Signals from dying hepatocytes trigger growth of liver progenitors

Youngmi Jung,1 Rafal P Witek,1 Wing-Kin Syn,1 Steve S Choi,1 Alessia Omenetti,1 Richard Premont,1 Cynthia D Guy,2 Anna Mae Diehl1

ABSTRACT
Objective The death rate of mature hepatocytes is chronically increased in various liver diseases, triggering responses that prevent liver atrophy, but often cause fibrosis. Mice with targeted disruption of inhibitor kappa B kinase (Ikk) in hepatocytes (ΔHEP mice) provide a model to investigate this process because inhibiting Ikk—nuclear factor-κB (NF-κB) signalling in hepatocytes increases their apoptosis.

Methods Cell proliferation, apoptosis, progenitors, fibrosis and production of Hedgehog (Hh) ligands (progenitor and myofibroblast growth factors) were compared in ΔHEP and control mice before and after feeding methionine choline-deficient ethionine-supplemented (MCDE) diets. Ikkβ was deleted from primary hepatocytes to determine the effects on Hh ligand production; Hh signalling was inhibited directly in progenitors to determine the effects on viability. Liver sections from patients were examined to assess relationships between hepatocyte production of Hh ligands, accumulation of myofibroblastic cells and liver fibrosis.

Results Disrupting the Ikk—NF-κB pathway in hepatocytes inhibited their proliferation but induced their production of Hh ligands. The latter provided viability signals for progenitors and myofibroblasts, enhancing accumulation of these cell types and causing fibrogenesis. Findings in the mouse models were recapitulated in diseased human livers.

Conclusion Dying mature hepatocytes produce Hh ligands which promote the compensatory outgrowth of progenitors and myofibroblasts. These results help to explain why diseases that chronically increase hepatocyte death promote cirrhosis.

INTRODUCTION
Repetitive exposure to various insults chronically increases the death rate of mature hepatocytes and triggers reparative responses to reconstitute damaged hepatic epithelium. Chronic liver injury is often accompanied by progressive liver fibrosis. However, it is not fully understood why situations that cause chronic, but relatively incremental, increases in the rate of mature hepatocyte death typically result in some degree of hepatic architectural distortion and tend to promote liver fibrosis, when such outcomes rarely complicate recovery from acute injuries that abruptly cause massive hepatocyte loss.

In order to investigate regenerative mechanisms associated with chronically increased rates of hepatocyte death, we took advantage of mice with targeted disruption of the kinase that promotes nuclear accumulation of nuclear factor-κB (NF-κB), a transcription factor that controls expression of genes that encode hepatocyte viability factors. NF-κB is held in the cytoplasm in its inactive form by a specific inhibitor, inhibitor κB (IκB). The IκB kinase (Ikk) complex phosphorylates IκB, allowing NF-κB to translocate into the nucleus. Ikk is comprised of three subunits, Ikkα, Ikkβ and Ikkγ. Hepatocyte-specific deletion of Ikkβ inhibits nuclear localization of NF-κB, reduces expression of hepatocyte viability factors and increases hepatocyte susceptibility to apoptosis.

The present study evaluates the hypothesis that hepatocytes that are prone to apoptosis have an impaired proliferative capacity, but elaborate paracrine signals that trigger compensatory growth of...
progenitors which ultimately regenerate the hepatic parenchyma. Further, we theorise that this process also promotes hepatic fibrogenesis. Hedgehog (Hh) ligands activate a morpho- 
genic signalling pathway that orchestrates tissue construction during development and adult wound healing. Because Hh pathway activity is low in healthy livers but high in various types of diseased livers, and Hh signalling promotes the growth of liver progenitors and myofibroblasts,1, 2 our aims were to determine if, and how, disrupting NF-kB survival signals in hepatocytes activates the Hh pathway in other liver cell types that are involved in fibrogenic repair of damaged livers. The results confirm our hypothesis and demonstrate an unsuspected mechanism that couples hepatocyte death to regenerative strategies that result in fibrogenic repair of liver damage.

**MATERIAL AND METHODS**

**Experimental animals**

IKKβ1/2 and IKKβHep mice were generous gifts from Dr Michael Karin (University of California San Diego). IKKβ1/2 mice were bred with Alb-Cre mice to generate mice in which all albumin-expressing cells (eg, hepatocytes) and their progeny are IKKβ deficient (referred to as IKKβHep). All mice were housed in a facility with a 12 h light/dark cycle and allowed free access to food and water. Adult mice were used between 5 and 8 months of age. To induce oxidative liver injury, inhibit replication of mature hepatocytes and activate hepatic stellate cells (HSCs) and liver progenitor populations, IKKβ1/2 and IKKβHep mice (n=9) were fed a methionine/choline-deficient diet supplemented with 0.15% ethionine (MCDE). Surviving mice were sacrificed after being fed MCDE diets for 1 week (IKKβ1/2 mice n=7; IKKβHep n=5). Chow-fed IKKβ1/2 (n=6) and IKKβHep mice (n=6) were also sacrificed at the same time point.

Animal care and surgical procedures were approved by the Duke University Medical Center Institutional Animal Care and Use Committee as set forth in the ‘Guide for the Care and Use of Laboratory Animals’ published by the National Institutes of Health.

**Human subjects**

Anonymised liver biopsy-proven non-alcoholic steatohepatitis (NASH) (n=5) and NASH-related cirrhosis (n=6) specimens were obtained from the Duke University, Division of Gastroenterology and Department of Pathology. The control liver tissues were obtained from the Duke University School of Medicine Tissue Bank Shared Resource and studied in accordance with NIH and Institutional guidelines for human subject research.

**Liver histology and immunohistochemistry**

Specimens fixed in formalin and embedded in paraffin were cut into 4 μm sections, dewaxed, hydrated, and subsequently incubated for 10 min in 3% hydrogen peroxide to block endogenous peroxidase. Antigen retrieval was performed by heating in 10 mM citrate buffer (pH 6.0) for 10 min or incubation with 0.5% H2O2 for 10 min. Sections were blocked in Dako protein block (X9090; Dako Envision, Dako, USA) for 30 min and incubated with primary antibodies, active caspase 3 (9661, 1:250; Cell Signaling, Danvers, Massachusetts, USA), Ki67 (NCL-Ki67, 1:100; Novocastra, Leica Microsystems, Newcastle Upon Tyne, UK), AE1/AE3 (15-0132, 1:500; Invitrogen, Carlsbad, California, USA), A6 (a gift from Dr Valentina Factor, National Institutes of Health, Bethesda, Maryland, USA), NF-kB p65 (ab7970, 1:5000; Abcam, Cambridge, Massachusetts, USA), GlI2 (26056; 1:4500; Abcam) and Sonic hedgehog (Shh; 1843-1, 1:7500; Epitomics, Burlingame, CA) at 4°C overnight. Other sections were also incubated at 4°C overnight in non-immune sera. Polymer- 

horseradish peroxidase (HRP) antirabbit (K4003; Dako), antimouse (K4001; Dako) or MACH3 mouse AP polymer kit (MP530, Biocare Medical, Concord, California, USA) was used as secondary antibody. 3,3’-Diaminobenzidine (DAB) was employed in the detection procedure. Omitting primary antibodies from the reactions eliminated staining, demonstrating staining specificity.

Sirius red staining and AE1/AE3 and A6 immunohistochemical staining were assessed by morphometry (MetaView software, Universal Imaging Corp, Downingtown, Pennsylvania, USA). To quantify Sirius red staining, 10 randomly chosen 20× fields/section were evaluated for each mouse. The quantification of AE1/AE3 and A6 staining was evaluated by randomly choosing eight portal tracts (PTs) excluding the major bile duct, at 40× fields/section for each mouse. After excluding the major bile duct in each PT from consideration, cells staining positively for Ki67, p65 (cytosol or nuclear) or GlI2 (a Hh-regulated transcription factor) in eight PTs/slide were counted at ×40 magnification. PTs selected for analysis contained a portal vein that ranged from 120 to 180 μm. The average number of Ki67- or GlI2-positive ductular cells was obtained by dividing the total number of positive cells by the total number of PTs. Ki67-, p65- or GlI2-positive hepatocytes were quantified by counting the total number of Ki67-, p65- or GlI2-positive hepatocytes/field and dividing by the total number of hepatocytes/field.

**RNA and protein analysis**

Total RNA and protein were extracted from freeze-clamped liver samples that had been stored at −80°C. A detailed protocol and the antibodies used are listed in the Supplementary Materials and methods. The sequences of primers for mice are summarised in Supplementary table 1.

**Hepatic hydroxyproline assay**

The hydroxyproline content in whole liver specimens was quantified colourimetrically as described in the Supplementary Materials and methods.

**Isolation and study of primary liver cells from mice**

Primary hepatocytes were obtained as described6 9 To isolate primary liver progenitors, we performed a two-step collagenase perfusion according to the protocol of Selgen et al.10 and collected the non-parenchymal cell (NPC) fraction that contained progenitors using gradient centrifugation. Isolated cells were incubated with anti-Sca (stem cell antigen)-1 fluorescent isothiocyanate (FITC)-conjugated antibody (130-092-529 MACs, Miltenyi Biotec, GmbH, Bergisch Gladbach, Germany), followed by anti-FITC microbeads. After incubation, cells were positively selected using a MACS (magnetic-activated cell separation) sort.11 12 As determined by trypan blue exclusion, cell viability was >90% in all experiments.

To evaluate apoptotic activity, hepatocytes and progenitors were cultured for 24 h and then cytoscenfirmed to slides and examined for annexin V (ab14196, Abcam). For double immuno-fluorescent staining, cytopsin cells were fixed, permeabilised, and processed for immunostaining with primary antibody annexin V and albumin (MAB1455, 1:100; R&D Systems, Minneapolis, Minnesota, USA). Alexa Fluor 568 and Alexa Fluor 488 (Molecular Probes, Eugene, Oregon, USA) were used as secondary antibodies. 4,6-Diamidino-2-phenylindole (DAPI) counterstaining was employed to demonstrate nuclei. Hepatocytes were also labelled with anti-mouse CD26 (559652, BD Biosciences, San Jose, California, USA) and annexin V (559933, BD Biosciences) or matched isotype control antibodies, and analysed using the FACS VantageSe (Beckton Dickinson, Franklin Lakes, New Jersey, USA).
In separate experiments other freshly isolated progenitors were seeded in 96-well tissue culture plates (2000 cells/well) and grown in Iscove’s modified Dulbecco’s medium (IMDM) supplemented with 10% fetal bovine serum (FBS). After 72 h, the medium was replaced with 1% FBS IMDM; the next morning cells were treated with either cyclopamine (3 μM, Toronto Research Chemcals, North York, Ontario, Canada) or tomatidine (3 μM, Calbiochem, San Diego, California, USA) for 24 h. Control wells were treated with an equal volume of vehicle (dimethylsulfoxide (DMSO)) for the same treatment period. At the end of the treatment period, cell numbers were determined by Cell Counting Kit-8 (CCK-8, Dojindo, Rockvilee, Maryland, USA). A calibration curve was prepared using wells containing a fixed numbers of viable cells. A FLUOstar OPTIMA microplate reader (BMG Labtech, Durham, North Carolina, USA) was used for absorbance measurements.

Primary hepatocytes isolated from wild-type and IkkβF/F mice (n=5/group) were cultured in the presence of adenoviral vectors carrying green fluorescent protein (AdGFP) or Cre recombinase (AdCre) (multiplicity of infection (MOI) 25). Cultures were analysed at 24 and 48 h.

**Statistical analysis**
Results are expressed as the mean±SEM. Significance was established using the Student t test and analysis of variance when appropriate. Differences were considered significant when p<0.05.

**RESULTS**
IkkβDΔHep mice have more apoptotic hepatocytes and increased liver progenitors

Although livers of IkkβDΔHep mice appeared grossly normal at baseline (Supplementary figure 1), they had increased caspase 3 activity by immunohistochemistry (IHC) (figure 1A, B) and western blot analysis (figure 1C). IHC of primary hepatocytes that were double-stained for annexin V, a marker of cells carrying green fluorescent protein (AdGFP) or Cre recombinase (AdCre) (multiplicity of infection (MOI) 25). Cultures were analysed at 24 and 48 h.

**Statistical analysis**
Results are expressed as the mean±SEM. Significance was established using the Student t test and analysis of variance when appropriate. Differences were considered significant when p<0.05.

**RESULTS**
IkkβDΔHep mice have more apoptotic hepatocytes and increased liver progenitors

Although livers of IkkβDΔHep mice appeared grossly normal at baseline (Supplementary figure 1), they had increased caspase 3 activity by immunohistochemistry (IHC) (figure 1A, B) and western blot analysis (figure 1C). IHC of primary hepatocytes that were double-stained for annexin V, a marker of cells carrying green fluorescent protein (AdGFP) or Cre recombinase (AdCre) (multiplicity of infection (MOI) 25). Cultures were analysed at 24 and 48 h.

**Statistical analysis**
Results are expressed as the mean±SEM. Significance was established using the Student t test and analysis of variance when appropriate. Differences were considered significant when p<0.05.

**RESULTS**
IkkβDΔHep mice have more apoptotic hepatocytes and increased liver progenitors

Although livers of IkkβDΔHep mice appeared grossly normal at baseline (Supplementary figure 1), they had increased caspase 3 activity by immunohistochemistry (IHC) (figure 1A, B) and western blot analysis (figure 1C). IHC of primary hepatocytes that were double-stained for annexin V, a marker of cells carrying green fluorescent protein (AdGFP) or Cre recombinase (AdCre) (multiplicity of infection (MOI) 25). Cultures were analysed at 24 and 48 h.

**Statistical analysis**
Results are expressed as the mean±SEM. Significance was established using the Student t test and analysis of variance when appropriate. Differences were considered significant when p<0.05.

**RESULTS**
IkkβDΔHep mice have more apoptotic hepatocytes and increased liver progenitors

Although livers of IkkβDΔHep mice appeared grossly normal at baseline (Supplementary figure 1), they had increased caspase 3 activity by immunohistochemistry (IHC) (figure 1A, B) and western blot analysis (figure 1C). IHC of primary hepatocytes that were double-stained for annexin V, a marker of cells carrying green fluorescent protein (AdGFP) or Cre recombinase (AdCre) (multiplicity of infection (MOI) 25). Cultures were analysed at 24 and 48 h.

**Statistical analysis**
Results are expressed as the mean±SEM. Significance was established using the Student t test and analysis of variance when appropriate. Differences were considered significant when p<0.05.

**RESULTS**
IkkβDΔHep mice have more apoptotic hepatocytes and increased liver progenitors

Although livers of IkkβDΔHep mice appeared grossly normal at baseline (Supplementary figure 1), they had increased caspase 3 activity by immunohistochemistry (IHC) (figure 1A, B) and western blot analysis (figure 1C). IHC of primary hepatocytes that were double-stained for annexin V, a marker of cells carrying green fluorescent protein (AdGFP) or Cre recombinase (AdCre) (multiplicity of infection (MOI) 25). Cultures were analysed at 24 and 48 h.

**Statistical analysis**
Results are expressed as the mean±SEM. Significance was established using the Student t test and analysis of variance when appropriate. Differences were considered significant when p<0.05.
undergoing apoptosis, and albumin, a marker for hepatocytes, confirmed that many albumin-positive hepatocytes from IkkBΔHep mice were annexin V positive, whereas albumin-positive hepatocytes from IkkB/F/F mice were generally negative for annexin V (figure 1D). Fluorescence-activated cell sorting (FACS) analysis of primary hepatocytes which were double-stained for annexin V and CD26, another marker for mature hepatocytes, verified that more hepatocytes from IkkBΔHep mice were positive for annexin V (figure 1E). Hence, hepatocyte-specific deletion of IkkB increased baseline hepatocyte apoptotic activity.

To determine if this led to expansion of hepatocyte progenitor populations, livers from IkkBΔHep mice and IkkB/F/F controls were stained for AE1/AE3 and A6, two different markers of liver progenitors. Compared with livers of IkkB/F/F mice, livers of IkkBΔHep mice contained more AE1/AE3-positive cells, and more A6-positive cells at baseline (figure 2A–C). Hence, hepatocyte-specific deletion of IkkB expanded hepatic progenitor populations in seemingly healthy IkkBΔHep mice.

Figure 2 Increased hepatic progenitor populations in IkkBΔHep mice before and after hepatic damage. (A) Immunohistochemical staining for progenitor markers, AE1/AE3 and A6, in liver sections from representative IkkB/F/F and IkkBΔHep mice before (baseline) and after methionine choline-deficient ethionine-supplemented (MCDE) diets for 1 week (×40). Quantitative AE1/AE3 (B, D) or A6 (C, E) immunohistochemistry data from IkkB/F/F and IkkBΔHep mice before (B, AE1/AE3; C, A6) and after injury (D, AE1/AE3; E, A6). AE1/AE3- or A6-positive cells were measured by morphometry in eight portal tracts (PTS)/section. Mean±SD are graphed (*p<0.05, **p<0.005 vs the IkkB/F/F group).

To determine if these changes in hepatic progenitor cell populations are associated with altered hepatocyte proliferative responses, liver sections from MCDE-fed IkkB/F/F and IkkBΔHep mice were stained for Ki67, a marker of S phase. The injured livers of IkkB/F/F mice contained 20-fold more Ki67-positive hepatocytes than those of IkkBΔHep mice (figure 3C, E). Interestingly, although IkkBΔHep mice had reduced hepatocyte replication, these mice demonstrated greater proliferation of ductular cells than IkkB/F/F mice.
both at baseline and after liver injury (figure 3D, E). Because the ductular cell population is enriched with liver progenitors, these findings suggested that hepatocyte-specific deletion of Ikkβ promoted proliferation of liver progenitor populations.

**Increased progenitors in IkkβΔHEp mice during liver damage**

To investigate this possibility further, livers from MCDE diet-fed IkkβF/F and IkkβΔHEp mice were stained for the progenitor markers, AE1/3 and A6. Liver injury induced greater accumulation of AE1/AE3-positive cells in IkkβΔHEp mice than in IkkβF/F mice (figure 2A, D). Similarly, numbers of A6-positive progenitors were also greater in IkkβΔHEp mice than in IkkβF/F mice (figure 2A, E). Therefore, hepatocyte-specific deletion of Ikkβ promoted compensatory expansion of progenitor populations after liver injury.

**Increased Hh signalling in IkkβΔHEp mice promotes progenitor viability**

Evidence that mature hepatocytes from IkkβΔHEp mice were prone to apoptosis, but that progenitors accumulated in IkkβΔHEp livers, suggested that hepatic progenitors might have survival mechanisms that were distinct from those that operate in mature hepatocytes. Since activation of the Hh pathway is known to

---

**Figure 3** Reduced nuclear localisation of nuclear factor-κB (NF-κB) p65 and decreased hepatocyte Ki67 expression in IkkβΔHEp mice after hepatic injury. (A) Immunohistochemical staining for NF-κB p65 in liver from representative IkkβF/F and IkkβΔHEp mice before (baseline) and after methionine choline-deficient ethionine-supplemented (MCDE) diets for 1 week (×40). (B) Quantitative NF-κB p65 immunohistochemistry data from MCDE-fed mice (n=4 mice/group). The numbers of hepatocytic cells (HEP) or ductular cells with nuclear or cytosolic NF-κB p65 were counted in eight portal tracts (PTs)/section. Results are expressed as numbers of NF-κB p65(+) cells per PT and graphed as mean±SD results (*p<0.05, **p<0.005 vs the IkkβF/F MCDE-fed group). (C, D) Immunohistochemical staining with Ki67 (cell proliferation marker) in liver sections from representative IkkβF/F and IkkβΔHEp mice after MCDE diet-induced liver injury. (E) Quantitative Ki67 immunohistochemistry data from all mice (n=4 mice/group). The numbers of Ki67(+) HEPs and ductular cells were counted in eight PTs/section. Ki67(+) HEPs were quantified by counting the total number of Ki67(+) HEPs/field and dividing by the total number of HEPs/field. Ki67(+) ductular cells were quantified by dividing the total number of positive cells by the total number of PTs. Mean±SD results are graphed (*p<0.05, **p<0.005 vs the IkkβF/F chow-fed control group).
regulate the viability of many types of progenitors, including fetal liver progenitors and ductular-type progenitors in adult livers, we next evaluated Hh pathway activity in Ikk\(^{\beta}\Delta\text{HEP}\) mice.

First we clarified which types of liver cells were capable of Hh signalling by isolating progenitor populations and mature hepatocytes from healthy Ikk\(^{\beta}\Delta\text{HEP}\) mice (n=4 mice/group). Due to the relatively low numbers of progenitors in adult livers, progenitor cells were pooled from 2–3 mice/group for RNA and protein extraction. Quantitative reverse transcription—PCR (qRT—PCR) analysis of RNA expression of the mutant Ikk\(^{\beta}\) allele, albumin, Cre recombinase (cre) and glyceraldehyde phosphate dehydrogenase (gapdh) in representative mice. Western blot analysis of Gli2 (133 kDa) and \(\beta\) actin expression in the same mice. (B) Primary progenitors from both groups were then cultured with cyclopamine (3 \(\mu\)M) or tomatidine (3 \(\mu\)M) for 24 h. Control wells were treated with an equal volume of vehicle (dimethylsulfoxide (DMSO)) for the same treatment period. At the end of the treatment period, growth (cell number) was assessed using the CCK-8 assay. Each experiment was replicated three times. Mean±SD results are graphed (*p<0.05, **p<0.005 vs respective the 0 h control group). (C) QRT—PCR analysis of liver RNA from chow-fed Ikk\(^{\beta}\)F/F and Ikk\(^{\beta}\Delta\text{HEP}\) mice (open bars), and methionine choline-deficient ethionine-supplemented (MCDE) diet-fed Ikk\(^{\beta}\)F/F and Ikk\(^{\beta}\Delta\text{HEP}\) mice (black bars) for Indian hedgehog (Ihh), Patched (Ptc), Gli2 and frizzled-related peptide (Frp)1 (n=4 mice/group/treatment). Mean±SD results are graphed (*p<0.05, **p<0.005 vs the Ikk\(^{\beta}\)F/F chow-fed control group).
progenitor populations from both Ikkβ F/F and IkkβΔHep mice expressed Gli2 proteins, whereas mature hepatocytes from neither strain expressed Gli2 (figure 4A). Thus, the hepatic progenitor populations were capable of activating Hh signalling and retained this capability even when their NF-kB pathway was disrupted.

To investigate the role of Hh signalling in maintaining progenitor viability during liver injury more directly, we isolated progenitor populations from MCDE-fed Ikkβ F/F and IkkβΔHep mice, placed them in culture and treated the cultures with vehicle, cyclopamine, a specific pharmacological inhibitor of the Hh pathway, or tomatidine, an inactive cyclopamine analogue. The effects on cell numbers were quantified. Neither vehicle nor tomatidine altered the number of Ikkβ F/F or IkkβΔHep progenitors. In contrast, cyclopamine significantly reduced numbers in both groups, decreasing cell counts by >50% within 24 h (figure 4B). Hence, Hh signalling permitted the survival of liver progenitor populations during chronic liver injury, including the IkkβΔHep progenitors that had a defective NF-kB pathway.

Increased expression of Hh ligands and target genes in livers of IkkβΔHep mice

Because progenitor populations were increased in the livers of IkkβΔHep mice, both at baseline and during superimposed injury, we next asked if the Hh pathway was generally activated in mice with disruption of Ikk–NF-kB signalling in hepatocytes. Compared with Ikkβ F/F mice, IkkβΔHep mice had greater mRNA expression of the Hh ligand, Indian Hh (Ihh), and several Hh target genes, including the Hh receptor, Patched (Ptc), Gli2, a Hh-regulated transcription factor, and frizzled-related peptide (Frp)-1, a Hh target gene and soluble inhibitor of canonical Wnt signalling, both before and after liver injury (figure 4C).

Disrupting the Ikk–NF-kB pathway in hepatocytes triggers their production of Hh ligands

Since differences in Hh pathway activation between IkkβΔHep and Ikkβ F/F mice accompany targeted deletion of Ikkβ in mature hepatocytes, we isolated primary hepatocytes from Ikkβ F/F and wild-type mice and treated the cells with adenoviral vectors carrying either Cre recombinase (AdCre) or green fluorescent protein (AdGFP) to determine if acutely deleting Ikkβ function influenced hepatocyte production of Hh ligands (figure 5). Incubation with AdGFP efficiently transduced virtually all of the hepatocytes and did not reduce cell viability or affect Hh ligand expression. Similarly, treating wild-type hepatocytes with AdCre had no effect on either of these parameters. In contrast, AdCre treatment significantly reduced viability and increased mRNA expression of Ihh ligand in hepatocytes from Ikkβ F/F mice (figure 5A,B). Western blot analysis of protein lysates from Ikkβ F/F hepatocytes that were treated with AdGFP or AdCre demonstrated that acute AdCre-mediated disruption of Ikkβ led to caspase 3 activation and increased cellular production of both Shh and Ihh proteins (figure 5C,D). However, AdCre-treated hepatocytes did not exhibit increased expression of Hh target genes, such as Ptc or Gli2 (data not shown). Therefore, interrupting Ikkβ function in isolated hepatocytes reduced their viability and induced them to produce Hh ligands that function as viability factors for liver progenitors.

Increased production of Hh ligands causes hepatic accumulation of Hh-responsive cells

Subsequent experiments used IHC for the Hh-regulated transcription factor Gli2, to identify which types of liver cells were Hh responsive in intact animals. Although few Gli2-positive cells were evident in the uninjured livers of either Ikkβ F/F or IkkβΔHep mice, quantitative analysis demonstrated that...
IkkβD mice had more Gli-positive hepatocytic cells and Gli-positive ductular cells than IkkβF/F mice at baseline. The numbers of Gli2-positive cells increased in both groups after MCDE diet treatment, but the greatest numbers of these Hh-responsive cells accumulated in IkkβD Hep livers (figure 6A–C). After only 1 week of diet-induced liver injury, the net numbers of Hh-responsive cells were about threefold greater in IkkβD Hep mice than in IkkβF/F mice.

Evidence that neither mature hepatocytes from IkkβF/F mice nor hepatocytes from IkkβD Hep mice expressed Gli2 at baseline, and that targeted disruption of Ikkβ function in cultured hepatocytes failed to activate expression of Hh-regulated genes suggested that the Gli2-positive hepatocytic cells in injured livers might have been derived from Hh-responsive progenitors that survived liver injury and then partially differentiated to replace dead hepatocytes. This concept was evaluated by double immunostaining liver sections for Gli2 and pancytokeratin, a marker of immature hepatocytes. Co-localisation of Gli2 and pancytokeratin was demonstrated in hepatocytic cells, particularly periportally (figure 6D), supporting the possibility that Gli2-expressing hepatocytic cells in IkkβD Hep mice were immediate progeny of Gli2-positive progenitors that survived liver injury.

Enhanced fibrogenesis and liver fibrosis in IkkβD Hep mice, and increased hepatocyte expression of Shh in patients with liver fibrosis

Hepatocytes in healthy livers lack a basement membrane and, hence, are immediately adjacent to HSCs that reside in the space of Disse. Cells that produce Hh ligands release these factors in

Figure 6 Increased accumulation of Hedgehog (Hh)-responsive cells in IkkβD Hep mice after liver injury. (A) Immunohistochemical staining for Gli2 in representative IkkβF/F and IkkβD Hep mice before (baseline) and after methionine choline-deficient ethionine-supplemented (MCDE) diets for 1 week (×40). (B, C) Quantitative Gli2 immunohistochemistry data from all mice (n=4 mice/group). Gli2(+) hepatocytic cells (HEPs) or ductular cells were counted in eight (×40 magnification) fields that contained portal tracts (PTs). (B) Gli2(+) hepatocytic cells were expressed as a percentage of Gli2(+) hepatocyte nuclei/total HEPs. (C) Gli2(+) ductular cells were quantified by dividing the total number of positive cells by the total number of PTs. Mean±SD results are graphed (*p<0.05, **p<0.005 vs the IkkβF/F chow-fed control group). (D) Double immunohistochemical staining with Gli2 (brown) and pancytokeratin (blue) in IkkβD Hep mice after MCDE treatment (original magnification ×63).
membranous particles, and this process increases during apoptosis. Thus, HSCs are proximal targets of Hh ligands that are released from hepatocytes in injured livers. This may promote fibrogenesis because Hh ligands activate quiescent HSCs to become myofibroblastic (MF) and promote the viability and proliferation of MF-HSCs. To assess this issue, we performed western blot analysis to compare expression of α-smooth muscle actin (α-SMA), a marker of MF-HSCs, in IkkβF/F and IkkβD Hep mice. Levels of α-SMA protein were significantly greater in the IkkβD Hep group, both before and after MCDE diet treatment (figure 7A). Sirius red staining demonstrated that IkkβD Hep mice also had more pericellular and sinusoidal deposition of collagen fibrils than IkkβF/F mice at both time points (figure 7B). Biochemical determination of hepatic hydroxyproline content, a quantitative measure of liver fibrosis, confirmed that IkkβD Hep mice had significantly more liver fibrosis than IkkβF/F mice (figure 7C). Therefore, activation of the Hh pathway promoted accumulation of MF cells and increased fibrogenesis in mice with chronically increased hepatocyte apoptosis.

To determine if a similar process might occur in humans, we performed immunostaining for Shh and α-SMA on liver biopsy samples from 12 patients with early (stage F2) or advanced (stage F4) fibrosis due to chronic NASH. In parallel, similar staining was done on healthy control liver tissue. NASH livers were selected for scrutiny because levels of hepatic apoptotic activity correlate with the severity of liver fibrosis in this disease. No Shh-positive hepatocytes were identified in healthy control livers (data not shown). In NASH livers, hepatocytes that stained for Shh were variably detected, with greater numbers of more strongly stained hepatocytes occurring in livers with more fibrosis (figure 8A–C, Supplementary figure 3A,B). Fibrotic NASH livers typically demonstrated foci of hepatocytes

Figure 7 Enhanced fibrogenesis and liver fibrosis in IkkβD Hep mice. (A) α-SMA protein expression was assessed in whole liver proteins from IkkβF/F and IkkβD Hep mice (n=3/group/treatment) using western blot analysis. The graph demonstrates the mean±SD morphometric data from all mice (n=4/group/treatment). (B) Sirius red staining in liver sections from representative mice (×20 or ×40). The graph demonstrates the mean±SD morphometric data from all mice (n=4/group/treatment). (C) Hepatic hydroxyproline content in all mice (n=4 mice/group). All results are displayed as mean±SD (*p<0.05, **p<0.005 vs the IkkβF/F chow-fed control group). MCDE, methionine-deficient ethionine-supplemented.
that were surrounded by stromal cells that expressed the myofibroblast marker, \(\alpha\)-SMA. Double-immunostaining for Shh (brown) and \(\alpha\)-SMA (blue) confirmed that \(\alpha\)-SMA-positive myofibroblastic cells tended to localise near regions of Shh-expressing hepatocytes within the hepatic parenchyma (figure 8D,E). Thus, in NASH, a type of human liver disease in which there is increased hepatocyte apoptosis, increased hepatocyte expression of Hh ligands occurs, and this is accompanied by evidence of HSC activation and increased fibrogenesis.

**DISCUSSION**

Our results in \(\text{IkB}\beta\Delta\text{Hep}\) mice, a murine model of increased hepatocyte death, demonstrate that optimal viability and proliferative capacities of mature hepatocytes are heavily dependent upon an intact NF-\(\kappa\)B pathway, whereas Hh signalling in liver progenitors provides an alternative survival mechanism that permits them to survive and proliferate even when NF-\(\kappa\)B nuclear localisation is impaired. Moreover, we discovered that when the \(\text{IkB}\beta\Delta\text{Hep}\) pathway is disrupted in mature hepatocytes, they produce Hh ligands. This enriches the hepatic microenvironment with factors that enhance the growth of \(\text{IkB}\beta\)-deficient progenitor cells. As these progenitors mature, they eventually lose the ability to transduce Hh-initiated signals, and become more reliant on NF-\(\kappa\)B-regulated viability factors. Thus, their vulnerability to apoptosis increases because \(\text{IkB}\beta\Delta\text{Hep}\) signalling has been disrupted, and this perpetuates the increased rate of mature hepatocyte apoptosis. The latter maintains heightened production of Hh ligands and stimulates the continued outgrowth of Hh-responsive non-parenchymal cells that rely on Hh signalling to survive, including myofibroblasts. Progressive hepatic accumulation of such cells promotes fibrogenic repair, helping to explain why liver fibrosis is increased in \(\text{IkB}\beta\Delta\text{Hep}\) mice.

Increased hepatocyte production of Hh ligands and accumulation of Hh target cells (ie, MF cells) were also demonstrated in patients with NASH. Increased rates of hepatocyte apoptosis are known to occur in this common type of chronic human liver disease. Moreover, hepatic apoptotic activity predicts the severity of liver fibrosis in NASH, and is significantly greater in patients with advanced fibrosis than in patients with early fibrosis.\(^{20}\) MF-HSCs play major roles in excessive matrix accumulation during many types of chronic liver disease, including NASH. Earlier work showed that phagocytosis of apoptotic bodies activates quiescent HSCs (Q-HSCs) to become MF-HSCs.\(^{21}\) Therefore, the early stages of fibrosis in NASH are characterised by deposition of fibrous matrix around hepatocytes and along sinusoids.\(^{22}\)

The new evidence that injured hepatocytes in NASH produce Shh ligands provides another mechanism that helps to explain the pattern of liver fibrosis that occurs in NASH because HSCs...
are capable of responding to Hh ligands. In healthy livers, Q-HSCs produce relatively large amounts of Hip (Hh-interacting protein), a potent Hh ligand antagonist. However, HSC expression of Hip is abolished by liver injury (or culture), permitting Hh-dependent induction of MF genes and enhancing the viability and proliferative activity of MF-HSCs.1819 MF-HSCs themselves produce and release Hh ligands,19 further enriching the hepatic microenvironment with these factors.

Mature Hh ligands diffuse away from the cells that produce them, and variably modulate gene expression in distant Hh-responsive cells. Target cell effects depend upon the concentration and duration of Hh ligand exposure.20 Membranous particles containing biologically active Hh ligands have been purified from the blood and bile of experimental animals with liver injury, suggesting routes by which Hh ligands generated by apoptotic hepatocytes and MF-HSCs might be disseminated both extrahepatically and to other parts of the liver lobule. Hepatic progenitors that reside in the canals of Hering and bile ductular cells in PTs are Hh responsive, and Hh ligands released from MF-HSCs have been shown to enhance the viability and proliferative activity of both cell types.24 This promotes the ductular response that often accompanies chronic liver injury,2425 Hh ligands also stimulate immature ductular cells to undergo epithelial to mesenchymal transition (EMT),26 a process that may further contribute to liver fibrosis.26 Indeed, quantitative reverse transcription–PCR (RT–PCR) analysis of liver RNA from Ikkβ−/− mice demonstrated increased expression of various EMT-related factors (Supplementary figure 4). Hence, evidence that dying hepatocytes release morphogens that trigger hepatic accumulation of cell types that mediate fibrogenic repair suggests a unifying mechanism that helps to explain the strong association between chronic liver injury and fibrogenesis. This discovery identifies novel diagnostic and therapeutic targets for further research that are relevant to various types of chronic liver injury.

Acknowledgements We like to thank Dr Michael Karin (University of California, San Diego) for providing the Ikk−/− and Ikk−/−HEP mice and helpful discussion.

Funding This work was supported by NIH grant 5R01-DK077794 to AMD.

Competing interests None.

Ethics approval This study was conducted with the approval of the Duke University, Division of Gastroenterology and Department of Pathology, Duke University School of Medicine Tissue Bank Shared Resource.

Provenance and peer review Not commissioned; externally peer reviewed.

REFERENCES
Signals from dying hepatocytes trigger growth of liver progenitors

Youngmi Jung, Rafal P Witek, Wing-Kin Syn, Steve S Choi, Alessia Omenetti, Richard Premont, Cynthia D Guy and Anna Mae Diehl

*Gut* 2010 59: 655-665
doi: 10.1136/gut.2009.204354

Updated information and services can be found at:
http://gut.bmj.com/content/59/5/655

These include:

**Supplementary Material**
Supplementary material can be found at:
http://gut.bmj.com/content/suppl/2010/06/30/59.5.655.DC1

**References**
This article cites 25 articles, 7 of which you can access for free at:
http://gut.bmj.com/content/59/5/655#BIBL

**Email alerting service**
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

**Topic Collections**
Articles on similar topics can be found in the following collections

- Editor's choice (120)
- Cirrhosis (331)

**Notes**

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/