Mobilisation of enterocyte fat stores by oral glucose in humans

M D Robertson, M Parkes, B F Warren, D J P Ferguson, K G Jackson, D P Jewell, K N Frayn

Background and aims: When a high fat oral load is followed several hours later by further ingestion of nutrients, there is an early postprandial peak in plasma triacylglycerol (TG). The aim of this study was to investigate the location and release of lipid from within the gastrointestinal tract.

Methods: Ten healthy patients undergoing oesophago-gastro-duodenoscopy (OGD) were recruited. At t=0, all patients consumed a 50 g fat emulsion and at t=5 hours they consumed either water or a 38 g glucose solution. OGD was performed at t=0 hours and jejunal biopsy samples were evaluated for lipid storage. A subgroup of five subjects then underwent a parallel metabolic study in which postprandial lipid and hormone measurements were taken during an identical two meal protocol.

Results: Following oral fat at t=0, samples from patients that had subsequently ingested glucose exhibited significantly less staining for lipid within the mucosa and submucosa of the jejunum than was evident in patients that had consumed only water (p=0.028). There was also less lipid storage within the cytoplasm of enterocytes (p=0.005) following oral glucose. During the metabolic study, oral glucose consumed five hours after oral fat resulted in a postprandial peak in plasma TG, chylomicron-TG, and apolipoprotein B48 concentration compared with oral water.

Conclusion: After a fat load, fat is retained within the jejunal tissue and released into plasma following glucose ingestion, resulting in a peak in chylomicron-TG which has been implicated in the pathogenesis of atherosclerosis.

MATERIALS AND METHODS

Patients

Ten patients (six females, mean age 36.7 years (range 21–55); mean weight 57.8 kg) undergoing diagnostic oesophago-gastro-duodenoscopy (OGD) at the John Radcliffe Hospital, Oxford, were recruited into the study. All patients were free from concomitant cardiovascular, pulmonary, and renal disease, and exhibited no symptoms of fat malabsorption.

Abbreviations: TG, triacylglycerol; ER, endoplasmic reticulum; apo B48, apolipoprotein B48; MTP, microsomal triglyceride transfer protein; OGD, oesophago-gastro-duodenoscopy; Sf, Svedberg flotation rate; CV, coefficient of variation; VLDL, very low density lipoprotein; DGAT, acyl-CoA:diacylglycerol acyltransferase.
Informed written consent was obtained and the study was approved by the Oxfordshire Clinical Research Ethics Committee.

**Study design**

The study design was based on an established protocol in which a high fat breakfast is followed five hours later by a “second meal”. Small volumes of water and glucose solution were chosen as “second meal” negative and positive controls, respectively, due to the experimental constraints of performing an OGD only one hour after nutrient ingestion. Patients were randomised into two groups to consume either a small glass of water (100 ml) or a glucose drink (38 g of glucose in 100 ml water) five hours following a high fat load. The two treatments will be referred to as “fat load-water” and “fat load-glucose” and the groups as the “water group” and the “glucose group”.

After an overnight fast, an oral fat load (50 g liquid fat: Calogen; SHS, Merseyside, UK) was given to all patients at 07:00 on the morning of their endoscopy. At 12:00, patients received their second nutrient stimulus (glucose or water). OGD was performed at 13:00 (six hours after the oral fat load and one hour after the glucose or water) using a paediatric colonoscope which allowed intubation and biopsy of the jejunum. Frozen sections were processed with oil red-O to identify neutral fat. Histological preparations stained with oil red-O were quantified for lipid content using a semiautomatic analysis system (IBAS, Kontron, Germany) with 5–10 fields of mucosal and submucosal tissue included. Tissue for electron microscopy was fixed with 4% glutaraldehyde. Samples were post-fixed in 1% osmium tetroxide in a 0.1 M phosphate buffer, dehydrated in ethanol, followed by treatment with propylene oxide prior to embedding in Spurr’s epoxy resin. Thin sections were cut and stained with uranyl acetate and lead citrate prior to examination with a JEOL 1200EX microscope. Between six and 10 electron micrographs were taken at magnifications ranging from ×5700 to ×23 250 from a single biopsy in each patient. Samples were examined in a blinded fashion and graded independently from 0 to ++++ for lipid content present within the sections.

**Metabolic study**

As it was not possible to take blood samples during OGD, a subgroup of five patients (two females) from the original biopsy group underwent parallel metabolic investigations. This was a randomised crossover design with patients undergoing both protocols (fat load-water; fat load-glucose) on separate days two weeks apart.

After an overnight fast, patients attended the metabolic research unit and an indwelling cannula was inserted into the antecubital vein of the forearm and two fasting blood samples were taken. The 50 g of liquid fat was given at time=0 with blood samples taken at hourly intervals. At t=300 minutes, the second drink of either water or glucose solution was given and blood samples were collected every 15 minutes for a further 90 minutes.

Blood samples were collected into heparinised tubes (Sarstedt, Leicester UK) for analysis of plasma glucose, insulin, and TG, and chylomicron rich fraction (Svedberg flotation rate (Sf) >400) TG and apo B48 concentrations. A preservative was added to plasma to prevent proteolytic degradation of apo B48 (EDTA (0.5 ml/l), phenylmethylsulphonyl fluoride (10 mmol/l dissolved in isopropanol) and aprotinin (10 000 MIE/l)). The Sf >400 lipoprotein fraction was isolated by density gradient ultracentrifugation. Briefly, the density of plasma was adjusted to d=1.10 kg/l with addition of d=1.42 kg/l KBr and then overlayered with d=1.063, 1.020, 1.000, and 1.000 kg/l KBr.

**Table 1**

<table>
<thead>
<tr>
<th>Grade of biopsy</th>
<th>100 ml water (n=5)</th>
<th>38 g glucose in 100 ml water (n=5)</th>
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<td>Mean grade for group</td>
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**Figure 1**

(A) Electron micrograph of jejunal tissue from a female patient one hour after oral glucose ingestion and six hours after a 50 g fat load. The microvilli (MV) and apical junctional complex (AJ) denotes the apical surface of the enterocyte. Numerous mitochondria (M), strands of rough endoplasmic reticulum (ER), and a lysosome (Ly) are visible. This represents grade 0 lipid storage. Bar is 1 µm. (B) Electron micrograph of jejunal tissue from a female patient one hour after oral water ingestion and six hours after a 50 g fat load. Large numbers of chylomicron sized lipid droplets (L) are present in the apical cytoplasm identified by the presence of microvilli (MV). Lipid is present in both the apical and basal regions of the cell. This represents grade ++++ lipid storage. M, mitochondrion. Bar is 1 µm.
and 1.006 kg/l NaCl solutions. Ultracentrifugation was performed at 40 000 rpm (202 000 g) for 32 minutes in a SW 40Ti swinging bucket rotor (Beckman Instruments, High Wycombe, UK) at 15°C. The top 1 ml of the gradient (Sf >400 fraction) was carefully isolated. Apo B48 was measured using a specific ELISA with a few modifications. A heptapeptide-thyroglobulin conjugate consisting of the terminal residues of the apo B48 molecule was used as the coating material in the ELISA format. Samples were incubated with a specific polyclonal anti-apo B48 antibody, which recognises the C terminal region of the protein on the surface of lipoprotein particles and does not show cross reactivity for apo B100. The intra-assay coefficient of variation (CV) was 4.5%. Plasma glucose concentrations were measured enzymatically using an IL Monarch automated analyser (Instrumentation Laboratory, Warrington UK). Insulin was measured by radioimmunoassay using a commercially available kit (Pharmacia and UpJohn, Milton Keynes, UK) exhibiting both inter- and intra-assay coefficients of variation of less than 10%. TG concentrations in plasma and (Sf >400) lipoprotein fraction were measured using an enzyme based colorimetric kit (Instrumentation Laboratory). The intra-assay CV was 1.7%.

**Statistics**

Data were analysed using SPSS version 10 (SPSS inc.; Chicago, Illinois, USA). Plasma results presented in time courses are mean (SEM) values. Postprandial data were analysed using repeated measures ANOVA. Summary data such as areas under the curve were calculated by the trapezoid method and analysed by paired Student's t test. Biopsy data (grade of electron microscopy samples and quantitative staining with oil red-O) were analysed by one way ANOVA for unpaired samples. A p value of <0.05 was taken as the level of significance.

**RESULTS**

During endoscopic examination, all 10 patients were found to have normal jejunal mucosa (2 × lactose intolerant; 4 × irritable bowel syndrome; 2 × non-ulcer dyspepsia; 1 × oesophagitis). There was no evidence of liquid fat emulsion remaining within the lumen of the stomach, duodenum, or jejunum six hours after the oral fat load in either treatment group.

**Biopsy study**

Table 1 displays the grade of fat deposit present within the jejunal biopsy sample. Six hours after a fat load there were numerous lipid vacuoles of varying size within the cytoplasm of the enterocytes for 6/10 patients (1/5 fat load-glucose (fig 1A); 5/5 fat load-water; p=0.005 (fig 1B)). The vacuoles were not limited by a “unit membrane” which distinguishes the lipid structures from other cell organelles. It is likely that there were some deposits of phospholipids forming a limiting layer but we were unable to detect these using our histological methods. The lipid droplets ranged in size from approximately 0.5 µm to 6 µm, with significant variation both within and between subjects. These were not confined to the basolateral region of the cell and did not appear to be associated with the secretory apparatus (ER or Golgi) which was clearly visible following electron microscopy (fig 2). Lipid was also evident in the interstitium between adjacent enterocytes. On light microscopy, jejunal samples from the water group exhibited significantly more lipid staining than did samples from the
glucose group. Lipid was evident within both the enterocytes and the lamina propria of the villus tip (fig 3A).

**Metabolic study**

As the metabolic load was identical in both groups for the first 300 minutes of the study, data are only presented for 300–390 minutes (90 minutes after the second stimulus).

The plasma chylomicron-TG (SF >400) response following water or glucose ingestion is shown in fig 4. There was a significant peak in chylomicron-TG 45 minutes after glucose ingestion which was not evident in the water group (p=0.033). A similar pattern was seen for plasma TG concentration, with a peak observed 45 minutes following glucose ingestion but not with oral water (p=0.008) (data not shown). Apo B48 concentrations within the chylomicron rich fraction (SF >400) were also significantly higher in the glucose group, with a peak again reached by 45 minutes post ingestion (fig 5). The TG/apo B48 ratio in the chylomicron rich fraction in the first 300 minutes of the study (after fat ingestion alone) was significantly lower than that obtained (300–390 minutes) following glucose ingestion (10.6 v 15.6 mmol TG per μg apo B48; p=0.043).

As expected, the plasma glucose concentration rose with a peak at 45 minutes after oral glucose ingestion whereas there was no effect of oral water. There was an increase in the plasma insulin concentration which remained elevated for the remainder of the study despite a drop in plasma glucose concentration (fig 6).

**DISCUSSION**

The combination of metabolic data with electron microscopy images has allowed demonstration for the first time that during the second meal effect for plasma TG, the cytosolic storage pool of lipid contained within the jejunal enterocytes is significantly reduced. Six hours after the high fat load there were significant amounts of lipid visible within the enterocytes, the intercellular spaces, and the lamina propria of the villus tip. The lipid visible within the jejunal tissue was significantly reduced one hour after oral glucose, coinciding in the parallel metabolic studies with a peak in endogenous lipid particles in the plasma. The second meal effect appears to reflect both an increase in the mobilisation of cytosolic TG and an increase in the movement of these lipid particles through the secretory pathway and from lymph to circulation; an effect which we have shown may be independent of further fat ingestion.

The large lipid droplets (up to 6 μm) observed in this study and the lack of integrity with the ER membrane led to the conclusion that following fat absorption, re-esterified fat is stored within the cytoplasm. This adds credence to the theory that the transfer of TG from the ER into the secretory pathway can become saturated, resulting in storage pools of TG within the cytosol. Chylomicron assembly is similar to the assembly of very low density lipoprotein (VLDL) particles in the liver. In hepatocytes, it is known that a proportion of the secreted TG as VLDL-TG is derived from this cytosolic pool; that the same phenomenon should occur within enterocytes is not perhaps surprising, although until now difficult to demonstrate in vivo and in humans.

For the stored TG to be made available, it must undergo hydrolysis within the cytosol followed by re-esterification in the ER. It has been shown recently that rat enterocytes express pancreatic TG lipase and in Caco-2 cells the majority of this lipase activity is found in the cytosolic fraction. Addition of exogenous lipase to the apical surface of Caco-2 cells results in increased fat hydrolysis and enhanced basolateral lipid secretion, an observation which would explain our in vivo second meal effect. The relationship between oral glucose and cytosolic lipase concentration in this study may be modulated via the concomitant increase in circulating plasma insulin. Increased TG hydrolysis has been demonstrated in hepatocytes cultured with insulin although the effect of circulating insulin on cytosolic lipase within enterocytes is currently...
unknown. In Caco-2 cells, short term incubation with insulin also increases intestinal fatty acid binding protein levels, which may be important for the movement of these newly formed products of lipolysis to the ER. The potential for the rate of chylomicron release from the enterocyte to be “controlled” by the ER itself cannot be disregarded. Recent studies in mice lacking the enzyme acyl-CoA:diacylglycerol acyltransferase 1 (DGAT1), which catalyses the final step in TG synthesis, have revealed large lipid droplets within the cytoplasm of enterocytes. 12 DGAT activity in the heart 28 and adipocyte 29 has been shown to increase in animal models of diabetes although whether this is related to increased circulating levels of glucose per se remains to be determined. Following exocytosis from the basolateral membrane, the newly secreted chylomicron particles first enter the intercellular space before migrating via the lacteals into the circulation; however, a proportion of fat is retained in the lymphatics. 30 Following oral glucose, there was significantly less fat present within the lamina propria compared with that following water or the original high fat load also contributing to the observed second postprandial peak in plasma lipids. Factors affecting the rate of movement of preformed chylomicrons through the lymph are therefore likely to be important in determining the magnitude of the second meal effect. Glucose in the luminal fluid has been shown to stimulate both fluid uptake and lymph flow, 31 increasing sympathetic activity leading to vasodilatation of the submucosal venules and enhanced intestinal blood flow. 32 Insulin also leads, directly or indirectly, to vasodilatation in muscle 33 and adipose tissue. 34

Apo B48, synthesised by bound ribosomes in the ER, is the major structural protein component and is essential for the formation and secretion of chylomicrons. Following oral glucose, the chylomicron apoB48 particle also differed significantly from the ratio following the earlier fat load which may be important as a difference in the composition of the secreted lipoproteins between the original fat load and the glucose load. The second meal effect increased the chylomicron TG/apoB48 ratio, indicating the secretion of larger particles, as each chylomicron is known to contain only a single apo B48 particle. 35 Addition of lipid to the newly formed apo B48 to form the lipoprotein particle occurs in the lumen of the ER/Golgi and is mediated by MTP and so regulation of MTP activity is likely to be important in determining particle size. Inhibition of MTP in Caco-2 cells has been shown to reduce the amount of TG per particle and conversely increases in MTP result in the secretion of larger particles. 36 Whether there is acute regulation of MTP by either glucose or insulin has yet to be demonstrated. This secondary release of lipids from the “enteral” store has been studied metabolically for many years 23–25 without knowing the site of this “enteral” store. There is no reported difference in the magnitude of the second meal effect between low and high fat meals; nothing is known on the effects of individual amino acids on this process although from our metabolic studies of chylomicron content would appear to be important. 2 In this study, we demonstrated an almost complete mobilisation of all of the stored cytosolic TG following oral glucose (4/5 subjects showed no fat storage following glucose ingestion) with only minimal amounts of fat still visible within the lamina propria.

In conclusion, following a high fat meal in humans, a proportion of the fat load remains both within the enterocytes as neutral lipid droplets contained within the cytosol and as preformed chylomicrons retained within the lymph. When glucose is ingested as a second meal, TG mobilisation within the enterocyte is stimulated, producing new chylomicron particles which are exported from the cell. A concomitant increase in lymph flow results in movement of chylomicron particles, resulting in a rapid peak in plasma TG. In this study we have made significant progress in locating the storage pool of dietary TG responsible for the second meal effect in vivo in humans; however, further work is necessary, potentially utilising cultured human enterocytes, in determining the precise mechanisms involved.

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**Authors’ affiliations**

M D Robertson, K N Frayn, Metabolic Research Group, Oxford Centre for Diabetes, Endocrinology, and Metabolism, University of Oxford, Oxford, UK

M Parkes, D P Jewell, Nuffield Department of Gastroenterology, University of Oxford, Oxford, UK

B F Warren, D J Ferguson, Department of Pathology, John Radcliffe Hospital, Oxford, UK

K G Jackson, Hugh Sinclair Unit for Human Nutrition, School of Food Biosciences, University of Reading, Reading, UK

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