Gastric cancer is the second leading cause of cancer-related death worldwide.1 Notwithstanding the global declining incidence of gastric cancer, mortality is still rising in Asian countries. To date, there is no effective measure to prevent development of gastric cancer. Although *Helicobacter pylori* infection has been identified as the most important causative factor,2 there is little evidence to substantiate the fact that eradication of the bacterium alone can stop the process of gastric carcinogenesis.3,4

Since the observation from the Physician’s Health Study that usage of aspirin may reduce the risk of colorectal cancer, intense interest has been directed towards investigation of the antitumor properties of aspirin and non-steroidal anti-inflammatory drugs (NSAIDs). There are at least 12 published observational studies showing the protective effects of NSAIDs against colorectal cancer. More recently, studies in colon cancer show that induction of cyclooxygenase 2 (COX-2) is associated with inhibition of apoptosis, increase in angiogenesis, and metastatic potential.5 Celecoxib, a COX-2 inhibitor, has been shown to reduce polyp formation in a cohort of patients with familial adenomatous polyposis syndrome.6

COX-2 expression is upregulated in *H pylori* induced mucosal inflammation.6 It is frequently expressed in gastric cancer7,8 as well as in premalignant gastric lesions.9 Inhibition of COX-2 in vitro results in growth inhibition of gastric cancer cells.10 Furthermore, the use of COX-2 inhibitors has been shown to suppress the growth of gastric cancer xenografts in nude mice.11 Unlike colorectal cancers, however, there are a lack of animal and human data demonstrating the effectiveness of COX-2 inhibition and NSAIDs in the prevention of gastric cancer.

In this study, we evaluated the use of celecoxib and indomethacin in the prevention of *N*-methyl-*N*-nitro-*N*-nitrosoguanidine (MNNG) induced gastric cancer in rats.

**Background:** Overexpression of cyclooxygenase 2 (COX-2) is frequently detected in gastric cancer and is believed to play a crucial role in gastric carcinogenesis. Aims: We examined the chemopreventive effect of a COX-2 inhibitor in an animal model of stomach carcinogenesis.

**Methods:** Eighty-six male Wistar rats were divided into six different treatment groups: group A, water alone (n = 16); group B, *N*-methyl-*N*-nitro-*N*-nitrosoguanidine (MNNG 100 μg/ml) (n = 16); group C, indomethacin (3 mg/kg/day) (n = 16); group D, celecoxib (5 mg/kg/day) (n = 17); group E, celecoxib (10 mg/kg/day) (n = 16); and group F, celecoxib (20 mg/kg/day) (n = 16). Group B-E animals were treated with 10% sodium chloride (in the initial six weeks) and MNNG in drinking water to induce adenocarcinoma in the stomach. All animals received treatment for 40 weeks, and were sacrificed after death or at 48 weeks. Gastric neoplasm was evaluated by histology.

**Results:** The incidences of gastric cancer were 0% in group A, 75% in group B, 68.8% in group C, 70.6% in group D, 18.8% in group E, and 31.3% in group F (p = 0.004). Compared with MNNG controls, treatment with celecoxib 10 mg/kg/day also showed lower tumour multiplicity (0.19 (0.40) vs. 1.00 (0.73); p = 0.004) and lower mean tumour volume (2.4 ± 2805 mm³; p = 0.02). Although tumours had significantly higher COX-2 expression than their adjacent normal tissues (p < 0.02), there was no significant difference in COX-2 levels among tumours in the different treatment groups. The lowest tumour prostaglandin E2 level was found in the indomethacin treated group, suggesting that the chemopreventive effect of celecoxib may be mediated by a COX independent pathway.

**Conclusion:** While treatment with indomethacin had no significant effect on tumour development, treatment with celecoxib reduced gastric cancer incidence and growth in rats.

**MATERIAL AND METHODS**

**Animals**

Administration of MNNG in drinking water is a well established animal model for the study of the differentiated type of human stomach cancer.12 Eighty-six four week old grade 2 male Wistar rats (approximately 50 g in weight) were obtained from the Laboratory Animal Centre of the Sun Yat-Sen University. Rats were kept in metal cages at 21°C, humidity 50%, with a 12 hour light-dark cycle. Rats had free access to regular chow pellets and drinking water. There was one week of acclimatisation prior to the initiation of this experiment. The study protocol was approved by the animal ethics committee of the Sun Yat-Sen University.

**Chemicals**

MNNG (Fluka, Germany) solution was prepared three times per week with distilled water at a concentration of 100 μg/ml. It was protected from light and given ad libitum to animals in their drinking water. In addition to MNNG, all animals were given 1 ml of 10% sodium chloride weekly by oral gavage in the initial six weeks to enhance gastric cancer development.17

**Study design**

Rats were allocated to one of six different groups (groups A–F). Group A was a control group whereas groups B–F were treated with MNNG. In addition, they were given water (control, group B), indomethacin (3 mg/kg/day, group C), or celecoxib (Pfizer Pharmaceuticals, New York) at 5 mg/kg/day

**Abbreviations:** COX-2, cyclooxygenase 2; MNNG, *N*-methyl-*N*-nitro-*N*-nitrosoguanidine; NSAIDs, non-steroidal anti-inflammatory drugs; PCR, polymerase chain reaction; PGE2, prostaglandin E2; NFκB nuclear factor κB
Histopathology
For histological examination, the stomach was fixed in 10% neutral buffered formalin. Paraffin embedded sections (5 μm) were cut and stained with haematoxylin and eosin for histological examination by a pathologist who was unaware of the treatment assignments. Adenocarcinoma, as defined by the presence of atypical glands that locally invaded the submucosa, muscularis propria, or serosa, was recorded.19

RNA extraction and quantitative PCR
Gastric tissue specimens were homogenised with an ultrasound homogeniser. Total RNA was extracted by RNA Tri Reagents (CINNA/MRC; Cincinnati, Ohio, USA). Total RNA (1 μg) was reverse transcribed into cDNA using dNTPs (1 mM), 5X reverse transcription buffer (500 mM Tris HCl, pH 8.3, 250 mM KCl, 50 mM MgCl₂, and 50 mM DTT), 16 units RNasin, and 2.5 units of AMV reverse transcriptase (GibcoBRL, Life Technologies Gaithersburg, Maryland, USA). Real time quantitative polymerase chain reaction (PCR) was performed on a ABI PRISM 7000 sequence detection system using SybBgreen, PCR mastermix (Perkin Elmer, Branchburgh, New Jersey), and primers. Primer sequences were designed from the Genbank as follows: COX-2 (L25925) (forward, 1408–1435) 5'-ACAGGAGAGAAAGAAATGGCT-3', (reverse, 1598–1573) 5'-CAGTATTGAGGAGA-CACAGATGGGATT-3'; and β-actin (NM-031144) (forward, 476–500), 5'-TCACCCACATCTGGCCCATCTATGA-3', (reverse, 633–610) 5'-GTCAGCGACCATTCCCTCCACGC-3'. A 24 μl reaction mix was aliquoted with 1 μl/rePLICATE of cDNA. A DNase treated free template control (containing water) was included and each sample was added in duplicate. Reaction tubes were scaled with optical caps, and the PCR reaction was run at 50°C for two minutes, 95°C for 10 minutes, followed by 40 cycles at 96°C for 45 seconds, 60°C for 45 seconds, and 72°C for one minute. The specificity of the PCR products was characterised by melting curve analysis and followed by gel electrophoresis. Quantification was determined by the threshold cycle. Actin was used as a housekeeping gene to normalise mRNA levels and compared against mRNA expression levels in normal control stomach. 

PGE₂ assay
Prostaglandin E₂ (PGE₂) levels were measured in snap frozen tissue specimens using an ELISA based assay (Amersham Pharmacia Biotech), and the column was washed with distilled water and hexane. PGE₂ was eluted with two 0.75 ml volumes of ethyl acetate. This fraction was evaporated to dryness under nitrogen and stored at −80°C. Samples were resuspended in 1 ml of buffer and assayed in 96 well plates. The quantity of PGE₂ in supernatants was determined using ELISA.

Statistics
Body weight, tumour incidence (percentage of animals with tumour development), tumour multiplicity (mean number of tumours per animal), mean tumour volume (mean volume of tumour in tumour bearing rats), COX-2, and PGE₂ levels were compared among animals fed MNNG control alone, indomethacin, and celecoxib. Parametric data were analysed by ANOVA with Bonferroni's multiple comparison; non-parametric data were computed by χ² test or Fisher's exact test with Bonferroni's correction. A p value of <0.05 was considered to be statistically significant.

RESULTS
General observation
Body weights of group A control animals were higher than those of the other groups in the early phase of the study (fig 1). However, there was no significant difference in body weight among other treatment groups during the whole study period. There were in total 26 deaths during the study period: none in group A, six in group B, six in group C, five in group D, six in group E, and three in group F. The causes of death are listed in table 1. Most animals died from gastric (n = 14) and small bowel (n = 8) cancer. Two animals died from intestinal haemorrhage after receiving the high dose (20 mg/kg/day) of celecoxib. Two animals died from non-digestive tract diseases.

Tumour incidence
Table 2 summarises the incidences of MNNG induced gastric tumours in the six treatment groups. Seventy five per cent of rats treated with MNNG developed gastric cancer at the end of this study whereas none of the control rats in group A had a gastric tumour. There was a significant difference in tumour incidences among different treatment groups (p = 0.002). Rats treated with celecoxib 10 mg/kg/day (group E) had the lowest tumour incidence (18.8%) which was significantly lower than the MNNG group (75.0%; p = 0.004). The tumour incidence of group F rats (celecoxib 20 mg/kg/day) also tended to be lower than the MNNG control (75.0%) (p = 0.062). The absolute risk reductions of gastric cancer in animals treated with celecoxib 10 mg/kg/day and 20 mg/kg/day were 56.3% (95% confidence interval (CI) 16.7–80.4%) and 43.8% (95% CI 3.9–71.8%), respectively. On the other hand, the numbers of gastric tumours were comparable between the other treatment groups (indomethacin or celecoxib at 5 mg/kg/day) and the MNNG control group. All gastric tumours were confirmed to be adenocarcinomas (fig 2) and the majority (90.7%) were high grade tumours.

Moreover, premalignant gastric lesions such as dysplasia were frequently detected in MNNG treated rats. Of the 10 remaining viable rats in group B, nine had dysplasia on histological examination of the gastric mucosa. In contrast, only four of the 10 remaining rats in group E (celecoxib 10 mg/kg/day) had gastric dysplasia (p = 0.23). The frequencies of gastric dysplasia in rats treated with indomethacin and other doses of celecoxib were comparable with group B.

Tumour multiplicity
In addition to tumour incidence, there was a significant difference in tumour multiplicity, or number of cancers per rat, among the different treatment groups (p = 0.001, table 2).
Celecoxib prevents gastric cancer in rats

Fig. 1. Body weight of animals in the different treatment groups. The body weight of group A control rats was higher than in the N-methyl-N'-nitro-N-nitrosoguaniidine (MNNG) treated groups in the initial phase of the experiments. However, there was no significant difference in body weight among all MNNG treated groups (groups B–F).

Compared with the MNNG group, rats fed celecoxib 10 mg/kg/day or 20 mg/kg/day had significantly lower tumour multiplicities compared with animals treated with MNNG alone (0.2 (0.4) × 1.0 (0.7), p = 0.004 and 0.3 (0.5) × 1.0 (0.7), p = 0.025). However, treatment with indomethacin (group C) and low dose celecoxib (5 mg/kg/day, group D) did not have any apparent suppressive effect on tumour multiplicity.

Tumour volume

Mean tumour volume was significantly different among the treatment groups (p = 0.009). Specifically, rats treated with celecoxib had a markedly reduced tumour volume compared with the MNNG control group (group B). Mean tumour volumes were significantly lower in animals treated with celecoxib 5 mg/kg/day (group D) (188.5 (377.8) mm³; p = 0.036), 10 mg/kg/day (group E) (2.4 (7.0) mm³; p = 0.022) and 20 mg/kg/day (group F) (38.9 (110.5) mm³; p = 0.025) compared with those treated with MNNG alone (group B) (2805 (5540.1) mm³). In contrast, mean tumour volume in indomethacin treated animals (group C) was only marginally lower than the MNNG control group (359 (859.8) mm³; p = 0.07).

Non-gastric tumours

There were eight animals that developed small bowel adenocarcinoma, three in the indomethacin treated group (group C) and five in the celecoxib treated group (group E). One animal in group E also developed lung cancer. Overall, there was no significant difference in the number of small bowel and lung tumours with different treatment allocations.

COX-2 and PGE₂ levels

COX-2 was expressed at low levels in the stomach of control rats (0.53 (0.11)) (fig 3). In contrast, COX-2 was upregulated in tumours. Gastric tumours had higher COX-2 expression than their adjacent normal tissues in all treatment groups (p<0.02). Treatment with celecoxib or indomethacin did not reduce tumour COX-2 levels but COX-2 was significantly lower in adjacent normal tissues of celecoxib or indomethacin treated groups (p<0.01).

In addition to induction of COX-2, PGE₂ levels were increased in tumours (fig 4). Gastric tumours in all treatment groups tended to have higher PGE₂ levels than their adjacent normal tissues but a significant difference was only observed in the low dose celecoxib (5 mg/kg/day) group (p = 0.015). Treatment with indomethacin (3 mg/kg/day) or high doses of celecoxib (>10 mg/kg/day) were associated with mildly reduced tumour PGE₂ levels, but the difference did not reach statistical significance. Moreover, there was no significant difference in PGE₂ levels of normal tissues among the different treatment groups.

DISCUSSION

In this study, we determined the role of COX-2 inhibition in the prevention of sodium chloride enhanced gastric carcinogenesis induced by MNNG in Wistar rats. MNNG induced gastric cancer is a well established animal model of stomach carcinogenesis. The mutagen, when given in drinking water, induces intestinal metaplasia and adenocarcinoma in the pyloric mucosa of Wistar rats. The histology of this induced gastric malignancy resembles the differentiated type of stomach cancer in humans. To enhance the carcinogenic effects of MNNG, highly concentrated sodium chloride solution was given to these animals in the initial six weeks. In the present study, 75% of MNNG treated animals developed gastric cancer at the end of 48 weeks, confirming that this is a highly successful model of gastric tumorigenesis.

Although the exact mechanism underlying MNNG induced gastric cancer remains poorly understood, previous studies showed that the genetic makeup of the animals may play a role. For example, ACI/N rats are highly susceptible to MNNG induced stomach carcinogenesis but BUF/Nac rats are relatively resistant. Recently, COX-2 and Bcl-2 were found to be coexpressed in the glandular corpus epithelium of rats treated with MNNG. This upregulated expression is associated with cell proliferation, atrophy, and intestinal metaplasia of the stomach. It is therefore logical to anticipate that treatment with a COX-2 inhibitor may have an antiproliferative and hence chemopreventive effect on MNNG induced gastric cancer.

The results of this study showed, for the first time, that both the incidence and multiplicity of MNNG induced gastric cancer can be significantly reduced in rats treated with celecoxib. The chemopreventive effect of celecoxib was demonstrated when a moderate dose (10 mg/kg/day) was given to these animals. With the use of celecoxib 10 mg/kg/day, there was an approximate 56% reduction in tumour incidence, 80% reduction in tumour multiplicity, and 1169-fold reduction in tumour volume. This remarkable degree of tumour suppression by celecoxib is comparable with that reported in the azoxymethane induced colon cancer model in rats. Moreover, it exceeds that previously reported in MNNG induced gastric cancer by other agents, such as

Table 1. Tumour incidences and causes of death in the different groups of study animals

<table>
<thead>
<tr>
<th>Group</th>
<th>No of rats</th>
<th>Treatment</th>
<th>Causes of death (No of animals)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>5</td>
<td>Control</td>
<td>Gastric cancer (6)</td>
</tr>
<tr>
<td>B</td>
<td>16</td>
<td>MNNG alone</td>
<td>Gastric cancer (2), small bowel cancer (3), unknown (1)</td>
</tr>
<tr>
<td>C</td>
<td>16</td>
<td>MNNG+indomethacin (3 mg/kg/day)</td>
<td>Gastric cancer (5)</td>
</tr>
<tr>
<td>D</td>
<td>17</td>
<td>MNNG+celecoxib (5 mg/kg/day)</td>
<td>Small bowel cancer (5), lung cancer (1)</td>
</tr>
<tr>
<td>E</td>
<td>16</td>
<td>MNNG+celecoxib (10 mg/kg/day)</td>
<td>Gastric cancer (11), intestinal haemorrhage (2)</td>
</tr>
<tr>
<td>F</td>
<td>16</td>
<td>MNNG+celecoxib (20 mg/kg/day)</td>
<td>Gastric cancer (11), intestinal haemorrhage (2)</td>
</tr>
</tbody>
</table>

MNNG, N-methyl-N’-nitro-N-nitrosoguanidine.
differentiated gastric adenocarcinoma in the stomach of a Wistar rat. (B) Haematoxylin-eosin staining of well differentiated gastric adenocarcinoma in the stomach (×20).

Table 2  Tumour incidences and multiplicity in the different treatment groups

<table>
<thead>
<tr>
<th>Group</th>
<th>Treatment</th>
<th>No of deaths (%)</th>
<th>No of rats with gastric tumours (%) (incidence)*</th>
<th>No of gastric cancers per rat (SD) (multiplicity)†</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>Control</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>B</td>
<td>MNNG alone</td>
<td>6 (37.5)</td>
<td>12 (75.0)</td>
<td>1.0 (0.7)</td>
</tr>
<tr>
<td>C</td>
<td>MNNG+indomethacin</td>
<td>6 (37.5)</td>
<td>11 (68.8)</td>
<td>0.8 (0.80)</td>
</tr>
<tr>
<td>D</td>
<td>MNNG+celecoxib 5 mg/kg/day</td>
<td>5 (29.4)</td>
<td>12 (70.6)</td>
<td>0.8 (0.6)</td>
</tr>
<tr>
<td>E</td>
<td>MNNG+celecoxib 10 mg/kg/day</td>
<td>6 (37.5)</td>
<td>3 (18.8)</td>
<td>0.2 (0.4)</td>
</tr>
<tr>
<td>F</td>
<td>MNNG+celecoxib 20 mg/kg/day</td>
<td>3 (18.8)</td>
<td>5 (31.3)</td>
<td>0.3 (0.5)</td>
</tr>
</tbody>
</table>

* p = 0.002 (ANOVA): C versus B, p = 1.00; D versus B, p = 1.00; E versus B, p = 0.004; F versus B, p = 0.052.
† p = 0.001 (ANOVA): C versus B, p = 1.00; D versus B, p = 1.00; E versus B, p = 0.004; F versus B, p = 0.025.

Figure 2  Macroscopic and microscopic appearance of N-methyl-N'-nitro-N-nitrosoguanidine (MNNG) induced gastric tumour in a rat. (A) Macroscopic view of MNNG induced tumour formation in the distal stomach of a Wistar rat. (B) Haematoxylin-eosin staining of well differentiated gastric adenocarcinoma in the stomach (×20).

Figure 3  Cyclooxygenase 2 (COX-2) mRNA expression levels of tumours and adjacent normal tissues in the different treatment groups. COX-2 mRNA expression levels were determined by quantitative reverse transcription-polymerase chain reaction. Mean (SEM) values are shown. There was upregulation of COX-2 in all tumours compared with MNNG controls. The reason for these discrepancies between indomethacin and celecoxib is unclear. One plausible explanation may be related to the dose of indomethacin used in this study. Our selection of this dose was based on two facts. Firstly, the recommended dose of indomethacin in humans is 1–3 mg/kg/day. Secondly, previous animal studies demonstrated inhibitory effects on the formation of aberrant crypt foci in the colons of dimethyl hydrazine treated rats using a dose of 2 mg/kg/day. As shown in figure 4, tumour PGE2 levels in the indomethacin

celecoxib used in this study may be associated with more toxicity, such as intestinal haemorrhage. Moreover, the COX-2 selectivity of celecoxib may be lost at high doses, resulting in more COX-1 inhibition. Based on our data with high dose celecoxib and indomethacin, concurrent COX-1 inhibition may have a paradoxical effect on chemoprevention. It remains undetermined whether concurrent COX-1 inhibition has a promotional effect on tumour development.
Celecoxib prevents gastric cancer in rats

The Mongolian gerbil was recently found to be a good animal model to study H pylori associated gastric carcinogenesis.\(^\text{18}\) Moreover, emerging data show that COX-2 is upregulated in the gerbil stomach after H pylori infection.\(^\text{19}\) It will be interesting to characterise the role of COX-2 inhibition in the chemoprevention of gastric cancer in this gerbil model. Another issue that is worth further study is the role of celecoxib in the therapy of established gastric cancer, as this drug was introduced at the same time as the carcinogen in this study. The exact therapeutic role of celecoxib against established cancer remains unknown and a study that introduces celecoxib at different time points may be helpful in clarifying this point. Moreover, this type of study may help address the important question of the optimal time of intervention if it is found that celecoxib only prevents gastric cancer development but fails in the treatment of established cancer.

In summary, our study showed that treatment with celecoxib, a specific COX-2 inhibitor, suppressed MNNG induced gastric cancer in rats. This finding lends further support to the use of COX-2 inhibitors in the chemoprevention of gastric cancer. Whether this result can be translated into clinical benefit requires further confirmation in human clinical studies.

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REFERENCES


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