Dietary fructo-oligosaccharides and inulin decrease resistance of rats to salmonella: protective role of calcium

S J M Ten Bruggencate, I M J Bovee-Oudenhoven, M L G Lettink-Wissink, M B Katan, R Van der Meer

Gastrointestinal infections induced by food borne pathogens are a major clinical problem. Survival of food borne pathogens within the intestinal tract can potentially be modulated by dietary prebiotics. Prebiotics, which include fructo-oligosaccharides (FOS) and inulin, are defined as non-digestible food ingredients that beneficially affect the host by selectively stimulating the growth and/or activity of one or a limited number of bacteria in the colon that can improve the health of the host. Stimulation of the endogenous microflora by dietary prebiotics may inhibit colonisation of intestinal pathogens by production of organic acids and by competing for nutrients and mucosal adhesion sites. However, we showed previously that FOS decreases the resistance towards the invasive food borne pathogen Salmonella enterica serovar Enteritidis. Rapid fermentation of FOS by the intestinal microflora may lead to high luminal concentrations of organic acids. These organic acids may induce damage to the mucosal barrier and hence decrease resistance to intestinal pathogens.

Both inulin and FOS are composed of linear chains of fructose units, linked by \(\beta(2\rightarrow1)\) bonds and often terminated by a glucose unit. The number of fructosyl moieties ranges from 2 to 60 for inulin and from 2 to 7 for FOS. In vitro fermentation experiments revealed that molecules with a degree of polymerisation (DP) >10 are fermented, on average, half as quickly as molecules with a DP of <10. Thus the possible adverse consequences of rapid fermentation might be more pronounced with FOS than with inulin.

The consequences of excessive fermentation can potentially be inhibited by dietary calcium phosphate. Calcium forms an insoluble complex with phosphate in the upper small intestine. This complex increases the buffering capacity throughout the intestinal lumen. Thus it can be speculated that dietary calcium phosphate counteracts the effects of rapid fermentation of inulin and FOS.

The aim of the present study was to compare the effects of FOS and inulin on the resistance of rats to the invasive pathogen Salmonella enterica serovar Enteritidis. In addition, we determined whether dietary calcium phosphate can counteract the effects of rapid fermentation.

METHODS

Diets, infection, and dissection of the animals

The animal welfare committee of Wageningen University and Research Centre (Wageningen, the Netherlands) approved the experimental protocol. Specific pathogen free male Wistar rats (WU, Harlan, Horst, the Netherlands), eight weeks old, with a mean body weight of 276 (2) g, were housed individually in metabolic cages. All animals were kept in a temperature (22–24°C) and humidity (50–60%) controlled environment. The mean food intake was 266 kJ/day of a purified diet. Restricted food intake was necessary to prevent differences in food consumption and hence differences in vitamin and mineral (especially calcium) intake. The control diet contained (per kg) 200 g acid casein, 49 g mineral mix (without calcium), 552 g glucose, 160 g palm oil, 40 g corn oil, 20 g cellulose, 49 g mineral mix (without calcium), and 14 g vitamin mix according to AIN93 recommendation. The experimental diets were supplemented with 60 g/kg cellulose (Arbocel type)

Abbreviations: CFU, colony forming units; DP, degree of polymerisation; FOS, fructo-oligosaccharides; ICP-AES, inductively coupled plasma-atomic emission spectrophotometry; NOx, sum of nitrate and nitrite; PCR, polymerase chain reaction
The supernatants were diluted with 0.5 g/l CsCl and analysed.

Temperature and centrifuged for two minutes at 14,000 rpm. One hour at room temperature and centrifuged for one hour at 37°C, and subsequently centrifuged for one hour at 14,000 g (Hettich, Micro-rapid 1306, Tuttingen, Germany). Cytotoxicity of faecal water was determined with an erythocyte assay, as previously described, and validated earlier with intestinal epithelial cells. Incubations were of physiological ionic strength (300 mosmol/l) and buffered at pH 7.0 (final 100 mosmol/l 3-N-morpholino-propanesulfonic acid; Sigma) to prevent acid induced haemolysis.

Microbiological analyses of faeces

Faeces were quantitatively collected and pooled per animal for five days before salmonella infection. Faeces were freeze dried and subsequently ground to obtain homogeneous powdered samples. Real time quantitative polymerase chain reaction (PCR), targeting a 110 bp transaldolase gene sequence, was used to specifically quantify bifidobacteria in faecal samples, as described and validated previously. The cycle threshold values generated by real time PCR from DNA extracts of dilutions of caecal contents in saline were plated on Modified Brilliant Green Agar (Oxoid) to suppress swarming bacteria such as proteus species. The detection limit of this method was 10² CFU/g faecal wet weight.

To determine salmonella numbers in caecal contents seven days after infection, the caecum was excised and caecal contents were removed. The pH of the caecal contents was measured at 37°C and caecal tissue was weighed. Tenfold dilutions of caecal contents in saline were plated on Modified Brilliant Green Agar (Oxoid) and incubated aerobically overnight at 37°C. Lactobacilli were quantified by plating 10°fold dilutions in saline on Levine EMB Agar (Difco Laboratories, Detroit, Michigan, USA) and incubating aerobically overnight at 37°C. Lactobacilli were quantified by plating on Rogosa Agar (Oxoid, Basingstoke, UK) and incubating in anaerobic jars (MART Microbiology, Lichtenvoorde, the Netherlands) at 37°C for three days. The detection limit of this method was 10² CFU/g faecal wet weight.

Biochemical analyses of faeces

Total faecal lactic acid was measured using a colorimetric enzymatic kit (Boehringer Mannheim, Germany), as described previously. To measure calcium, faeces were treated with 50 g/l trichloroacetic acid for one h at room temperature and centrifuged for two minutes at 14,000 g. The supernatants were diluted with 0.5 g/l CsCl and analysed by ICP-AES (Varian). To measure phosphate, dry ashed faeces was destructed (15 minutes at 180°C) with a perchoric acid (70%)/hydrogen peroxide (30%) mixture (5:1 vol/ vol, total volume 600 μl). Subsequently, samples were centrifuged for two minutes at 14,000 g. The supernatants were diluted with 0.5 g/l CsCl and analysed by ICP-AES (Varian).

Murin was extracted from freeze dried faeces and quantified fluorometrically, as described previously. Interfering oligosaccharides of dietary origin were removed by filtration (data not shown). Standard solutions of N-acetylgalactosamine (Sigma, St Louis, Missouri, USA) were used to calculate the amount of oligosaccharide side chains liberated from murins. Therefore, faecal murins were expressed as μmol oligosaccharide equivalents.

Analyses of faecal water

Fresh faeces were freeze dried and reconstituted with double distilled water to obtain a physiological osmolarity of 30 mosmol/l. Samples were mixed, incubated for one hour at 37°C, and subsequently centrifuged for one hour at 14,000 g (Hettich, Micro-rapid 1306, Tuttingen, Germany). Cytotoxicity of faecal water was determined with an erythocyte assay, as previously described, and validated earlier with intestinal epithelial cells. Incubations were of physiological ionic strength (300 mosmol/l) and buffered at pH 7.0 (final 100 mosmol/l 3-N-morpholino-propanesulfonic acid; Sigma) to prevent acid induced haemolysis.

Analyses of urine samples

Complete 24 hour urine samples were collected starting one day before infection until six days after infection. Oxytetracycline (1 mg; Sigma) was added to the urine collection vessels of the metabolic cages to prevent bacterial deterioration. The concentration of NOx (sum of nitrate and nitrite) was determined using a colormetric enzymatic kit (No 1746081; Roche Diagnostics, Mannheim, Germany). Briefly, urinary nitrate is reduced to nitrite by nitrate reductase. Subsequently, nitrite reacts with sulphanilamide and N-(1-naphthyl)-ethylenediamine dihydrochloride, resulting in a red diazo dye, which was measured spectrophotometrically at 540 nm.

Statistical analysis

Results are expressed as means (SEM). A commercially available package (Statistica 6.1; StatSoft Inc., Tulsa, Oklahoma, USA) was used for all statistics. We tested for differences between the low calcium/control, FOS, and inulin groups. In addition, we tested for differences between the high calcium groups and their low calcium counterparts. In case of normally distributed data, differences between means were tested for significance using a one way ANOVA, followed by Fisher's protected least significant difference test (two sided). Differences were considered statistically significant when p<0.05. When data were not normally distributed, differences between means were tested for significance using a Kruskall-Wallis ANOVA, followed by the non-parametric Mann-Whitney U test (two sided). Bonferroni correction was made for the number of comparisons (n = 6). Differences were considered statistically significant when p<0.05.

RESULTS

Animal growth and food intake

One animal in the high calcium/FOS group was excluded from all study results because of oropharyngeal reflux of the salmonella suspension, resulting in pneumonia. Before infection, there were no differences in animal growth between the diet groups. Mean animal growth was 1.26
output of these minerals was substantially higher in the high calcium groups.

In addition, low calcium/inulin and FOS increased the cytotoxicity of faecal water (fig 1) and stimulated daily faecal mucin excretion in the low calcium groups before infection (fig 2). Both cytotoxicity and faecal mucin excretion were inhibited by dietary calcium.

Colonisation and translocation of salmonella

Both inulin and FOS increased salmonella numbers in caecal contents six days after infection (fig 3). Dietary calcium did not affect colonisation of salmonella in the caecum. Major differences were observed in the effects of low calcium/inulin and FOS on translocation of salmonella, as measured by the infection induced urinary NO\textsubscript{x} excretion in time (fig 4). Both low calcium/inulin and FOS increased urinary NO\textsubscript{x} excretion in time. At day 6 after infection, urinary NO\textsubscript{x} excretion was threefold higher in the low calcium/inulin group and fourfold higher in the low calcium/FOS group compared with the low calcium/control group. Dietary calcium inhibited translocation of salmonella to a large extent in the FOS group. Total infection induced urinary NO\textsubscript{x} excretion (corrected for baseline output) for the low calcium groups was 122 (12) mmol/6 days in the control group, 290 (59) mmol/6 days in the inulin group, and 448 (66) mmol/6 days in the FOS group. Total NO\textsubscript{x} excretion for the high calcium counterparts was 104 (13) mmol/6 days in the control group, 174

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### Table 1

<table>
<thead>
<tr>
<th></th>
<th>Growth (g/day)</th>
<th>Caecal pH</th>
<th>Caecal weight (g)</th>
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<tbody>
<tr>
<td><strong>Low calcium</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>1.4 (0.2)</td>
<td>7.2 (0.2)</td>
<td>1.2 (0.1)</td>
</tr>
<tr>
<td>Inulin</td>
<td>0.5 (0.1)</td>
<td>6.0 (0.1)*</td>
<td>1.9 (0.1)*</td>
</tr>
<tr>
<td>FOS</td>
<td>-0.6 (0.5)*</td>
<td>6.5 (0.3)*</td>
<td>2.1 (0.1)*</td>
</tr>
<tr>
<td><strong>High calcium</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>1.4 (0.2)</td>
<td>7.5 (0.2)</td>
<td>1.2 (0.1)</td>
</tr>
<tr>
<td>Inulin</td>
<td>0.7 (0.5)</td>
<td>7.0 (0.3)*</td>
<td>1.8 (0.1)</td>
</tr>
<tr>
<td>FOS</td>
<td>0.6 (0.3)*</td>
<td>6.8 (0.2)</td>
<td>1.7 (0.1)</td>
</tr>
</tbody>
</table>

†Results are expressed as mean (SEM) (n = 7 in the high calcium FOS group and n = 8 in the other diet groups).

*Significantly different from the low calcium/control group (p < 0.05).

††Significantly different from its low calcium counterpart (p < 0.05).

### Table 2

<table>
<thead>
<tr>
<th></th>
<th>Bifidobacteria (No log 10/g wet faeces)</th>
<th>Lactobacilli</th>
<th>Enterobacteria</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Low calcium</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>9.5 (0.1)</td>
<td>5.1 (0.4)</td>
<td>7.6 (0.1)</td>
</tr>
<tr>
<td>Inulin</td>
<td>9.8 (0.1)</td>
<td>6.6 (0.4)*</td>
<td>8.8 (0.1)*</td>
</tr>
<tr>
<td>FOS</td>
<td>9.9 (0.2)</td>
<td>7.1 (0.4)*</td>
<td>9.0 (0.1)*</td>
</tr>
<tr>
<td><strong>High calcium</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>8.9 (0.1)*</td>
<td>6.7 (0.2)*</td>
<td>7.4 (0.2)</td>
</tr>
<tr>
<td>Inulin</td>
<td>10.1 (0.1)</td>
<td>7.5 (0.5)*</td>
<td>7.7 (0.3)*</td>
</tr>
<tr>
<td>FOS</td>
<td>10.2 (0.1)</td>
<td>7.9 (0.2)</td>
<td>7.7 (0.2)*</td>
</tr>
</tbody>
</table>

††Bifidobacteria were specifically quantified by real time quantitative polymerase chain reaction. Lactobacilli and enterobacteria were quantified by standard culturing on appropriate agar plates.

Results are expressed as mean (SEM) (n = 7 in the high calcium FOS group and n = 8 in the other diet groups).

*Significantly different from the low calcium/control group (p < 0.05); ††significantly different from its low calcium counterpart (p < 0.05).

CFU, colony forming units.

### Table 3

<table>
<thead>
<tr>
<th></th>
<th>Lactate (μmol/day)</th>
<th>Calcium (μmol/day)</th>
<th>Phosphate (μmol/day)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Low calcium</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>4.5 (1.3)</td>
<td>109 (13)</td>
<td>113 (8)</td>
</tr>
<tr>
<td>Inulin</td>
<td>6.9 (1.5)*</td>
<td>28 (3)*</td>
<td>64 (6)*</td>
</tr>
<tr>
<td>FOS</td>
<td>6.4 (2.1)†</td>
<td>30 (6)*</td>
<td>60 (5)*</td>
</tr>
<tr>
<td><strong>High calcium</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>9.4 (2.6)†</td>
<td>766 (30)*</td>
<td>468 (40)†</td>
</tr>
<tr>
<td>Inulin</td>
<td>21.3 (4.3)†</td>
<td>619 (39)*</td>
<td>374 (29)†</td>
</tr>
<tr>
<td>FOS</td>
<td>26.9 (7.9)†</td>
<td>615 (53)*</td>
<td>367 (42)†</td>
</tr>
</tbody>
</table>

†Lactic acid was measured using a calorimetric enzymatic kit. Calcium and phosphate were analysed by ICP-AES. Results are expressed as mean (SEM) (n = 7 in the high calcium FOS group and n = 8 in the other diet groups).

*Significantly different from the low calcium/control group (p < 0.05).

††Significantly different from its low calcium counterpart (p < 0.05).
DISCUSSION

This study showed that inulin and FOS significantly stimulated intestinal colonisation and translocation of salmonella to extraintestinal sites. These adverse effects were largely induced by dietary calcium phosphate.

The general opinion is that inulin and FOS may increase resistance to intestinal infections by selectively stimulating the growth and/or activity of one or a limited number of bacteria in the colon. Indeed, previous studies showed that inulin and FOS stimulated intestinal bifidobacteria and lactobacilli in rats and humans. However, in the present study, no effect of inulin or FOS on faecal bifidobacteria was observed. An explanation could be that when initial bifidobacteria counts are high to start with, a further increase in their numbers might not be demonstrable. Although lactobacilli were stimulated in the present study by inulin and FOS, our results, as well as those of others, do not support the concept of selective stimulation because intestinal enterobacteria, which are associated with gut derived septiccaemia, were also stimulated. The endogenous microflora is known to be important for host resistance. However, the effects of inulin and FOS on resistance to intestinal bacterial infections have rarely been investigated in vivo studies. The few previously published studies showed inconsistent effects of oligosaccharides on colonisation of salmonella. In the present study, inulin and FOS increased salmonella colonisation in the caecum, despite stimulation of the intestinal microflora. These results do not confirm the results obtained in a previous study of our group. However, there is one important difference: in the previous study, salmonella was administered to animals in a fed state while in the present study the salmonella suspension was given after an overnight fast (that is, on an acidic empty stomach). This may have provoked an acid tolerance response in salmonella which may have increased its virulence and subsequent survival within the caecum and colon.

Apart from the increase in colonisation, both prebiotics markedly stimulated salmonella translocation to extraintestinal sites, as measured by urinary excretion of NOx metabolites. Urinary NOx is a sensitive and quantitative biomarker of intestinal bacterial translocation which correlates with organ cultures and severity of systemic infectious diseases in rats and humans. The increased colonisation and translocation of salmonella in the present study, concomitant with decreased animal growth after infection, indicates that the infection was worse in the low calcium/inulin and FOS groups. These data confirm the results of our previous studies in which FOS increased translocation of salmonella.

Why do inulin and FOS increase translocation of salmonella? Fermentation of both inulin and FOS results in the production of organic acids in the distal gut, indicated by the high faecal lactate concentration and the low caecal pH. Accumulation of organic acids and other fermentation metabolites may lead to irritation and impairment of the mucosal barrier. The intestinal mucosa responds to these irritating components by increasing mucus excretion. Indeed, inulin and FOS increased faecal mucin excretion in the present study. In addition, previous studies showed that fermentable fibres increase faecal mucin excretion. Thus rapid fermentation of FOS and inulin may impair the mucosal barrier in the distal gut. Translocation of salmonella is believed to occur through ileal Peyer’s patches but this issue is still debated. Impairment of the barrier by rapid fermentation of inulin and FOS might expand the possibilities for salmonella to translocate in the distal gut.
phosphate complex precipitates cytotoxic components in the intestinal lumen, which reduces epithelial cell damage and increases resistance towards salmonella translocation. Indeed, dietary calcium inhibited the cytotoxicity of faecal water, induced by inulin and FOS. The cytotoxicity assay was performed in a buffer of neutral pH, precluding simple acid induced lysis. Thus calcium phosphate increases the buffering capacity of the intestinal lumen and has cytoprotective effects, which may preserve the mucosal barrier.

In conclusion, the present study shows that FOS and inulin impair resistance to salmonella infections in rats. This effect is likely due to the rapid production of fermentation metabolites and subsequent impairment of the mucosal barrier. Dietary calcium phosphate inhibited the adverse effects of acidic fermentation and largely prevented damage to the intestinal barrier. The results of the present study await verification in other controlled animal and human studies. However, considering the current interest in health foods supplemented with fermentable fibres, concern is warranted.

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The authors wish to thank Wim van Doesburg for analytical analysis and the biotechnicians working at the Small Animal Centre of Wageningen University for expert biotechnical assistance.

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Transcellularly. Surprisingly, the effects of inulin and FOS were similar. Probably both prebiotics were equally rapidly fermented in the distal gut resulting in damage to the mucosal barrier. In addition, although organic acids inhibit growth of salmonella in vitro, prebiotic induced alterations in organic acid concentrations may increase salmonella virulence and hence increase translocation.

Most adverse effects of inulin and FOS were inhibited by dietary calcium. Calcium forms an insoluble complex with phosphate in the upper small intestine. Fermentation of inulin and FOS results in a considerable production of organic acids and hence acidification of gut contents. This can subsequently be counteracted by solubilisation of the calcium phosphate complex. However, when the dietary calcium phosphate supply is limited, the amounts of insoluble calcium phosphate may be insufficient to counteract acidification. Moreover, the present study and previous studies show that both inulin and FOS stimulate calcium absorption. Subsequently, less calcium phosphate will be available within the intestinal lumen to counteract the adverse effects of acidic fermentation.

Another mechanism by which calcium may improve the mucosal barrier is precipitation of cytotoxic components within the intestinal lumen. The amorphous calcium phosphate complex precipitates cytotoxic components in the intestinal lumen, which reduces epithelial cell damage and increases resistance towards salmonella translocation. Indeed, dietary calcium inhibited the cytotoxicity of faecal water, induced by inulin and FOS. The cytotoxicity assay was performed in a buffer of neutral pH, precluding simple acid induced lysis. Thus calcium phosphate increases the buffering capacity of the intestinal lumen and has cytoprotective effects, which may preserve the mucosal barrier.

In conclusion, the present study shows that FOS and inulin impair resistance to salmonella infections in rats. This effect is likely due to the rapid production of fermentation metabolites and subsequent impairment of the mucosal barrier. Dietary calcium phosphate inhibited the adverse effects of acidic fermentation and largely prevented damage to the intestinal barrier. The results of the present study await verification in other controlled animal and human studies. However, considering the current interest in health foods supplemented with fermentable fibres, concern is warranted.

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