Administration of protegrin peptide IB-367 to prevent endotoxin induced mortality in bile duct ligated rats

A Giacometti, O Cirioni, R Ghiselli, F Mocchegiani, G D’Amato, M Simona Del Prete, F Orlando, W Kamysz, J Lukasiak, V Saba, G Scalise

**Background:** Postoperative morbidity in patients with obstructive jaundice remains high because of increased susceptibility to endotoxin and the inflammatory cascade.

**Aims:** An experimental study was designed to investigate the efficacy of protegrin peptide IB-367, an antimicrobial positively charged peptide, in neutralising *Escherichia coli* 0111:B4 lipopolysaccharide (LPS) in bile duct ligated rats.

**Methods:** Adult male Wistar rats were injected intraperitoneally with 2 mg/kg *E coli* 0111:B4 LPS one week after sham operation or bile duct ligation (BDL). Six groups were studied: sham with placebo, sham with 120 mg/kg tazobactam- piperacillin (TZP), sham with 1 mg/kg IB-367, BDL with placebo, BDL with 120 mg/kg TZP, and BDL with 1 mg/kg IB-367.

**Results:** Main outcome measures were: endotoxin and tumour necrosis factor α (TNF-α) concentrations in plasma, evidence of bacterial translocation in blood and peritoneum, and lethality. After LPS, TNF-α plasma levels were significantly higher in BDL rats compared with sham operated animals. IB-367 caused a significant reduction in plasma endotoxin and TNF-α concentrations compared with placebo and TZP treated groups. In contrast, both TZP and IB-367 significantly reduced bacterial growth compared with saline treatment. Finally, LPS induced 60% and 55% lethality in BDL placebo and TZP treated rats and no lethality in sham operated rats, while only IB-367 significantly reduced lethality to 10%.

**Conclusions:** By virtue of its dual antimicrobial and antiendotoxin properties, IB-367 could be an interesting compound to inhibit bacterial translocation and endotoxin release in obstructive jaundice.
Materials and methods

Animals
Adult male Wistar rats weighting 250–300 g were used for all experiments. All animals were housed in individual cages under constant temperature (22°C) and humidity with a 12 hour light/dark cycle, and had access to chow and water ad libitum throughout the study. The study was approved by the animal research ethics committee of the INRCA IRRCS, University of Ancona, Ancona, Italy.

Agents
IB-367 was synthesised manually (Faculty of Pharmacy, Medical University of Gdansk, Poland) by 9-fluoroenylmethoxycarbonyl solid phase chemistry using the protocol described previously.25 The peptide was purified by solid phase extraction on Kromasil sorbent (C8, 5 μm, 100 Å).25 The resulting fractions with purity greater than 97–98% were tested by high pressure liquid chromatography. The peptide was analysed by chemical analysis and matrix assisted laser desorption ionisation mass spectrometry.

IB-367 was dissolved in distilled H2O at 20 times the required maximal concentration. Successively, it was diluted in physiological saline. Tazobactam-piperacillin (Wieth Lederle, Aprilia, Italy) was diluted in accordance with the manufacturer’s recommendations. Solutions were made fresh on the day of assay.

Experimental design
Rats were assigned to one of six groups, each containing 20 animals: sham operation with placebo, sham operation with tazobactam-piperacillin (TZP), sham operation with IB-367, bile duct ligation (BDL) with placebo, BDL with TZP and BDL with IB-367. Rats were anaesthetised at time t=−7 days by intramuscular injection of ketamine 30 mg/kg body weight. The surgical procedures were either a sham operation in which, after opening the abdomen via a 2 cm midline incision, the bile duct was dissected from surrounding tissues, or BDL, in which the bile duct was prepared free and doubly ligated using 5-0 silk and divided according to the procedure described by Lee. After either operation the abdomen was closed in two layers using 4-0 chromic catgut. All procedures were performed observing strict asepsis. One week later (time 0) each animal was injected intraperitoneally with 2.0 mg of Escherichia coli serotype 0111:B4 LPS in a total volume of 500 μl of sterile saline. Immediately after injection, the animals in the six groups received intraperitoneally isotonic sodium chloride solution (placebo), 1 mg/kg IB-367, and 120 mg/kg TZP, respectively. The animals were returned to individual cages and examined daily. Toxicity was evaluated on the basis of the presence of any drug related adverse effect—that is, local signs of inflammation, anorexia, weight loss, vomiting, diarrhoea, fever, and behavioural alterations. The animals were monitored for the subsequent 48 hours.

Blood sampling procedure
For blood cultures and determination of endotoxin and TNF-α in plasma, 0.2 ml blood samples were collected from a tail vein into a sterile syringe and transferred to tubes containing ethylenediaminetetraacetic acid tripotassium salt (EDTA-K3).

Blood samples were taken before operation (t=−7 days), before (t=0) LPS administration, and 2, 6, 24, and 48 hours after injection.

Biochemical assays
Endotoxin concentrations were measured by the Limulus amoebocyte lysate test (E-TOXATE; Sigma-Aldrich). Plasma samples were serially diluted twofold with sterile endotoxin free water and heat treated for five minutes in a water bath at 75°C to destroy inhibitors that can interfere with activation. Endotoxin content was determined as described by the manufacturer. Endotoxin standards (0, 0.015, 0.03, 0.06, 0.125, 0.25, and 0.5 EU/ml) were tested in each run, and the concentration of endotoxin in the test samples was calculated by comparison with the standard curve.

TNF-α levels were measured with a commercially available solid phase sandwich enzyme linked immunosorbent assay (Nuclear Laser Medicine, Srl, Settala, Italy) according to the protocol supplied by the manufacturer. Standards and samples were incubated with a TNF-α antibody coated 96 well microtitre plate. The wells were washed with buffer and incubated with biotinylated anti-TNF-α antibody conjugated to streptavidin-peroxidase. This was washed away and colour was developed in the presence of chromogen (tetramethylbenzidine) substrate. The intensity of the colour was measured in a MR 700 Microplate Reader (Dynatech Laboratories, Guernsey, UK) by reading absorbance at 450 nm. Results for the samples were compared with the standard curve to determine the amount of TNF-α present. All samples were run in duplicate. The lower limit of sensitivity for TNF-α by this assay was 0.05 ng/ml. The intra-assay and interassay coefficients of variation were 5.8% and 7.7%, respectively.

Microbiological studies
To evaluate the presence of BT, bacterial analysis was performed on blood and intra-abdominal fluid obtained under sterile conditions.

Blood samples for aerobic and anaerobic cultures were obtained by aseptic percutaneous transthoracic cardiac puncture from all animals, 48 hours after the end of LPS infusion or at the time of death. Before cardiac puncture, surviving animals were killed with chloroform.

In addition, to perform quantitative bacterial evaluation of intra-abdominal fluid from each animal, 10 ml of sterile saline were injected intraperitoneally and samples of the peritoneal lavage fluid were serially diluted and a 0.1 ml volume of each dilution was spread onto blood agar plates for enumeration of developed colonies. For isolation of anaerobes, specimens were inoculated onto Columbia blood agar plates enriched with hemin and menadione, and incubated in an anaerobic chamber in 80% N2-10% H2-10% CO2. The limit of detection was <1 log5 colony forming units (CFU)/ml. Plates were incubated both in air and under anaerobic conditions at 35°C for 48 hours.

Statistical analysis
Survival data were compared by the log rank test. Qualitative results for blood cultures were analysed by the χ2 test, Yates’ correction, or Fisher’s exact test, depending on the sample size. Quantitative evaluation of bacteria in intra-abdominal
fluid cultures are presented as mean (SEM); statistical comparisons between groups were made using analysis of variance (ANOVA). Post hoc comparisons were performed by Bonferroni’s test. Plasma endotoxin and TNF-α levels were analysed using the Kruskal-Wallis test; multiple comparisons between groups were performed using the appropriate standard procedure. Each comparison group contained 20 rats. Significance was accepted when \( p < 0.05 \).

**RESULTS**

The main outcome measures were: endotoxin and TNF-α concentrations in plasma, evidence of BT, and lethality.

**Endotoxin and TNF-α levels**

On \( t=-7 \) days, endotoxin levels were not significantly different between the groups (table 1). However, at \( t=0 \), one week after operation, there were important changes in endotoxin levels (0.098 (0.01) EU/ml in all sham rats and 0.330 (0.07) EU/ml in all BDL rats, respectively) (\( p < 0.001 \)).

Two hours after induction of endotoxaemia, endotoxin levels were 5030.6 (1076.7) EU/ml and 5871.8 (1115.6) EU/ml in the sham and BDL placebo groups, respectively. Similar results were observed in the two TZP treated groups. In contrast, IB-367 caused a significant reduction in plasma endotoxin concentration compared with placebo and TZP treated groups (487.3 (106.7) EU/ml and 544.5 (121.3) EU/ml in sham and BDL rats, respectively) (\( p < 0.001 \)).

Comparable results were found six hours after LPS injection (table 1). At 24 and 48 hours, in all surviving animals, plasma endotoxin concentrations returned towards levels detected before LPS injection (\( t=0 \)). The lowest values were detected in the sham protegrin treated group (table 1).

TNF-α levels in all groups were \( < 0.05 \) ng/ml at \( t=-7 \) days. Overall, at \( t=0 \) there were significant differences (\( p < 0.001 \)) in TNF-α levels between sham operated and BDL rats (0.08 (0.02) ng/ml and 0.11 (0.03) ng/ml, respectively). Within two hours of induction of endotoxaemia, TNF-α levels reached a peak in all groups, being significantly higher in BDL treated with placebo or TZP followed by sham treated with placebo or TZP (table 2). IB-367 achieved a significant reduction in plasma TNF-α concentration compared with the four above mentioned groups (\( p < 0.01 \)). In all groups, TNF-α levels returned towards prechallenge concentrations six hours after injection of LPS (table 2).

**Bacterial translocation**

Bacteriological evaluation showed no positive blood or intra-abdominal fluid cultures in sham operated animals. In contrast, for the BDL model, blood cultures grew bacteria in 15%, 5%, and 10% of placebo, TZP, and IB-367 treated rats, respectively. Finally, in the BDL model, evidence of bacterial colonisation in peritoneal fluid was demonstrated in 25%, 10%, and 15% of placebo, TZP, and IB-367 treated rats, respectively. Specifically, for BDL placebo treated rats, quantitative cultures of intra-abdominal fluid demonstrated the highest bacterial numbers, averaging \( 3.6 \times 10^7 \) (1.1 \( \times 10^7 \)) CFU/ml, while TZP exerted the strongest antibacterial activity producing a mean of \( 2.5 \times 10^5 \) (0.7 \( \times 10^5 \)) CFU/ml (fig 1). Gram negative and Gram positive bacteria were simultaneously isolated from more than 70% of animals. The pathogens isolated from both blood and abdominal fluid were mainly members of the Enterobacteriaceae family, including E coli, Enterobacter spp.

**Table 1** Endotoxin levels (EU/ml) in sham operated rats and bile duct ligated (BDL) rats at different times

<table>
<thead>
<tr>
<th>Group*</th>
<th>-7 days</th>
<th>Time 0</th>
<th>2 h</th>
<th>6 h</th>
<th>24 h</th>
<th>48 h</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham placebo</td>
<td>0.093 (0.021)</td>
<td>0.101 (0.035)</td>
<td>5030.6 (1076.7)</td>
<td>4122.5 (1055.2)</td>
<td>0.272 (0.042)</td>
<td>0.115 (0.035)</td>
</tr>
<tr>
<td>Sham TZP</td>
<td>0.099 (0.013)</td>
<td>0.097 (0.029)</td>
<td>4884.3 (1003.2)</td>
<td>4633.6 (1015.1)</td>
<td>0.259 (0.044)</td>
<td>0.109 (0.032)</td>
</tr>
<tr>
<td>Sham IB-367</td>
<td>0.090 (0.026)</td>
<td>0.096 (0.019)</td>
<td>487.3 (106.7)</td>
<td>422.9 (98.4)</td>
<td>0.088 (0.027)</td>
<td>0.098 (0.012)</td>
</tr>
<tr>
<td>BDL placebo</td>
<td>0.096 (0.034)</td>
<td>0.357 (0.092)*</td>
<td>5871.8 (1115.6)</td>
<td>5573.7 (1095.3)</td>
<td>0.476 (0.089)*</td>
<td>0.398 (0.068)*</td>
</tr>
<tr>
<td>BDL TZP</td>
<td>0.090 (0.017)</td>
<td>0.331 (0.088)*</td>
<td>5699.6 (1077.4)</td>
<td>5491.1 (1021.8)</td>
<td>0.451 (0.094)*</td>
<td>0.412 (0.073)*</td>
</tr>
<tr>
<td>BDL IB-367</td>
<td>0.095 (0.028)</td>
<td>0.302 (0.104)*</td>
<td>544.5 (121.3)*</td>
<td>501.3 (98.5)*</td>
<td>0.279 (0.093)</td>
<td>0.291 (0.102)*</td>
</tr>
</tbody>
</table>

Results are given as mean (SD).

*TZP, tazobactam-piperacillin 120 mg/kg; IB-367 1 mg/kg.

Each measurement refers to:b20 animals;c13 animals;dseven animals;esix animals;f15 animals;g10 animals;hnine animals;i18 animals.

**Table 2** Tumour necrosis factor levels (ng/ml) in sham operated rats and bile duct ligated (BDL) rats at different times

<table>
<thead>
<tr>
<th>Group*</th>
<th>-7 days</th>
<th>Time 0</th>
<th>2 h</th>
<th>6 h</th>
<th>24 h</th>
<th>48 h</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham placebo</td>
<td>( &lt; 0.05 )</td>
<td>0.08 (0.02)</td>
<td>1.12 (0.45)</td>
<td>0.26 (0.07)*</td>
<td>( &lt; 0.05 )*</td>
<td>( &lt; 0.05 )*</td>
</tr>
<tr>
<td>Sham TZP</td>
<td>( &lt; 0.05 )</td>
<td>0.09 (0.02)</td>
<td>0.98 (0.22)</td>
<td>0.31 (0.08)*</td>
<td>( &lt; 0.05 )*</td>
<td>( &lt; 0.05 )*</td>
</tr>
<tr>
<td>Sham IB-367</td>
<td>( &lt; 0.05 )</td>
<td>0.07 (0.02)</td>
<td>0.43 (0.16)</td>
<td>0.13 (0.05)*</td>
<td>( &lt; 0.05 )*</td>
<td>( &lt; 0.05 )*</td>
</tr>
<tr>
<td>BDL placebo</td>
<td>( &lt; 0.05 )</td>
<td>0.12 (0.03)</td>
<td>4.65 (1.14)</td>
<td>0.91 (0.19)*</td>
<td>0.13 (0.03)*</td>
<td>0.11 (0.02)*</td>
</tr>
<tr>
<td>BDL TZP</td>
<td>( &lt; 0.05 )</td>
<td>0.12 (0.03)</td>
<td>4.87 (1.39)</td>
<td>0.82 (0.21)*</td>
<td>0.14 (0.04)*</td>
<td>0.12 (0.03)*</td>
</tr>
<tr>
<td>BDL IB-367</td>
<td>( &lt; 0.05 )</td>
<td>0.09 (0.03)</td>
<td>0.78 (0.22)</td>
<td>0.18 (0.04)*</td>
<td>0.06 (0.01)*</td>
<td>0.07 (0.02)*</td>
</tr>
</tbody>
</table>

Results are given as mean (SD).

*TZP, tazobactam-piperacillin 120 mg/kg; IB-367 1 mg/kg.

Each measurement refers to:b20 animals;c13 animals;dseven animals;esix animals;f15 animals;g10 animals;hnine animals;i18 animals.

**Figure 1** Antimicrobial activities of an intraperitoneal monodose of 120 mg/kg tazobactam-piperacillin (TZP) and 1 mg/kg IB-367 in sham operated and bile duct ligated (BDL) rats. CFU, colony forming units.
Lethality

LPS induced 60% lethality in BDL placebo treated rats and no lethality in sham operated rats. Eleven of 20 TZP treated rats (55%) with BDL did not survive past 48 hours. With IB-367 treatment, lethality was reduced to two of 20 BDL rats (10%). In addition, among BDL rats, only IB-367 significantly reduced lethality compared with placebo (table 3). Finally, none of the agents showed any toxicity; in fact none of the animals had clinical evidence of drug related adverse effects, such as local signs of inflammation, anorexia, weight loss, vomiting, diarrhoea, fever, or behavioural alterations.

DISCUSSION

Endotoxaemia is strongly implicated in the complications common to jaundiced patients undergoing surgery. Endotoxaemia in biliary obstruction is probably the result of disturbance of the homeostatic gut-liver system. In fact, under the influence of various critical illnesses, such as total biliary obstruction, intestinal mucosal integrity is compromised and barrier function can be lost, permitting the passage of indigenous intestinal bacteria and endotoxins into the circulation and organs which are normally sterile, a term which has been coined bacterial translocation (BT). BT may have profound systemic effects, and may result in secondary infections, such as intra-abdominal abscesses, sepsis, and bacteraemia, as well as endotoxaemia. In addition, endotoxin may increase the permeability of the intestinal mucosa, thereby facilitating BT and endotoxin absorption. In this way the gut can become the source of an ongoing septic state in individuals without a defined infectious focus.

The aim of this study was to investigate the effect of IB-367, a new synthetic protegrin, on endotoxin induced mortality in rats that had undergone BDL. The results of the study showed that LPS administration was followed by mortality only in BDL rats. Specifically, although one week after operation there were significant changes in endotoxin levels between sham and BDL rats only, induction of high endotoxaemia in BDL rats resulted in a 48 hour lethality rate of 60%, 55%, and 10% in placebo, TZP, and IB-367 treated animals.

The amount of endotoxin detected in plasma after E. coli serotype 0111:B4 LPS administration was higher in placebo treated BDL rats. Previous reports have described that endotoxaemia may be the result not only of increased absorption of endotoxins through disrupted gut barrier but also of decreased capacity of mononuclear cells for inactivating endotoxins. In contrast, reduction in endotoxaemia and endotoxin induced mortality among IB-367 treated BDL rats, compared with placebo and TZP treated BDL rats, may be not only the result of direct neutralisation of LPS by binding of the lipid A moiety, inhibiting the biological activity of endotoxin, but also of a reduction in the amount of circulating endotoxin available for binding to Kupffer cells and inducing production of cytokines such as TNF-α. In fact, intraperitoneal administration of IB-367 immediately after injection of LPS would have directly inactivated the endotoxin, thus preventing its systemic effects.

It has been demonstrated that final plasma TNF-α levels are the result of a fine balance involving the whole mononuclear phagocyte system. In jaundiced animals, peritoneal macrophage function was significantly depressed seven days after BDL. Nevertheless, it has been demonstrated that the number of Kupffer cells increases during experimental biliary obstruction and it is known that Kupffer cells are unique in certain respects. In fact, they are strongly activated by bacterial endotoxins and normally in a low grade state of activation because of the small quantities of endotoxins that continually penetrate the intact intestinal mucosa.

The different number and state of activation of Kupffer cells in sham and BDL rats may explain the lethality reported in the last model. We observed that plasma TNF-α levels peaked two hours after exposure to LPS both in jaundiced and sham operated rats but TNF-α secretion in BDL animals exceeded that of sham rats by a factor of approximately 50. These findings indicate that plasma TNF-α, rather than endotoxin levels, are related to lethality.

Finally, the present study confirms that ligation of the bile duct increases the rate of translocation of enteric bacteria, even though this phenomenon did not appear to be directly related to survival, as demonstrated by the high lethality rate in TZP treated BDL rats in spite of the strong antibacterial activity of this compound.

The peculiar ability of antimicrobial peptides, such as IB-367, to inhibit not only bacterial growth but also biological activity of the endotoxin may offer an important opportunity for a new approach in the therapeutic strategies in obstructive jaundice. In fact, they act on Gram negative bacteria by initially binding to their surface polyanionic LPS, followed by self promoted uptake across the outer membrane. By virtue of this affinity for LPS, the cationic peptides operate as antiendotoxin agents and inhibit the production of cytokines such as TNF-α by macrophages stimulated with LPS.

This study has demonstrated that a single intraperitoneal dose of IB-367 produced significant reductions in plasma endotoxin and TNF-α levels compared with levels in the placebo and the TZP treated groups. On the basis of our results, IB-367 may have certain potential advantages. Firstly, the use of single doses of the compound resulted in significant inhibition of bacterial growth compared with that in the placebo groups. In addition, it produced a significant reduction in plasma endotoxin and TNF-α levels compared with those in the TZP treated groups, confirming its double antimicrobial and antiendotoxin activities.

Table 3: Effect of intraperitoneal monodose of 120 mg/kg tazobactam-piperacillin (TZP) and 1 mg/kg IB-367 on mortality in sham operated rats and bile duct ligated (BDL) rats

<table>
<thead>
<tr>
<th>Time (h)</th>
<th>Sham placebo</th>
<th>Sham TZP</th>
<th>Sham IB-367</th>
<th>BDL placebo</th>
<th>BDL TZP</th>
<th>BDL IB-367</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>6</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0.65</td>
<td>0.75</td>
<td>0.9</td>
</tr>
<tr>
<td>12</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0.5</td>
<td>0.65</td>
<td>0.9</td>
</tr>
<tr>
<td>24</td>
<td>1</td>
<td>1</td>
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<td>0.35</td>
<td>0.5</td>
<td>0.9</td>
</tr>
<tr>
<td>36</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0.3</td>
<td>0.45</td>
<td>0.9</td>
</tr>
<tr>
<td>48</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0.3</td>
<td>0.45</td>
<td>0.9</td>
</tr>
</tbody>
</table>
REFERENCES

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