MicroRNA-21 is a potential link between non-alcoholic fatty liver disease and hepatocellular carcinoma via modulation of the HBP1-p53-Srebp1c pathway

Heng Wu, Raymond Ng, Xin Chen, Clifford J Steer, Guisheng Song

ABSTRACT

Background Non-alcoholic fatty liver disease (NAFLD) is a major risk factor for hepatocellular carcinoma (HCC). However, the mechanistic pathways that link both disorders are essentially unknown.

Objective Our study was designed to investigate the role of microRNA-21 in the pathogenesis of NAFLD and its potential involvement in HCC.

Methods Wildtype mice maintained on a high fat diet (HFD) received tail vein injections of microRNA-21-anti-sense oligonucleotide (ASO) or miR-21 mismatched ASO for 4 or 8 weeks. Livers were collected after that time for lipid content and gene expression analysis. Human hepatoma HepG2 cells incubated with oleate were used to study the role of miR-21 in lipogenesis and analysed with Nile-Red staining. microRNA-21 function in carcinogenesis was determined by soft-agar colony formation, cell cycle analysis and xenograft tumour assay using HepG2 cells.

Results The expression of microRNA-21 was increased in the livers of HFD-treated mice and human HepG2 cells incubated with fatty acid. MicroRNA-21 knockdown in those mice and HepG2 cells impaired lipid accumulation and growth of xenograft tumour. Further studies revealed that Hbp1 was a novel target of microRNA-21 and a transcriptional activator of p53. It is well established that p53 is a tumour suppressor and an inhibitor of lipogenesis by inhibiting Srebp1c. As expected, microRNA-21 knockdown led to increased HBP1 and p53 and subsequently reduced lipogenesis and delayed G1/S transition, and the additional treatment of HBP1-siRNA antagonised the effect of microRNA-21-ASO, suggesting that HBP1 mediated the inhibitory effects of microRNA-21-ASO on both hepatic lipid accumulation and hepatocarcinogenesis. Mechanistically, microRNA-21 knockdown induced p53 transcription, which subsequently reduced expression of genes controlling lipogenesis and cell cycle transition. In contrast, the opposite result was observed with overexpression of microRNA-21, which prevented p53 transcription.

Conclusions Our findings reveal a novel mechanism by which microRNA-21, in part, promotes hepatic lipid accumulation and cancer progression by interacting with the Hbp1-p53-Srebp1c pathway and suggest the potential therapeutic value of microRNA-21-ASO for both disorders.

INTRODUCTION

The incidence of hepatocellular carcinoma (HCC) worldwide nearly matched its mortality, demonstrating the aggressive nature of this malignancy and limited therapeutic options. Although HBV and HCV are major risk factors of HCC, non-alcoholic fatty liver disease (NAFLD) remains a common underlying pathology to the majority of patients with HCC in the Western world. It is estimated that 90% of obese patients have some form of fatty liver, ranging from simple steatosis to more severe forms of non-alcoholic steatohepatitis (NASH) and cirrhosis with its associated high risk of HCC. In addition, given limited effects of chemotherapy and the relative insensitivity of HCC to radiotherapy, complete tumour extirpation represents the only choice for a long-term cure. Unfortunately, the
majority of patients are not eligible for surgical resection because of tumour extent or underlying liver dysfunctions including NAFLD. As described above, despite the strong association between NAFLD and HCC, the underlying mechanisms are largely unknown due in part to their complex nature of disease.

The discovery of a class of naturally occurring small non-coding RNAs, termed microRNAs (miRNAs), 4,5 has stimulated a new field of research on NAFLD and HCC. Alterations in miRNA expression have been reported in human individuals with NAFLD/NASH and HCC. 6,7 Reflective of their key roles in lipid metabolism and carcinogenesis, 8,9 miRNAs have been suggested as novel therapeutic targets for both metabolic diseases and human cancers. However, the miRNAs associated with both NAFLD and its potential sequel HCC are poorly described. Our interest in miR-21 arose initially from hepatocyte-specific miRNA profiling studies in mouse livers, in which we showed that miR-21 is highly expressed in hepatocytes. Furthermore, we observed that high fat diet (HFD) treatment significantly induced expression of miR-21 in livers of mice. By antagonising miR-21 in liver, we were able to prevent hepatic lipid accumulation in dietary obese mice. Consistent with our findings, miR-21 expression was significantly upregulated in human patients with NASH. 6 It is also known that miR-21 is a potent promoter of HCC and other human cancers. 7,9 These data led us to hypothesise that miR-21 plays an important role in the pathogenesis of NAFLD and its potential progression to HCC. In the present study, we have investigated the regulatory role of miR-21 in linking NAFLD and HCC in both in vivo and in vitro model systems.

MATERIALS AND METHODS

Bioinformatic analysis

Identification of miR-21 target genes was conducted as previously described with minor revision. 10 In detail, we compiled a list of downregulated genes in livers of patients with NAFLD/NASH by downloading their microarray data from GEO (http://www.ncbi.nlm.nih.gov/geo/). 11 mRNA profiles of six normal liver samples (male) and eight NAFLD/NASH liver samples (male) were compared using GeneSpring (Agilent Technologies Genomics). Differentially expressed genes were defined by a log-scale ratio ≤ 0.3 between paired samples with a p < 0.05. Based on these criteria, we identified 1219 downregulated probes in NAFLD/NASH samples (see online supplementary table S1). To identify genes with binding motifs for miR-21, we downloaded the target gene databases of miR-21 based on TargetScan, 12 Pictar 13 and Starbase. 14 Only hits from TargetScan or Pictar algorithm that were confirmed by Ago HITS-CLIP (high-throughput sequencing of RNAs isolated by cross-linking immunoprecipitation (HTS-CLIP) from Argonaute protein complex) were selected. These three databases were compared using Microsoft Access 2000, yielding 219 potential targets that have miR-21 binding motif (see online supplementary table S2). We then compared 1219 downregulated probes in livers of patients with NAFLD/NASH with 219 genes that have at least one binding motif for miR-21 using Microsoft Access 2010, which resulted in an overlap of 13 genes between two databases that were considered as potential targets of miR-21 (see online supplementary table S3). Gene ontology (GO) analysis was done using PathwayStudio software (Elsevier).

Animal, diet treatment and sample collection

Male Dicer1−/− mice on a mixed 129S4, C57Bl/6 strain background 15 were crossed with C57Bl/6 Alb-Cre+− mice 16 to generate Dicer1−/−, Alb-Cre+− mice (mice are from Dr Holger Willenbring’s lab at the University of California, San Francisco). To specifically investigate the impact of miRNAs on mature liver function, we initiated Cre recombinase expression in 8-week-old to 10-week-old mice. 17 To restrict Cre expression to hepatocytes, we used a hepatocyte-specific Transthyretin (Ttr) promoter and pseudotyped the vector genome with capsids from AAV8, a serotype that can transduce virtually all hepatocytes in vivo without causing toxicity. 10,17,18

To determine the effect of hepatic lipid accumulation on miRNA expression, 8-week-old wildtype male C57Bl/6 mice (Jackson Laboratory, n=6) were maintained on either a normal chow diet (Open Source D12450B: 10% Kcal fat) or an HFD (Open Source D12492: 60% Kcal fat) for 4 weeks as described by Vickers et al. 19 After 4 weeks of HFD administration, livers were collected for miRNA and gene expression analysis.

To determine the role of miR-21 in NAFLD, we synthesised locked nucleic acid anti-miR-21 anti-sense oligonucleotide (ASO) (Exiqon) specifically targeting miR-21 and also generated miR-21-mismatched-ASO (miR-21-MM-ASO), a control ASO that differs from miRNAs in four mismatched base pairs. The male C57Bl/6 mice kept on HFD for 4 weeks were divided into two groups; one group (n=8) was treated with miR-21-ASO and the other with miR-21-MM-ASO (control, n=8). Mice received a dose of 25 mg/kg miR-21-ASO or miR-21-MM-ASO (0.9% NaCl) weekly for 4 or 8 weeks via tail vein injection. At those times, the mice were anaesthetised, and blood was collected by way of cardiac puncture. Subsequently, the livers were harvested and immediately frozen in liquid nitrogen and stored at −80°C for gene expression and histological analysis.

Fatty acid treatment of HepG2 cells

HepG2 cells were obtained from Dr Xin Chen’s laboratory at the University of California, San Francisco. Sodium olate was obtained from Sigma-Aldrich and was dissolved in Dulbecco’s modified Eagle medium (DMEM) with 1% fatty acid free bovine serum albumin (BSA) (Sigma). Oleate treatment of HepG2 cells was carried out as previously described with minor revision. 10,20 Specifically, HepG2 cells were plated in four-well chamber slides with DMEM medium supplemented with 10% fetal bovine serum (Invitrogen). After 24 h, HepG2 cells were treated with either control medium (DMEM supplemented with 1% fatty acid free BSA) or medium containing olate (0.5 mM). The cells were cultured for another 24 h, then lipid accumulation and miR-21 expression were determined by Nile Red Staining (Sigma-Aldrich) and qRT-PCR, respectively (see online supplementary materials and methods for details).

Cell proliferation analysis

HepG2 cells were transfected with miR-21-ASO, scrambled control or miR-21-ASO plus HBPI- siRNA using lipofectamine 2000 (Invitrogen). After 48 h, cell proliferation was determined using a MTT (3-(4,5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazoliumbromide) cell proliferation kit (Cell Biolabs) according to the manufacturer’s instruction (see online supplementary material and methods for details).

Focus formation assay and flow cytometry analysis

HepG2 cells were used to determine the effect of miR-21 on focus formation and cell cycle progression (see online supplementary material and methods for details).

Xenograft tumour assay

Male BALB/c athymic nude mice (6 weeks old) purchased from Jackson Laboratory were used to study the role of miR-21 in...
promoting growth of xenograft tumour from HepG2 cells (see online supplementary material and methods for details).

**Histological analysis**
Frozen sections of liver were stained with Oil Red-O staining. Paraformaldehyde-fixed, paraffin-embedded sections of liver were stained with H&E (see online supplementary material and methods for details).

**Lipid and lipoprotein analysis**
Both plasma and hepatic lipid content were enzymatically measured in liver lysates and plasma via a colorimetric assay using a triglyceride assay kit from Roche Diagnostics, according to the manufacturer’s protocols (see online supplementary material and methods for details).

**Western blot and Q-RT-PCR**
Western blot and qRT-PCR were used to determine expression levels of genes. Primers used for quantitative RT-PCR are listed in online supplementary table S4 (see online supplementary material and methods for details).

**Statistical analysis**
Statistical analysis was performed using GraphPad Prism Software. Data derived from cell-line experiments were presented as mean±SD and assessed by a two-tailed Student’s t test. Statistical difference for cell cycle progression analysis was evaluated using χ² test. Mann–Whitney test was used to evaluate the statistical significance for mouse experiments. Each experiment was repeated at least three times; and the error bars represent the SD. p<0.05 was considered to be statistically significant.

**RESULTS**

**miR-21 is robustly induced in livers of mice on HFD and HepG2 cells exposed to high levels of fatty acid**
Hepatocytes are the major cells that control lipid metabolism and the primary site of NAFLD and HCC. To investigate the role of miRNAs in both disorders, we compiled hepatocyte-specific miRNA profiles by comparing miRNAs expression of livers of hepatocyte-specific Dicer1 knockout (DKO) and wild-type mice (see online supplementary table S5). We observed that miR-21 was the most significantly downregulated miRNA in livers of DKO mice (≥39-fold reduced), indicating that hepatocytes represent a main source of its expression in the liver (figure 1A). To assess the role of miR-21 in NAFLD, we fed wildtype C57Bl/6 mice an HFD (see online supplementary figure S1A–C) and measured its hepatic expression. The results showed that miR-21 had a twofold upregulation in the livers of HFD-treated mice (figure 1B), suggesting its potential role in NAFLD.

We also determined whether fatty acids can increase the expression of miR-21 in human hepatoma HepG2 cells. Oleic acids are the most abundant unsaturated fatty acids in liver triglycerides in both normal subjects and patients with NAFLD.\(^\text{21}\) In this study, HepG2 cells were used because of their increased sensitivity to fat accumulation.\(^\text{21}\) Nile-Red staining revealed that oleic acid treatment led to an increase in intracellular lipids in HepG2 cells (figure 1C, D), which was also associated with upregulation of miR-21 (figure 1E). Taken together, our in vivo and in vitro studies indicated that both HFD treatment of mice and exposure of HepG2 cells to fatty acid were able to induce expression of miR-21.

To further elucidate the role of miR-21 in hepatic lipid accumulation, we began to identify target genes of miR-21 by combining mRNA profiling of livers of NAFLD individuals with the bioinformatic prediction of miR-21 binding motifs within potential target mRNAs. This led us to identify 13 genes including HBP1, SOX7 and RHOB that showed reduced expression in human fatty liver and contained a conserved binding motif for miR-21 (see online supplementary table S3). GO analysis of the above 13 genes revealed that HBP1 was a potent tumour suppressor by preventing G1/S transition of cell cycle.\(^\text{22}\)\(^\text{23}\) In addition, our prediction from in silico algorithms showed that the 3’ UTR of HBP1 mRNA was 100% complementary to the miR-21 3’ seed region, exhibiting the highest prediction score and binding energy (figure 2A). Therefore, we selected HBP1 as a potential target of miR-21.

![Figure 1](http://gut.bmj.com/)

**Figure 1** Hepatocyte-specific miR-21 is significantly induced in livers of high fat diet (HFD)-treated mice and HepG2 cells treated with fatty acid. (A) qRT-PCR confirmed that miR-21 was highly expressed in hepatocytes of liver. miR-21 expression in liver was compared between Dicer1 knockout (DKO) (n=3) and wildtype mice (n=3) using qRT-PCR. Data are presented as mean±SD (p=0.0016, Student’s t test). (B) HFD treatment led to higher levels of miR-21 in livers of mice. Briefly, wildtype mice (C57Bl/6) were maintained on HFD for 4 weeks, and then mice were sacrificed and livers collected for miRNA expression analysis. Control mice received standard diet (SD). Data are presented as mean±SD (p=0.0022, Mann–Whitney test). (C and D) Oleate treatment increased lipid content and subsequently (E) expression of miR-21 in HepG2 cells. HepG2 cells were maintained in Dulbecco’s modified Eagle medium containing 0.5 mM oleate. Data are presented as mean±SD. In this multiple-groups experiment, we only performed comparison between two groups and Student’s t test was used for statistical analysis. Lipid droplets in human hepatocytes were labelled with arrows.

---

We next determined its expression in the livers of dietary obese mice treated with miR-21-ASO. It was not surprising that \textit{Hbp1} expression increased more than twofold in the livers of miR-21-ASO-treated mice compared with those treated with miR-21-MM-ASO (figure 2B). Taken together, hepatic expression of miR-21 was increased in dietary obese mice and livers of human patients with NAFLD/NASH, and \textit{Hbp1}, as a potential target of miR-21, showed reduced expression in livers of obese mice and human patients with NAFLD (see online supplementary table S3 and figure S1D). Our findings suggested that the crosstalk between miR-21 and \textit{Hbp1} might play an important role in hepatic lipid accumulation.

\textit{Hbp1} is a direct target of miR-21

To establish that miR-21 directly recognises the predicted target site within the 3' UTR of \textit{Hbp1}, the 3' UTR of mouse \textit{Hbp1} mRNA was cloned into a luciferase reporter vector (pMiR-Report) to generate pMiR-\textit{Hbp1}. Mouse Hepa1,6 cells were transfected with pMiR-\textit{Hbp1} and chemically synthesised miR-21 mimic or miR-21 inhibitor. We found that miR-21 mimics significantly downregulated luciferase activity in a dose-dependent fashion (figure 2C). Conversely, transfection with a miR-21 inhibitor antagonised the binding of miR-21 mimics to the 3' UTR of mouse \textit{Hbp1}, which was reflected by increased luciferase activity. Data are presented as mean±SD. Student's t test was used for statistical analysis. (E) Mutated binding motif for miR-21 within \textit{Hbp1} 3' UTR impaired miR-21 binding, which was reflected by (i) negligible change of luciferase activity after miR-21 mimics treatment; and (ii) miR-21 inhibitor treatment had no effect on luciferase activity. Data are presented as mean±SD (p≥0.1, Student's t test).

\textit{Hbp1} is a direct target of miR-21

To establish that miR-21 directly recognises the predicted target site within the 3' UTR of \textit{Hbp1}, the 3' UTR of mouse \textit{Hbp1} mRNA was cloned into a luciferase reporter vector (pMiR-Report) to generate pMiR-\textit{Hbp1}. Mouse Hepa1,6 cells were transfected with pMiR-\textit{Hbp1} and chemically synthesised miR-21 mimic or miR-21 inhibitor. We found that miR-21 mimics significantly downregulated luciferase activity in a dose-dependent fashion (figure 2C). Consistently, miR-21 inhibitor antagonised the inhibitory effect of miR-21 mimics on luciferase activity (figure 2D). Furthermore, we mutated the binding motif for miR-21 within the pMiR-\textit{Hbp1} 3' UTR and found that both
mimics and inhibitors of miR-21 had no effect on luciferase activity (figure 2E), indicating a potentially direct interaction between miR-21 and Hbp1 mRNA. To further validate that Hbp1 is a target of miR-21, we increased intracellular levels of miR-21 in HepG2 cells in the absence of fatty acid. qRT-PCR and western blot revealed that miR-21 significantly inhibited expression of HBP1 (figure 2F). In contrast, miR-21 knockdown led to an increase in mRNA and protein levels of HBP1 in HepG2 cells (figure 2G). Together, these results confirmed that Hbp1 is a direct target of miR-21.

HBP1 inhibits expression of SREBP1C, CCND1 and CCNB1 by activating p53

HBP1 is a well-described transcriptional repressor that modulates expression of genes involved in cell cycle progression.24 Therefore, we overexpressed HBP1 in HepG2 cells and determined the expression levels of genes involved in cell cycle using Human Cell Cycle RT² Profiler PCR Assay. Interestingly, we observed that p53 was the most upregulated after overexpression of HBP1 (figure 3A). p53 functions as a tumour suppressor and potent inhibitor of lipogenesis by inhibiting transcription of SREBP1C.24–26 leading to our hypothesis that miR-21 plays roles in both lipogenesis and carcinogenesis by interacting with the HBP1-p53 pathway.

Overexpression of HBP1 led to increased mRNA levels of p53, implying that HBP1 might activate transcription of p53 by binding to its promoter. Therefore, we cloned the p53 promoter into a luciferase reporter vector (pGL3-Basic) and generated pGL3-p53. Hepa1,6 cells were transfected with pGL3-p53 and HBP1 expression vector. As expected, overexpression of HBP1 induced luciferase activity (figure 3B), and HBP1 knockdown led to decreased luciferase activity (figure 3C). Furthermore, HBP1 knockdown impaired expression of endogenous p53 (figure 3D), suggesting that HBP1 was able to activate transcription of p53. HBP1 can function as a transcriptional activator by binding to a specific binding motif (GGGATGGG).22 However, we did not identify this binding motif within the promoter of p53, signifying that HBP1 might activate transcription of p53 by interacting with other transcription factors that have binding sites within the p53 promoter.

Srebp1c is a transcription factor that activates genes encoding enzymes required for lipid synthesis.27 28 Considering the role that p53 plays in inhibiting lipogenesis by modulating Srebp1c,26 we cloned the mouse Srebp1c promoter into pGL3-basic vector (pGL3-Srebp1c). As expected, co-transfection of p53 expression vector pGL3-Srebp1c and into Hepa1,6 cells significantly reduced luciferase activity in a dose-dependent fashion (see online supplementary figure S2A), and in contrast, p53 knockdown induced luciferase activity (see online supplementary figure S2B). Furthermore, we observed that overexpression of p53 led to a decrease in endogenous mRNA levels of SREBP1C and its targeted lipogenic genes SCD1 (stearyl-CoA desaturase-1), GPAT (glycerol 3-phosphate acyltransferase), and FASN (fatty acid synthase),28 and genes controlling cell cycle progression including CCNB1 and CCND1 in HepG2 cells (figure 3E).30 p53 knockdown led to the opposite effect (figure 3F), underscoring the central role of p53 in modulating the expression of genes involved in lipogenesis and cell cycle progression.

To further determine whether HBP1 prevents transcription of genes associated with lipogenesis and G1/S transition via p53-SREBP1C pathway, we overexpressed HBP1 in HepG2 cells and determined expression levels of p53, SREBP1C, the lipogenic genes and CCNB1 and CCND1. As confirmed by qRT-PCR, HBP1 overexpression led to increased p53, which subsequently prevented expression of SREBP1C and SCD1, FASN and GPAT, as well as CCNB1 and CCND1 (figure 3G). In summary, our findings suggested that HBP1 is able to simultaneously inhibit expression of CCNB1, CCND1 and SREBP1C by modulating p53.

miR-21 prevents expression of p53 but promotes transcription of SREBP1C by modulating HBP1 expression

We have shown that HBP1 is a target of miR-21, and HBP1 can activate transcription of p53. Meanwhile, p53 is a transcriptional repressor of Srebp1c.25 31 Thus, we hypothesised that miR-21 can simultaneously regulate expression of genes involved in lipogenesis and the G1/S transition by modulating the HBP1-p53-SREBP1C pathway. Indeed, overexpression of miR-21 in HepG2 cells inhibited expression of HBP1, which subsequently led to a reduction in p53 and an increase in mRNA levels of CCND1, CCNB1 and SREBP1C, as well as its target genes including SCD1, FASN and GPAT (figure 3H), while miR-21 knockdown led to an opposite effect (figure 3I). Our findings indicated that miR-21 is able to modulate expression of genes controlling lipogenesis and cell cycle progression via the p53-SREBP1C pathway.

miR-21 modulates lipid accumulation in HepG2 cells by interacting with the HBP1-p53 pathway

We then determined whether overexpression of miR-21 can promote lipogenesis. As expected, miR-21 overexpression prevented expression of HBP1 and p53 (figure 4A), which subsequently promoted lipid accumulation in HepG2 cells (figure 4B, C).

To determine loss of function for miR-21 in lipid accumulation, we transfected miR-21 inhibitor into oleate-treated HepG2 cells to knock down upregulated miR-21. Antagonising miR-21 led to a significant increase in HBP1 and p53 (figure 4D), which subsequently prevented lipid accumulation (figure 4E, F). These data demonstrated that miR-21 was sufficient for the downregulation of HBP1 and p53, which subsequently induced lipid accumulation. To further investigate the role of the interaction between miR-21 and HBP1 in hepatic lipid accumulation, we mutated the binding motif for miR-21 within the 3′ UTR of Hbp1 in the pMiR-Hbp1 (referred to as pMiR-Hbp1Mu) and introduced the pMiR-Hbp1 or pMiR-Hbp1Mu into oleate-treated HepG2 cells. Since oleate treatment increases miR-21 expression in HepG2 cells, it was expected that it would lead to a decrease of luciferase activity in HepG2 cells transfected with pMiR-Hbp1 compared with pMiR-Hbp1Mu. In fact, oleate treatment of HepG2 cells transfected with pMiR-Hbp1 resulted in robust repression of luciferase activity compared with pMiR-Hbp1-Mu (figure 4G). Together, our results indicated that HBP1 is a direct target of miR-21 during lipid accumulation in HepG2 cells and the crosstalk of miR-21 with HBP1 and p53 plays an important role in hepatic lipid accumulation.

Inhibitory effect of miR-21-ASO on hepatic lipid accumulation is mediated by HBP1

To confirm that miR-21 promotes lipogenesis via HBP1, we initially antagonised miR-21 by transfecting miR-21-ASO into HepG2 cells to induce expression of HBP1, and then knocked down the induced HBP1 using HBP1-siRNA. The results showed that miR-21 knockdown increased p53 and HBP1 and reduced lipid content in HepG2 cells, but additional treatment of HBP1-siRNA offset the effect of miR-21-ASO (figure 5A–C), which suggested that HBP1, in part, mediated the inhibitory effect of miR-21 inhibitor on lipid accumulation. To study the
Figure 3 HBP1 is a transcriptional activator of p53. (A) Overexpression of HBP1 increased mRNA levels of p53 in human HepG2 cells. Data are presented as mean±SD (*p=0.0003; Student’s t test). (B) Overexpression of HBP1 caused dose-dependent increase of the activity of a luciferase reporter gene linked to the p53 promoter. Data are presented as mean±SD. Student’s t test was used for statistical analysis. (C) HBP1 knockdown via its siRNA decreased luciferase activity driven by p53 promoter. Data are presented as mean±SD. Student’s t test was used for statistical analysis. (D) HBP1 knockdown via its siRNA decreased mRNA levels of p53. Data are presented as mean±SD (*p=0.0040; Student’s t test). (E) Overexpression of p53 led to decreased SREBP1C, lipogenic genes SCD1, FASN and GPAT as well as CCND1 and CCNB1 in HepG2 cells. Data are presented as mean±SD (*p<0.05; **p<0.001; Student’s t test). (F) Knockdown of p53 via its siRNA increased mRNA levels of SREBP1C, FASN, SCD1, CCND1 and CCNB1 in HepG2 cells. Data are presented as mean±SD (*p<0.05; **p<0.001; Student’s t test). (G) Overexpression of HBP1 reduced expression of p53, which subsequently led to decreased expression of genes involved in lipogenesis and G1/S transition. Data are presented as mean±SD (*p<0.05; **p<0.001; Student’s t test). (H) Overexpression of miR-21 inhibited HBP1 and p53, which subsequently promoted expression of the lipogenic genes including SCD1, FASN and GPAT and the genes controlling cell cycle progression including CCNB1 and CCND1. Data are presented as mean±SD (*p<0.05; **p<0.001; Student’s t test). (I) miR-21 knockdown via its inhibitor led to increased HBP1 and p53 and decreased expression of the lipogenic and cell cycle-related genes. Data are presented as mean±SD (*p<0.05; **p<0.001; Student’s t test).

role of the crosstalk between p53 and miR-21 in lipogenesis, we transfected oleated-incubated HepG2 cells with miR-21 mimics or a combination of miR-21 mimics and p53 expression vector. Nile-Red staining and qRT-PCR revealed that miR-21 mimics promoted lipid accumulation in HepG2 cells and the additional p53 overexpression rescued the effect of miR-21 (see online supplementary figure S3A, B). We further determined whether p53 deletion could offset the inhibitory effect(s) of miR-21-ASO on lipogenesis. The results showed that miR-21 knockdown led to decreased lipid content and increased p53, and additional treatment with p53 siRNA offset the effects of miR-21-ASO (see online supplementary figure S3C-E). Taken together, our results indicated that miR-21-induced lipid accumulation is, in part, mediated by HBP1 and p53.

HBP1 mediates the inhibitory effect of miR-21 inhibitor on proliferation, G1/S transition and xenograft tumour from HepG2 cells

To determine whether HBP1, at least in part, mediates the effects of miR-21 on proliferation and G1/S transition, we adopted a similar strategy as described above. MTT and soft agar colony assays revealed that miR-21-ASO administration caused suppression of cellular proliferation in HepG2 cells, but additional treatment of HBP1-siRNA counteracted the effects of miR-21-ASO (figure 5D, E). Cell cycle phase distribution of HepG2 cells further showed that miR-21 knockdown led to a significant increase in G1 phase cells and G1/S arrest, but additional treatment of HBP1-siRNA antagonised this effect of miR-21-ASO (figure 5F). Consistent with our in vitro findings, miR-21 knockdown inhibited growth of xenograft tumours in nude mice, and HBP1-siRNA treatment counteracted the effect of miR-21-ASO (figure 5G).

As we proposed, p53 is an important mediator of the miR-21-Hbp1-p53 axis. To determine whether p53 mediates the inductive effect of miR-21 on proliferation, we treated HepG2 cells with miR-21 mimic or a combination of miR-21 and p53 expression vector. MTT assay, soft agar colony formation assay and cell cycle analysis revealed that miR-21 promoted proliferation and cell cycle progression, and additional overexpression of p53 offset the inductive effects of miR-21 (see online supplementary figure S4A–E). Furthermore, miR-21-ASO treatment led to reduced proliferation, delayed G1/S transition and repressed growth of xenograft tumour, and the additional treatment of p53 siRNA rescued the inhibitory effects of miR-21-ASO (see online supplementary figure S4F–K). In summary, our findings indicated that HBP1 mediates, at least in part, the inhibitory effects of miR-21-ASO on G1/S transition, proliferation and growth of xenograft tumour.
Antagonising miR-21 in dietary obese mice prevents hepatic lipid accumulation

Next, we assessed the functional contribution of increased Hbp1 and p53 expression to hepatic lipid accumulation by reducing miR-21 expression in dietary obese mice. C57Bl/6 mice, which had been on an HFD, were injected with either miR-21-ASO or miR-21-MM-ASO for 4 weeks. We observed that antagonising miR-21 significantly reduced levels of triglycerides in livers of HFD-treated animals (figure 6A, B), in contrast to plasma triglyceride levels (figure 6C). On the other hand, miR-21-ASO treatment had no effect on body and liver weight (see online supplementary table S6). As expected, we also observed a 91% reduction of hepatic miR-21 expression in mice that received miR-21-ASO compared with miR-21-MM-ASO and a twofold increase of Hbp1 and p53 (figure 6D). Four-week treatment of miR-21-ASO showed a strong inhibitory effect on
Figure 6 Antagonising miR-21 prevented hepatic lipid accumulation in high fat diet (HFD)-treated mice. (A and B) miR-21 knockdown inhibited lipid accumulation in livers of HFD-fed mice injected with miR-21-anti-sense oligonucleotide (ASO). Representative images are shown. Lipid droplets in livers are labelled with arrows. Cellular triglyceride content was measured by Oil Red staining and triglyceride content was measured with a triglyceride estimation kit. Data are presented as mean±SD (p<0.0002, Mann–Whitney test). (C) miR-21 knockdown had no effect on plasma triglyceride in HFD-treated mice. Data are presented as mean±SD (p>0.1, Mann–Whitney test). (D) miR-21-ASO injection into dietary obese mice resulted in downregulated miR-21 and increased Hbp1 and p53 expression. Data are presented as mean±SD. Mann–Whitney test was used for statistical analysis. (E) qRT-PCR revealed that HFD-treated mice with decreased levels of miR-21 also retained reduced expression of Scd1, Fasn and Gpat after miR-21-ASO injection. C57Bl/6 mice at 8 weeks of age were kept on HFD for an additional 8 weeks. At 12 weeks of age, mice received a dose of 25 mg/kg miR-21-ASO or miR-21-MM-ASO (0.9% NaCl) weekly for 4 weeks via tail vein injection. Data are presented as mean ±SD. Mann–Whitney test was used for statistical analysis.
hepatosteatosis, but there were no differences in liver and body weight. Therefore, we increased miR-21-ASO treatment time to 8 weeks, which resulted in decreases in both liver weight (see online supplementary table S7) and hepatic lipid content (see online supplementary figure S5B). However, no difference in body weight (see online supplementary table S7), serum free fatty acid and glycerol still was observed (see online supplementary figure S6A, B). In addition, we also observed that miR-21-ASO treatment had no effect on plasma liver enzymes (see online supplementary table S8). These findings indicated that the crosstalk of miR-21 with Hbp1 plays an important role in hepatosteatosis.

We further compared expression levels of Srebp1c and lipogenic genes in livers of miR-21-ASO and miR-21-MM-ASO treated mice. As expected, miR-21-ASO treatment led to a significant reduction in Srebp1c in the livers of HFD-treated mice (figure 6E). Reduction of Srebp1c due to miR-21 knockdown should impair expression of the lipogenic genes. Indeed, in the miR-21-ASO treated group, the mRNA of three enzymes including Scd1, Fasn and Gpat was downregulated at least 1.5 times those of controls (figure 6D). Thus, the reduction of Srebp1c was associated with a dramatic reduction in the expression of the target enzymes responsible for lipogenesis, which prevented hepatic lipid accumulation. qRT-PCR also revealed that miR-21 knockdown had no effect on β-oxidation-related genes including Cpt1a, Acc2 and PGC1α (see online supplementary figure S6C, D).

In summary, our data have shown that HFD treatment led to increased miR-21, decreased Hbp1 and p53, which subsequently promoted hepatic lipid accumulation and the potential for HCC, whereas antagonising miR-21 led to the opposite and more therapeutic effect. Hbp1 is inhibited with increased miR-21 levels and its knockdown impaired transcription of p53 by Hbp1, which led to reduced p53. The loss of p53 then resulted in increased transcription of Srebp1c, CCNB1 and CCND1, which promoted both lipogenesis and cell replication (figure 7). Our findings indicate the miR-21 plays an important role in linking NAFLD to HCC by interacting with the Hbp1-p53-Srebp1c pathway.

**DISCUSSION**

Our study addresses a potentially important role for miR-21 in the development of NAFLD and HCC and defines a novel mechanism by which miR-21 contributes to lipogenesis and carcinogenesis via the Hbp1-p53-Srebp1c pathway. The observation that antagonising miR-21 in dietary obese mice potentially improves these metabolic parameters clearly indicates a functional role for increased miR-21 expression in the development of NAFLD. In addition, we also observed that miR-21 knockdown prevented an in situ model of tumorigenesis by targeting Hbp1. Increased miR-21 expression is not restricted to murine obesity models of NAFLD and HCC, but is also detected in human patients with NASH and HCC. Thus, miR-21-ASO may act as a unique potential therapeutic approach for the treatment of both disorders.

Despite its putative role in carcinogenesis, the mechanism by which miR-21 regulates NAFLD is unknown. In this study, we observed that HFD administration resulted in increased miR-21 and its knockdown prevented hepatic lipid accumulation. In addition, we have functionally validated Hbp1 as a bona fide target of miR-21, and Hbp1 is a transcriptional activator of p53. It is known that p53 acts as both a tumour suppressor and inhibitor of lipogenesis by inhibiting Srebp1c. Our findings, combined with those of others, indicate that miR-21 plays critical roles in the pathogenesis of NAFLD and HCC. In addition, miR-21 deletion in p53 knockout mice reduced the incidence of liver cancer; and p53 deletion alone promoted hepatosteatosis, further supporting the unique role for miR-21 in linking NAFLD to HCC via the Hbp1-p53-Srebp1c pathway. miR-21 significantly inhibits expression of PTEN, a negative regulator of NASH in mice, suggesting that it might, in part, mediate the inhibitory effect of miR-21 on hepatosteatosis. In this study, we found that Hbp1 is a direct target of miR-21 and confirmed that Hbp1 modulates the inhibitory function of miR-21-ASO on hepatosteatosis and carcinogenesis simultaneously.

We have shown that miR-21 is highly expressed in hepatocytes (figure 1A). Meanwhile, miR-21 is increased but its target Hbp1 is reduced in livers of HFD-treated mice and human individuals with NAFLD and HCC. All these features of miR-21 lead us to focus on its role in linking NAFLD to HCC. Although other miRNAs might contribute to the development of NAFLD and HCC, few other hepatocyte-specific miRNAs meet the above criteria like miR-21. These include increased expression in both NAFLD and HCC of human and mouse, reduced expression of their target genes in mouse models and human patients, and high expression in hepatocytes. We demonstrated that miR-21 knockdown led to upregulated Hbp1 and p53, downregulated Srebp1c and decreased expression of hepatic Scd1, Gpat and Fasn. These findings are consistent with earlier reports that p53 knockout promoted Srebp1c transcription and in turn increased expression of Scd1, Gpat and Fasn, and subsequently hepatic lipogenesis, suggesting that miR-21-dependent Hbp1-p53 pathway inhibition of Srebp1c transcription represents a candidate pathway to cause NAFLD.

There is evidence to suggest that inhibition of p53 attenuates steatosis and liver injury; which is inconsistent with our findings and those of others. One possible explanation is that p53 is a negative regulator of hepatic lipid accumulation in the early stages of NAFLD, but p53 is highly expressed at the advanced stages of NAFLD, which may contribute to the high level of apoptosis associated with NASH. In patients with liver steatosis without inflammation, p53 expression was significantly lower than in steatohepatitis, further suggesting that p53 plays different roles in the various developmental stages of NAFLD. Although the mechanism(s) by which miR-21 controls lipogenesis and carcinogenesis in liver clearly requires further investigation, our study has identified an important role for the interaction of miR-21 with Hbp1 in obesity-induced NAFLD and HCC.
Hepatology

Contributors HW and RN: acquisition of data. XC and CJS: analysis and interpretation of data. GS: obtaining funding, study supervision, study concept and design and drafting of the manuscript.

Funding Supported in part from grants received from the NIDDK R01 (1R01DK102601-01), Minnesota Medical Foundation, NIH Clinical and Translational Science Award at the University of Minnesota (ULTR000114).

Competing interests None declared.

Ethics approval All procedures involving mice were approved by the Institutional Animal Care Committee at the University of Minnesota, University of California San Francisco, and the Agency for Science Technology and Research Singapore.

Provenance and peer review Not commissioned; externally peer reviewed.

Open Access This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

REFERENCES
MicroRNA-21 is a potential link between non-alcoholic fatty liver disease and hepatocellular carcinoma via modulation of the HBP1-p53-Srebp1c pathway

Heng Wu, Raymond Ng, Xin Chen, Clifford J Steer and Guisheng Song

*Gut* published online August 17, 2015

Updated information and services can be found at: http://gut.bmj.com/content/early/2015/08/17/gutjnl-2014-308430

These include:

**Supplementary Material**

Supplementary material can be found at: http://gut.bmj.com/content/suppl/2015/08/18/gutjnl-2014-308430.DC1

**References**

This article cites 42 articles, 8 of which you can access for free at: http://gut.bmj.com/content/early/2015/08/17/gutjnl-2014-308430#BIBL

**Open Access**

This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

**Email alerting service**

Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

**Topic Collections**

Articles on similar topics can be found in the following collections

- Open access (354)
- Hepatic cancer (474)
- Nonalcoholic steatosis (127)

**Notes**

To request permissions go to: http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to: http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to: http://group.bmj.com/subscribe/