Chromatographic studies of the carbohydrate components of gastric and salivary mucopolysaccharides

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EDITORIAL SYNONYM These chromatographic studies suggest that the carbohydrate components of the gastric and salivary mucopolysaccharides contain glucose, galactose, mannose, and fucose. The gastric secretions of 12 patients with gastric carcinoma showed a marked increase in glucose.

An attempt has been made in our laboratory to study the human gastric mucopolysaccharides and their secretory patterns in normal and pathological conditions of the gastric mucosa. The mucopolysaccharides of the salivary secretions have also been investigated. At one stage of our investigation paper chromatography was used extensively. The results obtained are to be reported in this communication. The procedure for the fractional aspirations of the gastric secretion has already been described. (Schrager, 1961)

METHODS AND MATERIALS

Histamine and intravenous insulin were used as secretory stimulants. The gastric and salivary secretions were collected simultaneously and independently in ice-cooled test tubes. Eight aspirations of gastric secretions and eight specimens of saliva were obtained for every fractional gastric analysis. Adequate quantities for the routine investigations were removed from each specimen. The remaining gastric secretion and saliva were each pooled in separate containers and immediately stored in a deep freeze refrigerator overnight, thawed out, and centrifuged for 30 minutes in the refrigerated centrifuge (M.S.E. Super-Speed 40) at 10,000 r.p.m. to remove solid particles and cell debris. The supernatant was treated with diastase (total hexose: diastase—1 : 20 by weight) at 37°C. for five hours followed by dialysis at 4°C. against running water for five days and distilled water until free of chloride. The dialysed secretion was concentrated under reduced pressure at 30°C. to about one twentieth of its volume and hydrolysed with 2 N H₂SO₄ at 100°C. for two hours in sealed ampoules. The hydrolysate was neutralized with Ba(OH)₂, centrifuged, the supernatant evaporated to dryness in vacuo, and redissolved in a minimal quantity of water for chromatography on sheets of Whatman no. 1 paper (18 x 22.5 inches). At a later stage of our investigation we used Dowex-50 suspended in 0.05 N hydrochloric acid as a catalyst for the hydrolysis (Anastassiadis and Common, 1958). The sugars were developed by the ascending method using the solvent n-butanol: pyridine: water (6 : 4 : 3). Each chromatogram underwent three ascending developments, the paper was dried and replaced in fresh solvent for each development which lasted 36 hours. The total development of the chromatogram took about five and a half days. The sugars were detected with aniline hydrogen phthalate reagent (Partridge, 1949). Latterly we used the following reagent: 1-2 g. of p-anisidine, 1-7 g. of phthalic acid, 10 mg. of SnCl₂ H₂O in 100 ml. of ethanol (Moore, Spackman, and Stein, 1958). Between 100 and 200 ml. of gastric secretion and saliva is needed, depending on the amount of the mucopolysaccharides present, in order to obtain the concentration of the individual sugars necessary for chromatography.

The gastric secretion of 36 controls, consisting of 16 healthy volunteers and 20 patients who had no history of gastric symptoms or any present gastrointestinal complaints (Table I), 60 patients with duodenal ulcers (Table II), 18 patients with gastric ulcers (Table III), and

<table>
<thead>
<tr>
<th>TABLE I</th>
<th>SEX AND AGE OF 36 CONTROLS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age Groups (yr.)</td>
<td>0-20</td>
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<tr>
<td>Male</td>
<td>Nil.</td>
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<tr>
<td>Female</td>
<td>1</td>
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</table>

| 18 yr. |

<table>
<thead>
<tr>
<th>TABLE II</th>
<th>SEX AND AGE OF 64 PATIENTS WITH DUODENAL ULCER</th>
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</thead>
<tbody>
<tr>
<td>Sex</td>
<td>Age Groups (yr.)</td>
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<tr>
<td>Male</td>
<td>1</td>
</tr>
<tr>
<td>Female</td>
<td>Nil.</td>
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</table>
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TABLE III
SEX AND AGE IN 16 PATIENTS WITH GASTRIC ULCER

<table>
<thead>
<tr>
<th>Sex</th>
<th>Age Groups (yr.)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0-20</td>
</tr>
<tr>
<td>Male</td>
<td>3</td>
</tr>
<tr>
<td>Female</td>
<td>1</td>
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</tbody>
</table>

TABLE IV
SEX AND AGE IN 12 PATIENTS WITH CARCINOMA OF STOMACH

<table>
<thead>
<tr>
<th>Sex</th>
<th>Age Groups (yr.)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0-20</td>
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<tr>
<td>Male</td>
<td>1</td>
</tr>
<tr>
<td>Female</td>
<td>2</td>
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12 patients with carcinoma of the stomach (Table IV) were investigated. Only 15 controls, 20 patients with duodenal ulcer, eight patients with gastric ulcer, and four patients with gastric carcinoma provided sufficient quantities of saliva for chromatography.

RESULTS AND CONCLUSIONS

Our results do not support the common assumption that the human gastric mucopolysaccharides contain uronic acid. The chromatograms of the 120 gastric and 50 salivary secretions showed no uronic acid (Figs. 1-6), but when uronic acid was added to gastric secretion and hydrolysed it did appear on the chromatograms (Fig. 4). A solution of 1% chondroitin sulphate, treated as described above and chromatographed, also showed uronic acid (Fig. 4). These experiments indicate that our method of hydrolysis frees uronic acid and does not destroy it.

All our chromatograms invariably showed zones corresponding to glucose and mannose in addition to galactose and fucose. Glucose and mannose as components of human gastric mucopolysaccharides and glucose as a component of the salivary mucopolysaccharides are not mentioned in the literature as far as I know. Werner (1953) states that he could find no glucose in human gastric mucopolysaccharides after incubation with saliva and dialysis. Our investigation shows that both glucose and mannose are present in all the chromatographed secretions. The identification of glucose and mannose is based on the following: 1 The identity of the Rₖ values given with glucose and mannose standards. 2 The addition of glucose and mannose to the hydrolysates results in the intensification of the spots but not in the appearance of further ones. 3 Glucose disappears in the chromatograms of

FIG. 1.

FIG. 2.
GASTRIC SECRETION WITH URONIC ACID

NORMAL pH 7
NORMAL pH 2
STANDARDS

CARCINOMA CARCINOMA
NORMAL GASTRIC SECRETION NORMAL GASTRIC SECRETION
pH 7 pH 2
STANDARDS

FIG. 3.

FIG. 4.

GLUCURONIC ACID
GALACTOSAMINE
GLUCURONIC ACID
GALACTOSE
GLUCOSE
MANNOSE
FUCOSE
GLUCOLACTONE

FIG. 5.

FIG. 6.

GASTRIC SECRETIONS FROM PATIENTS WITH CARCINOMA OF THE STOMACH
STANDARD 2 STANDARD 1

GLUCURONIC ACID
GALACTOSE
GLUCOSE
MANNOSE
FUCOSE

FIG. 6.
gastric secretion hydrolysate incubated with glucose oxidase.

The presence of mannose has been demonstrated in other forms of human mucopolysaccharides, namely in duodenal (Dische, Di Sant’Agnese, Pallavicini, and Youlos, 1959) and bronchial mucopolysaccharides (Brogan, 1959), and in canine submaxillary saliva (Dische, Pallavicini, Kavasaki, Smirnow, Cizek, and Chien, 1962).

Glucose was demonstrated in all the salivas investigated (Fig. 2) as well as in all the gastric secretions from our controls and from patients suffering from duodenal ulcer and gastric ulcer after three redevelopments, each lasting 36 hours. It was not found on circular horizontal chromatograms after a 10-hour development (Fig. 3). Quantitative determination of the sugars, using Wilson’s (1959) elution method, show the ratios of galactose: glucose = 10:1 to 10:2. In order to separate the small quantities of glucose from the larger quantities of galactose prolonged and several redevelopments are needed.

The carbohydrate components of the gastric mucopolysaccharides of patients with carcinoma of the stomach show quantitative ratio differences from those of all other gastric secretions investigated. In the latter galactose and fucose are the predominant sugars. The secretion of the malignant stomach revealed a marked decrease in galactose and a great increase in glucose, the galactose being often reduced to a mere trace (Fig. 6). Horizontal circular chro-

matograms of gastric secretion of patients with gastric carcinoma always show glucose (Fig. 3). The large quantities of glucose present are separated even after one short development lasting 10 hours.

The quantitative relationship of the carbohydrate components of the gastric mucopolysaccharides is being further investigated.

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