagA may contribute to the characteristics of *H pylori* infection in Japan.

**Keywords:** VacA; *Helicobacter pylori*; CagA; ulcer; non-ulcer dyspepsia

**Helicobacter pylori** is a Gram negative microaerophilic bacterium that can infect human gastric mucosa. The organism is closely associated with not only active gastritis but also peptic ulcers and gastric cancer. The prevalence of *H pylori* infection is high worldwide, and about 80% of the general population are infected at age 40–50 in Japan. However, the infection leads to peptic ulcers or gastric cancer in only a small subset of infected patients. Although the pathogenesis of *H pylori* related gastroduodenal diseases is not well understood, there are several putative virulence factors of *H pylori* that may contribute to the exacerbation of mucosal damage.

Vacculating cytotoxin, which induces vacuolation in cultured eukaryotic cells, is secreted by about 50% of *H pylori* isolates in Western countries. Infection with vacculating cytotoxin positive strains is reported to be associated with particular gastroduodenal diseases. For instance, Figurea showed that strains with vacculating cytotoxin activity were found in 16 of 24 (67%) patients with peptic ulcers but in only 16 of 53 (30%) patients without. The cytotoxin is encoded by the vacA gene, and thus is also called VacA protein.

CagA protein is another putative virulence factor, encoded by the *cagA* gene (cytotoxin associated gene). It has been reported that this gene is present in about 60% of *H pylori* isolates, and nearly all *cagA* positive strains express the protein. Although the function of CagA protein is not known, the protein has been reported to be associated with particular gastroduodenal diseases. Ching et al. found serum anti-CagA antibody in 165 of 197 (84%) patients with duodenal ulcer, but in only 25 of 45 (56%) patients with non-ulcer dyspepsia. Recently, CagA positive *H pylori* strains have been shown to be associated with interleukin-8 (IL-8) induction in gastric epithelium. Neutrophilic infiltration into the gastric epithelium, which is characteristic of *H pylori* infection, may be due to increased production of IL-8. CagA protein may thus be related to gastric inflammation and gastroduodenal diseases.

Expression of CagA protein is closely associated with that of vacculating cytotoxin, although the underlying mechanism is not understood. Thus Xiang et al. classified *H pylori* strains into two groups, Type I and Type II. They and others reported that the Type I strains, which are positive for both vacculating cytotoxin and CagA, were strongly associated with peptic ulcer diseases in the host.

However, there have been few reports on the expression of VacA and CagA proteins in *H pylori* strains isolated in regions other than Western countries. In previous studies, we made recombinant CagA and VacA proteins and investigated serum anti-CagA and anti-VacA antibodies in Japanese patients with *H pylori* infection. These studies showed unexpectedly high seropositivity for both...
anti-VacA and anti-CagA antibodies irrespective of the gastroduodenal disease.

Since the presence of antibodies does not necessarily imply on-going infection, our previous studies suggested but did not prove high prevalence of infection with Type I strains in Japan. Therefore we conducted the current study to examine, by immunoblot analysis using specific antibodies raised against recombinant proteins, the expression of VacA and CagA proteins by *H pylori* isolates obtained from Japanese patients.

All *H pylori* isolates reported so far were found to possess the *vacA* gene, and strains without cytotoxin activity are thought to produce inactive cytotoxin and/or have defective secretion mechanisms. Thus we also examined the vacuolating activity and the genotypes of the *vacA* gene reported to be associated with the ability to secrete vacuolating cytotoxin.

**Materials and methods**

**Bacterial strains and growth**

*H pylori* strains were isolated from gastric biopsy samples of 68 patients (40 men and 28 women; age range 22–85 years; mean age 52.5) who underwent endoscopy in Tokyo University Hospital. The endoscopic findings were gastric ulcer in 16, duodenal ulcer in 17, non-ulcer dyspepsia in 25, and gastric cancer in 10 patients. Biopsy specimens were cultured on Columbia agar with 5% horse blood and Dent antibiotic supplement (Oxoid, Basingstoke, Hants, UK) at 37 °C for five days under microaerobic conditions. Isolates were identified as *H pylori* by Gram stain morphology and by positive urease, catalase, and oxidase activities. They were kept at −80 °C in brucella broth with 5% fetal bovine serum (FBS) containing 16% glycerol. ATCC43526 cells were used as VacA and CagA positive controls, and strain Tx30a was used as CagA negative control.

**Recombinant VacA and CagA proteins and antisera to recombinant VacA and CagA**

Recombinant VacA and CagA proteins and the rabbit antisera to each recombinant protein were prepared as described previously. In brief, the entire *vacA* gene (GenBank accession number AF001358) and *cagA* gene (GenBank accession number AF001357) of the ATCC43526 *H pylori* strain were cloned, and partial fragments of *vacA* (encoding amino acids 1, which is the ATG start codon, to 744) and *cagA* (encoding amino acids 107–531) genes were subcloned in frame with the sequence coding for the histidine tag of pQE vectors (Qiagen, Chatsworth, CA, USA). The histidine-tagged VacA and CagA proteins expressed in *Escherichia coli* were purified on Ni/nitrilotriacetic acid resin (Qiagen). Antibodies against the recombinant proteins were raised in New Zealand White rabbits.

**Detection of VacA and CagA proteins in *H pylori* isolates**

Immunoblot analysis was performed to detect VacA and CagA proteins produced by *H pylori* strains. VacA protein was examined using *H pylori* surface proteins prepared by the method of Xiang et al. In brief, the cultured bacterial cells were washed twice with distilled water, and, to separate the surface protein from bacterial cells, vortex mixed with distilled water for one hour at room temperature. After centrifugation at 10 000 g for ten minutes, the supernatants were stored at −30 °C until use.

CagA protein was examined using whole cell lysates. The whole cell lysates were washed twice in normal saline and centrifuged, and boiled at 100 °C for 5 min in Laemmli sample buffer before use. The surface proteins and the whole cell lysates were separated by electrophoresis in sodium dodecyl sulphate/10% polyacrylamide gel and transferred to a nitrocellulose membrane (Schleicher & Schuell, Dassel, Germany). The filter was blocked with buffer containing 5% skimmed milk in 25 mM Tris/HCl, pH 7.4, containing 150 mM NaCl and 5 mM KCl. The membrane was incubated with a 1:100 dilution of rabbit antisera against the recombinant VacA or CagA. After being washed, the membrane was incubated with 125I-labelled goat anti-rabbit immunoglobulin G, and then exposed to x ray film.

**Vacuolating assay**

*H pylori* strains were cultured at 37 °C in brucella broth containing 5% FBS under microaerobic conditions for 48 hours. The culture medium was centrifuged at 10 000 g for 20 min, and cell-free supernatant was sterilised by passage through a 0.2 μm pore size filter and
Table 1  Helicobacter pylori strains used in this study

<table>
<thead>
<tr>
<th>Strain* no.</th>
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<th>CagA</th>
<th>Vaculating assay</th>
<th>Type†</th>
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<tbody>
<tr>
<td>1 (T-1)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>I</td>
</tr>
<tr>
<td>2 (T-6)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>I</td>
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<td>+</td>
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<td>I</td>
</tr>
<tr>
<td>5 (T-72)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>I</td>
</tr>
<tr>
<td>6 (T-84)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>I</td>
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<td>7 (T-97)</td>
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</tr>
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<td>+</td>
<td>+</td>
<td>I</td>
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<td>10 (T-78)</td>
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<td>+</td>
<td>I</td>
</tr>
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<td>11 (T-99)</td>
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<td>12 (T-48)</td>
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<td>I</td>
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<td>13 (T-87)</td>
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<td>+</td>
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<td>I</td>
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<tr>
<td>15 (T-98)</td>
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<td>+</td>
<td>+</td>
<td>Intermediate</td>
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<tr>
<td>16 (T-57)</td>
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<td>+</td>
<td>+</td>
<td>Intermediate</td>
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Gastric ulcer

<table>
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<th>CagA</th>
<th>Vaculating assay</th>
<th>Type†</th>
</tr>
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<tbody>
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<td>69 (43526)</td>
<td>+</td>
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<td>+</td>
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Duodenal ulcer

<table>
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Non-ulcer dyspepsia

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<th>CagA</th>
<th>Vaculating assay</th>
<th>Type†</th>
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<td>49 (T-100)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>I</td>
</tr>
<tr>
<td>50 (T-102)</td>
<td>+</td>
<td>+</td>
<td>+</td>
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Gastric cancer

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<th>CagA</th>
<th>Vaculating assay</th>
<th>Type†</th>
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<tbody>
<tr>
<td>66 (T-113)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>I</td>
</tr>
<tr>
<td>67 (T-114)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>I</td>
</tr>
<tr>
<td>68 (T-108)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Intermediate</td>
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</table>

Controls

<table>
<thead>
<tr>
<th>Strain* no.</th>
<th>VacA</th>
<th>CagA</th>
<th>Vaculating assay</th>
<th>Type†</th>
</tr>
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<tr>
<td>69 (43526)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>I</td>
</tr>
</tbody>
</table>

Typing of vacA gene by polymerase chain reaction (PCR)

The vacA gene encodes a precursor protein made up of amino-terminal signal sequence, the cytotoxin proper, and a membrane protein which is removed during transmembrane transport. Genotyping of H pylori was proposed on the basis of the nucleotide sequence in the signal sequence (types s1a, s1b, and s2) and that in the cytotoxin region (m1, m2). We amplified the vacA gene by PCR using previously described genotype-specific primers.7 Thermal cycling was at 94 °C for 30 seconds, 50 °C for 15 seconds, and 72 °C for one minute, for a total of 35 cycles. PCR products were analysed by 1% agarose gel electrophoresis with ethidium bromide staining. ATCC43526 was used as a s1a/m1 control, and Tx30a was used as a s2/m2 control.

Statistical analysis

Fisher’s exact probability test and χ² test were used to test the independence on contingency tables. p<0.05 was considered to be statistically significant.

Results

Detection of vacA protein by immunoblot

VacA protein was detected among the cell surface proteins obtained from 59 of 68 (87%) H pylori isolates (fig 1A, tables 1 and 2). The positive rate was 14/16 (88%) in the isolates from patients with gastric ulcer, 15/17 (88%) from those with duodenal ulcer, 21/25 (84%) from those with non-ulcer dyspepsia, and 9/10 (90%) from gastric cancer patients. Statistically, the positive rate of VacA protein was not dependent on the gastroduodenal disease of the host (table 2).

The immunoblot analysis using whole cell lysates detected VacA protein even in the isolates that were negative by immunoblot analysis using cell surface proteins, suggesting that VacA protein was produced by all isolates but its secretion was disturbed in some of them.

Vacuolating assay

The vacuolating assay was positive in 54 of 59 (92%) VacA positive isolates and three of nine (33%) VacA negative ones (tables 1 and 2). Although discordant results were found in 8 of 68 (12%) isolates, the correlation between the two assays was strong enough (p<0.001) to assume that most isolates positive for VacA...
Virulence factors in *H pylori* isolates in Japan

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>Vacuolating activity (%)</th>
<th>VacA (%)</th>
<th>CagA (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gastric ulcer (n = 16)</td>
<td>13 (81)</td>
<td>14 (88)</td>
<td>15 (94)</td>
</tr>
<tr>
<td>Duodenal ulcer (n = 17)</td>
<td>15 (88)</td>
<td>15 (88)</td>
<td>15 (88)</td>
</tr>
<tr>
<td>Non-ulcer dyspepsia (n = 25)</td>
<td>20 (80)</td>
<td>21 (84)</td>
<td>21 (84)</td>
</tr>
<tr>
<td>Gastric cancer (n = 10)</td>
<td>9 (90)</td>
<td>9 (90)</td>
<td>10 (100)</td>
</tr>
<tr>
<td>Total (n = 68)</td>
<td>57 (84)</td>
<td>59 (87)</td>
<td>61 (90)</td>
</tr>
</tbody>
</table>

Table 3 Typing of vacA gene

<table>
<thead>
<tr>
<th>m1</th>
<th>m2</th>
<th>Neither</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>s1a</td>
<td>5</td>
<td>2</td>
<td>38</td>
</tr>
<tr>
<td>s1b</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Total</td>
<td>5</td>
<td>2</td>
<td>38</td>
</tr>
</tbody>
</table>


did secrete active cytotoxin.

Five strains were positive for VacA protein by immunoblot but negative for the vacuolating assay. These strains may be secretors of inactive cytotoxin similar to the Tx30a strain. Another three isolates were positive for the vacuolating assay but negative by the immunoblot analysis. The reason for this discrepancy is not known, but it may be that substances other than VacA induced vacuolation.

**Genotype of vacA gene**

The genotype of the vacA signal sequence was determined in 45 isolates, including six that were negative for VacA by both immunoblot and vacuolating assay. Interestingly, the signal sequence in all isolates was of type s1a, which is supposed to represent high vacuolating activity. The genotype of the mid region was also investigated in 45 isolates, as well as type m1 in five (11%) and type m2 in two (4%). PCR failed to amplify the mid region in the remaining 38 (84%) isolates (table 3).

**Detection of CagA protein by immunoblot**

The immunoblot analysis detected CagA protein in 61 of 68 (90%) *H pylori* isolates (fig 1B, tables 1 and 2). The positive rate was 15/16 (94%) in the isolates from patients with gastric ulcer, 15/17 (88%) from those with duodenal ulcer, 21/25 (84%) from those with non-ulcer dyspepsia, and 10/10 (100%) from gastric cancer patients. As in the case of the VacA protein described above, the positive rate of expression of CagA protein was high irrespective of the gastroduodenal disease of the host (table 2).

The estimated molecular mass of the CagA protein visualised by immunoblot varied from 120 to 140 kDa. This may indicate its heterogeneity among *H pylori* isolates, although we could find no correlation between its molecular mass and the gastroduodenal status of the host.

**Correlation between VacA and CagA**

The expression of CagA protein correlated strongly with that of VacA, as determined by immunoblot (p = 0.005), and with vacuolating cytotoxin activity in vitro (p = 0.002). Overall, 54 of 68 (79%) isolates were positive for cytotoxic activity and CagA protein. These isolates are defined as Type I strains in the following text. Type II strains were negative for these assays, and intermediate strains were neither Type I nor Type II.

The prevalence of Type I strains was 14/17 (82%) in the isolates from patients with duodenal ulcer, 13/16 (81%) from those with gastric ulcer, 18/25 (72%) from those with non-ulcer dyspepsia, and 9/10 (90%) from gastric cancer patients. The prevalence did not differ significantly between the groups.

Three isolates were negative for VacA protein, vacuolating activity, and CagA protein (Type II). The prevalence of these all-negative isolates was 3/25 (12%) in patients with non-ulcer dyspepsia and 0/43 (0%) in patients with peptic ulcer or gastric cancer. This difference was statistically significant (p = 0.046).

**Discussion**

Our current study shows that most *H pylori* strains isolated in Japan were positive for vacuolating cytotoxin and CagA protein. Indeed, 54/68 (79%) isolates were positive for the vacuolating assay and CagA protein and were therefore Type I strains. These results agree with our previous findings that the seropositivity of anti-VacA and anti-CagA antibodies is very high in Japanese patients with *H pylori* infection. We can conclude that most *H pylori* strains in Japan produce both VacA and CagA proteins not only in vitro but also in vivo.

**Recent studies on the cagA gene in Japan, Korea, and China, have shown that its prevalence is very high and not related to particular gastroduodenal diseases.**

For example, Pan et al. reported that 47 of 48 (98%) strains and 35 of 35 (100%) isolates were positive for the cagA gene in Chinese patients with peptic ulcers and gastritis respectively.

However, the current study does not rule out an association between the expression of VacA or CagA protein and the virulence of *H pylori*. The incidence of gastric cancer is high in Japan, with 38.2 deaths in 100 000 people in 1993. Epidemiological studies have shown that the prevalence of *H pylori* infection is significantly higher in patients with gastric cancer than in the general population in Japan as well as in other countries. It is possible that the high prevalence of infection with Type I strains shown in this study is associated with the high incidence of gastric cancer in Japan. In the current study, no patients with gastric cancer were
infected with VacA and CagA negative strains, suggesting a possibility that the incidence of gastric cancer may be low in the regions where infection with Type I strains is rare. It would require worldwide surveillance to elucidate whether Type I infection is actually related to gastric carcinogenesis. Recombinant proteins may be used in such studies for the examination of serum specific antibodies.

The vacuolating assay and analysis of VacA protein expression produced slightly different results. The difference between vacuolating activity and expression of VacA protein suggests that some strains secreted either inactive cytotoxin or only a small amount of active cytotoxin, so that the cytotoxin was detected by immunoblot analysis but its activity was not detected by the vacuolating assay. Indeed, some strains such as Tx30a secrete inactive cytotoxin,37 so that no vacuolation was observed by the vacuolating assay while VacA protein was detected by immunoblot analysis. Three isolates (T-12, T-47, T-87) were positive in the vacuolating assay but negative by immunoblot analysis. It is possible that substances other than VacA induced the vacuolation, or that VacA protein was decomposed. All isolates that were negative by immunoblot analysis of surface proteins showed VacA protein by immunoblot using cell lysates. These results suggest that VacA protein was produced, at least in bacterial cells.

The genotyping of the vacA gene showed genetic diversities between H pylori isolates from Japan and those from Western countries. The signal sequence of the vacA gene was of the s1 type (s1a subtypes) in all isolates examined in this study, and the mid region of the gene was of neither the m1 or m2 type in 38/45 (84%) isolates. A recent study reported similar results on the signal sequence in Japanese isolates34. These results were substantially different from the results obtained in Western countries. For example, Atherton et al57 reported that 21/23 (91%) isolates from patients with peptic ulcer but only 16/33 (48%) isolates from patients without ulcer were of the s1 type. They also reported that the mid region was either of the m1 or m2 subtype in 56 isolates, and no other subtypes were found. The distinct genotypes of the vacA gene of isolates in Japan may be associated with functional differences of the VacA protein, and may be a reason for the high incidence of gastric cancer in Japan.

CagA protein was reported to be produced in most cytotoxin positive strains of H pylori.16 However, recent studies have shown that the IL-8 induction may not be directly due to CagA protein, but to the proteins encoded by the genes in the pathogenicity island upstream of the cagA gene. These proteins are also co-expressed with CagA and therefore CagA may be used as a marker of enhanced virulence.35 36

In conclusion, we examined the expression of VacA and CagA proteins by H pylori strains clinically isolated in Japan. The prevalence of infection with strains that express vacuolating cytotoxin and CagA was very high compared with previous reports from Western countries. No significant correlation was found between the expression of these proteins by the isolates and gastroduodenal status of the host. Thus, these virulence factors cannot be used as markers to discern the risk of developing serious gastroduodenal disease in the host in Japan. However, it is possible that the high prevalence of infection with these Type I strains contributes to the characteristics of H pylori infection in Japan.

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