Understanding selective trafficking of lymphocyte subsets

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Bonzo/CXCR6 may be important in trafficking effector T cells making it a potential target for therapeutic modulation of inflammatory diseases

This important study provides a mechanism to explain how different functional subsets of lymphocytes can be selectively recruited to tissue during an immune response by chemotactic cytokines (chemokines). The selective expression of particular chemokine receptors on subsets of lymphocytes allows these cells to be preferentially recruited to sites of inflammation. The authors report that a recently described chemokine receptor called Bonzo or CXCR6 is preferentially expressed on lymphocytes during their differentiation into type 1 effector cells, thereby linking the type 1/type 2 paradigm of immune responses with the burgeoning field of chemokines.

REGULATION OF TYPE 1 AND TYPE 2 IMMUNE RESPONSES

Cytokines produced by T (helper) cells are of critical importance in determining the outcome of many infectious and immune mediated diseases. The ability of the immune system to produce the appropriate set of cytokines in response to an infection and then to regulate that response as the infecting agent is cleared determines whether the response is successful and whether long term inflammatory damage persists. A crucial insight points to effectively eradicate an infectious agent, and the T1/T2 dichotomy is established on the basis of different cytokines.

The characteristic type 1 cytokines, interferon-γ (IFN-γ) and tumour necrosis factor α (TNF-α), have also been shown to play a role in the pathogenesis of a number of autoimmune and inflammatory disorders. In the normal human ileum, jejunum, and colon, intraepithelial lymphocytes produce type 1 cytokines and display cytolytic activity. In inflammatory bowel disease (IBD), especially Crohn’s disease, there is evidence of an excessive type 1 response with elevated production of IFN-γ and increased numbers of IFN-γ secreting cells which is also found in murine IBD models. The successful therapeutic use of anti-TNF-α antibodies in Crohn’s disease underscores this. Although the signals that drive differentiation into type 1 or type 2 responses in vitro are known, until recently it has been unclear as to how type 1 effector cells can be selectively recruited to a site of infection or inflammatory damage. The current paper suggests that a novel chemokine receptor, Bonzo or CXCL16, defines type 1 effector lymphocytes and allows them to interact with the chemokine CXCR16 and thereby to be selectively recruited to tissue.

CHEMOKINES AND CHEMOKINE RECEPTORS

Chemokines are 8–12 kDa heparin binding cytokines with the ability to attract leucocyte subsets to specific sites. More than 50 human chemokines have been identified and are classified according to the arrangement of conserved cysteine residues into four groups: CXC, CC, C, and CX3C. Chemokines activate seven transmembrane spanning, G protein linked receptors resulting in activation of downstream signals that determine the response in a given cell. In most circumstances the result is cytoskeletal rearrangement and changes in motility as a consequence of phospholipase C and Rho activation, although activation of protein tyrosine kinases can, under particular circumstances, lead to cell activation and proliferation. Most chemokine receptors bind several chemokines although some are specific for a single chemokine and two non-signalling receptors, the Duffy antigen on erythrocytes and heparan sulphate proteoglycans in the glyocalyx and extracellular matrix, bind multiple chemokines via low affinity interactions to retain them at specific sites.

The most important function attributed to chemokines is their ability to regulate leucocyte recruitment, retention, and positioning in tissue. They do this not only by stimulating directional migration but also by activating leucocyte integrins to bind to other cells and the extracellular matrix. Critical roles for chemokines have been demonstrated in the recruitment and retention of stem cells in bone marrow, thymocytes during T cell differentiation in the thymus, and...
dendritic cells in lymph nodes, as well as the control of effector cell recruitment to sites of infection or tissue damage. Chemokines can be usefully subdivided into two functional groups: inducible chemokines, produced in response to inflammation that recruit leukocytes, dendritic cells, and activated T cells; and constitutive chemokines expressed in the bone marrow, thymus, and secondary lymphoid organs that regulate physiological leukocyte trafficking. Regulation of leukocyte recruitment is complex and involves both secretion and presentation of chemokines in the target tissues and regulated expression of chemokine receptors on leukocytes during differentiation and activation. For example, naïve T cells which traffic from blood to lymph nodes express low levels of the inflammatory chemokine receptors CXCR3 and CCR5 but high levels of CCR7 which allows them to respond to the constitutive chemokine SLC, expression of which is largely confined to secondary lymphoid tissue. After activation by antigen in lymph nodes, naïve T cells undergo differentiation into effector cells and downregulate CCR7 while increasing expression of CXCR3 and CCR5, a phenotypic change which effectively excludes them from lymph nodes but allows them to be readily recruited to inflamed tissue where inflammatory chemokines that activate CXCR3 (IP-10, Mig, and ITAC) and CCR5 (RANTES, MIP-1α, and MIP-1β) are expressed.

**CHEMOKINES AND THE REGULATION OF TYPE 1/TYPe 2 IMMUNE RESPONSES**

Chemokines are important components of polarised type 1 and type 2 responses and act at several points during T cell differentiation. They can have direct effects on T cell activation as well as altering the migratory potential of the T cell and thereby altering where it goes and which cell it interacts with. In addition, dendritic cells (DCs), which are responsible for T cell activation and polarity of cytokine secretion, are also affected by chemokines both in terms of the type and number of DCs that are recruited to a site of inflammation and in terms of how the DC responds to activation. Thus signals delivered to DCs via CCR5 can drive IL-12 secretion whereas signals delivered via CCR2 inhibit IL-12 and these chemokines will define the subsequent T cell differentiation. Finally, the chemokine receptors expressed by effector T cells determine where these cells migrate. The complexity of these processes is illustrated by the results of studying immune responses in genetically modified animals in which chemokines or their receptors are either over expressed or deleted. Disrupting chemokines or chemokine receptors has produced unanticipated results which vary depending on the type of inflammatory model studied and at what stage during the evolution of the process the chemokine is inhibited.1–3

The present paper concentrates on the role of a novel chemokine CXCL16 in driving the recruitment of type 1 effector cells into tissue. Polarised T1 and T2 populations (characterised by production of IFN-γ and IL-4, respectively) have been shown to display distinct chemokine receptor profiles in vitro with Th1 cells expressing CCR5 allowing them to respond to the IFN inducible chemokines ITAC, IP10, and Mig (CXCL9, CXCL10, and CXCL11) and the CCR5 ligands RANTES, MIP-1α, and MIP-1β. Conversely, the CC chemokines, macrophage derived chemokine (MDC, CCL22) and thymus and activation regulated chemokine (TARC, CCL17), are preferential attractants for polarised type 2 cells that express CCR4 and CCR3.4,5 MDC production is induced by IL-4 and IL-13 and inhibited by IFN-γ and IL-12. There is also evidence for bilateral regulation because chemokines that attract type 1 cells via CXCR3 can concomitantly block the migration of type 2 cells in response to CCR3 ligands thus enhancing the polarisation of T cell recruitment.6,7 These and other studies have outlined the existence of chemokine based gradients that induce and sustain polarised type 1 and type 2 responses.

Recently, a new CXC chemokine, CXCL168 which binds to the receptor CXCR6 (Bonzo), was described. CXCL16 is unique among the CC and CXC chemokines in being a transmembrane protein. It was initially thought to be involved in CD8–dendritic cell interactions based on its expression on naïve CD8 T cells in mice but it appears that functional CXCL16 is absent from naïve T cells in humans making this an unlikely role in humans. A recent study9 showed that regulation of CXCR6 on lymphocytes parallels that of CCR5 and is inversely correlated with CXCR4, further evidence of a role for CXCR6 in type 1 responses.10 Interestingly, the only other known transmembrane chemokine CXCL1 (Fractalkine) is also found to be preferentially expressed by type 1 compared with type 2 T cells.10,11 Kim et al have now extended these observations to demonstrate that CXCR6 expression is upregulated on T cells by interactions with dendritic cells and that the addition of IL-12 enhances the generation of CXCR6 cells whereas IL-4 reduces it. Furthermore, most of the CXCR6 cells in blood are IFN-γ secreting helper T cells, cytotoxic T cells, or CD56+ T cells that are enriched for effector functions and IFN-γ release. Thus upregulation of CXCR6 on primed T cells by DCs in an IL-12 rich environment will lead to the release of effector cells with the potential to mediate type 1 responses. In order to complete the story, the authors describe enrichment of CXCR6 cells in inflamed tissue implying the receptor is involved in effector cell recruitment to or retention in tissue. If this model is correct, further characterisation of the CXCR6+ T cells in specific diseases is of great importance. Their potential to mediate effector functions may be beneficial in infectious diseases where it is likely that CXCR6 cells include a high proportion of virus specific T cells in, for instance, chronic viral hepatitis. However, the same effector cells could lead to inappropriate recruitment and tissue damage in type 1 autoimmune disorders, as suggested by the finding that CXCR6+ T cells are enriched in the joint in rheumatoid arthritis. The fact that circulating CXCR6+ T cells include αβ+ T cells suggests they could also be recruited to the gut in IBD. CXCR6 and CXCL16 therefore become interesting therapeutic targets and provoke a dualistic approach to immune modulation. Whereas neutralisation of CXCR6+ effector cells might be expected to be beneficial in type 1 driven immune mediated diseases such as rheumatoid arthritis and Crohn’s disease, enhancement of these cells by either selective activation in vivo or adoptive transfer may promote antiviral responses in patients with chronic viral hepatitis. It would appear that it is not yet bedtime for Bonzo.

**REFERENCES**


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