Long term serological surveillance after treatment of *Helicobacter pylori* infection

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Abstract

Fifteen patients with type B gastritis caused by *Helicobacter pylori* infection were treated with 'triple' therapy consisting of colloidal bismuth subcitrate, amoxycillin, and metronidazole. All were followed up as outpatients every three months for at least one year. After 'triple' therapy a significant ($p < 0.01$) and persistent reduction in IgA and IgG antibody levels against *H. pylori* was detected. In three patients recurrent active infection with *H. pylori* at nine and 12 months was detected by a rise in IgA (three patients) and IgG (two patients) antibody levels against *H. pylori* and worsening of symptoms, and was confirmed by culture and histology. In 11 patients, the absence of infection at 12 months was confirmed by culture and histology. In a control group of 13 patients with type B gastritis who received no antibacterial treatment, specific IgA and IgG antibody levels against *H. pylori* remained unchanged during 12 months of follow up. Although specific IgG against *H. pylori* is the most widely used serological test for screening, our data indicate that specific IgA is also valuable in monitoring treatment. These serological tests are easy to perform, relatively inexpensive, devoid of radioactivity and are very acceptable to patients. It is concluded that serological testing is the preferred method for follow up after treatment for *H. pylori* infection and will probably replace endoscopy or the urea breath test.

Helicobacter pylori is a curved spiral Gram negative bacterium that lives in the human stomach. Since the first description of *H. pylori*,¹ it has been established as the cause of type B gastritis^{2,3} and as an important pathogenic factor in duodenal ulcer disease.^{4,5} Recently, eradication of *H. pylori* infection has been proposed as the new mainstay of duodenal ulcer treatment.⁵

There are several sensitive and specific tests available for the diagnosis of *H. pylori*. The presence of the bacterium in gastric biopsy specimens can be detected directly by culture⁶ and histology^{7,8} or indirectly by using the strong urease⁹ produced by the bacterium. For screening and follow up after treatment non-invasive tests are preferable. Urea breath tests are reliable and non-invasive but require an expensive mass spectrometer or produce radioactive waste with a long half-life.^{10,11}

The presence of specific IgA and IgG antibodies against *H. pylori* can be shown by the relatively inexpensive and generally available technique of enzyme linked immunosorbent assay (ELISA).¹²⁻¹⁴ We followed two groups of

patients – treated and untreated for *H. pylori* infection – to see whether monitoring of treatment by means of serology was informative with regard to eradication of *H. pylori*.

Patients and methods

Fifteen patients, whose mean age was 52 years, and male to female ratio (M:F) 1.5, with dyspeptic symptoms, positive *H. pylori* culture, and histological evidence of type B gastritis, were treated with triple therapy consisting of bismuth subcitrate 120 mg qds for 28 days, amoxycillin 500 mg qds, and metronidazole 500 mg qds for 10 days. All patients had raised serum antibodies against *H. pylori* determined by ELISA.

The patients have been followed as outpatients, initially at six weeks and then every three months for at least one year. Thirteen patients (mean age 61 years, M:F 5.5) with *H. pylori* positive gastritis who received no antibacterial treatment were also followed by means of serology every three months for a year. This group, with high antibody values confirming *H. pylori* infection, served as control group. In this non-randomised study controls were selected whose antibody values matched those of the patients receiving treatment and who shared the same ethnic background.

Patients who took antibiotics or bismuth-containing drugs before the study or during follow up were excluded. The controls received maintenance treatment with H_2 antagonists.

At the three monthly visits, symptoms were recorded and blood was taken for serological tests. Endoscopy (Olympus, GIF K10) was performed before treatment, and at three and 12 months afterwards. Additional endoscopies were performed if there was no improvement in symptoms, when symptoms recurred, or when there was an appreciable rise in antibody values. Each patient underwent an average of 4.2 endoscopies. Follow up was stopped when infection with *H. pylori* was diagnosed. Controls underwent an endoscopy at 12 months of follow up.

At upper gastrointestinal endoscopy, biopsy specimens were taken from the antrum with a sterilised biopsy forceps, 2 cm proximal to the pylorus in intact mucosa. One specimen was transported to the laboratory in 0.2 ml sterile 0.9% NaCl and inoculated by rubbing onto the surface of a blood agar plate (Blood Agar Base No 2, Oxoid CM 271, containing 5% sheep blood) and onto Skirrow's medium. The cultures were incubated at 37°C in a nitrogen atmosphere containing 8% CO₂ and 6% O₂ for five days. The bacteria were identified as *H. pylori* on the basis of their morphology, oxidase, catalase, and urease production. Another specimen was immediately

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Figure 1: Development of specific IgA anti-*H. pylori* values (mean (SEM)) in 11 patients (broken line) before and for 12 months after triple therapy without signs of *H. pylori* infection at 12 months, expressed as an absorbance index (AI) compared with the absorbance index of the 13 controls (solid line) without triple therapy. * $p < 0.01$.

fixed in 10% buffered formalin, embedded in paraffin and cut into 4 μ m thick serial sections, which were stained with haematoxylin and eosin. For the present study, infection with *H. pylori* was diagnosed if both active gastritis and bacteria were present.

IgA and IgG specific antibodies against *H. pylori* were measured by a modified ELISA technique using conjugates labelled with immunoperoxidase specific for human IgA and IgG. For standardisation of the measurement of these antibodies, test conditions were chosen: the mean (SD) values for absorbance of the standard reference serum was taken as 0.5 (0.1) for IgA and 1.0 (0.1) for IgG. These values were used to correct the absorbance given by the sera under study.

The absorbance index (AI) was calculated from the mean of two readings of optical density (OD) of serum. The results were expressed as follows:

$$AI = \frac{\text{Patient's OD} - \text{OD of blank reading}}{\text{Reference OD} - \text{OD blank reading}}$$

An AI > 0.32 for anti-*H. pylori* IgG and an AI > 0.35 for anti-*H. pylori* IgA was considered evidence of infection.¹² The antigen preparation and determination intra- and inter-assay variability of the ELISA technique has been described in detail.¹²

Wilcoxon's rank sum test was used for statistical analysis.

Results

Thirty subjects (15 in each group) entered the study, but two patients in the triple therapy group failed to meet follow up appointments and were withdrawn. During treatment, side effects of nausea and diarrhoea were frequent but mild, and stopped when treatment ended: no patients dropped out because of these.

H. pylori eradication was confirmed by culture and histology in 14 patients three months after beginning treatment.

The ELISA results for specific IgA and IgG anti-*H. pylori* antibodies in the triple therapy and control groups are shown in Figures 1 and 2. Thirteen patients in the control group showed no variation in the mean AI for IgA and IgG specific antibodies. Those patients in the triple therapy group without evidence of *H. pylori* infection at 12 months (n=11) showed a significant decrease in the mean AI for both specific IgA and IgG anti-*H. pylori* antibodies.

Twelve months after the start of triple therapy, the AI for specific IgA was ≤ 0.35 , which is considered the upper limit of normal¹² in five out of the 11 patients without evidence of *H. pylori* infection at that time. After 12 months, the AI for specific IgG was ≤ 0.32 (considered the upper limit of normal¹²) in only one out of these 11 patients. Nine of the 11 patients are still without evidence of *H. pylori* infection 18 months after starting treatment. In six of these nine the AI for specific IgA was ≤ 0.35 and in four the AI for specific IgG was ≤ 0.32 , illustrating a slow but persistent decrease in AI for both immunoglobulins after 12 months of follow up. After treatment, recurrent active *H. pylori* infection and active gastritis were confirmed by culture and histology in three patients (20%) in whom worsening symptoms and a rise in specific IgA anti-*H. pylori* antibodies occurred. In two of these patients a rise in specific IgG anti-*H. pylori* antibody was also found. Individual values of their AI are shown in the Table.

Although it was not the aim of the study, the effect of treatment on symptoms was also monitored. In five patients, symptoms resolved after triple therapy: in the remaining 10, there was no change or only a short term improve-

Figure 2: Development of specific IgG anti-*H pylori* levels in 11 patients before and after triple therapy expressed as an absorbance index (AI) compared with the 19 controls without triple therapy. * $p < 0.01$.

Individual values for specific IgA and IgG anti-*H pylori* (expressed as absorbance index) in three patients with recurrent active *H pylori* infection after treatment with triple therapy

Patient No		Before treatment	At 6 weeks	At 3 months	At 6 months	At 9 months	At 12 months
1	IgA	0.81	—	0.46	0.33	0.52*	—
	IgG	0.81	—	0.65	0.58	0.67*	—
2	IgA	2.56	1.44	1.57	1.37	—	1.48*
	IgG	0.66	0.63	0.48	0.42	—	0.27*
3	IgA	0.86	0.60	0.62	0.53	0.86*	—
	IgG	0.63	0.59	0.57	0.50	0.78*	—

*Infection with *H pylori* confirmed by culture or histology.

ment. In the three patients whose symptoms recurred and who showed an increase in specific IgA anti-*H pylori*, infection with *H pylori* was detected. In six patients, symptoms returned without a concurrent rise in specific IgA or IgG anti-*H pylori*. In these patients, *H pylori* infection was not detected by culture or histological examination of gastric biopsy specimens (which showed minimal chronic gastritis only) nor was any abnormality found during endoscopy.

Discussion

Since the detection of *H pylori* and its association with chronic active gastritis,¹ rapid developments have taken place. Not only is *H pylori* now generally accepted as the cause of type B gastritis but there is also mounting evidence that it is one of the most important factors in duodenal ulcer disease.^{2,3,15,16} With the development of effective treatment regimens, long term eradication of the bacterium is possible in over 90% of patients.¹⁷ Using triple therapy against *H pylori*, Rauws and Tytgat have recently suggested that a permanent cure for duodenal ulcer disease may be within reach.⁵

Development of non-invasive tests for evaluating the effect of treatment on *H pylori* in larger numbers of patients has somewhat lagged behind these rapid developments in the field of thera-

peutics. There are several excellent tests for the detection of *H pylori* in untreated patients.¹² After treatment, however, when the number of bacteria in the mucosa is substantially reduced, the number of false negative test results may increase.

Determination of specific IgA and IgG anti-*H pylori* antibodies by means of an ELISA technique is suitable for screening and follow up in larger patient populations, and several authors have reported a high sensitivity and specificity for diagnosing untreated *H pylori* infection.¹²⁻¹⁴

There are few well documented studies in which patients have been followed by serological means for longer than six months. In our study, we found that after triple therapy, eradication of the bacterium was reflected in a rapid and significant ($p < 0.01$) decrease in specific IgA and IgG anti-*H pylori* antibody values at six weeks. Specific IgG anti-*H pylori* antibody values showed a more gradual decrease than those specific for IgA. This rapid decrease in specific IgA anti-*H pylori* antibodies agrees with the results reported in two short term follow up studies.^{18,19}

In our study, recurrence of *H pylori* infection was indicated by a rise in specific IgA antibody values in three patients and a rise in specific IgG antibody levels in two of them. In all patients with recurrent or persisting complaints, endoscopy and serological examination was performed. Only when recurrent complaints were accompanied by a rise in specific IgA anti-*H pylori*, was infection with *H pylori* diagnosed. In most patients with no signs of *H pylori* infection at 12 months, both specific IgA and specific IgG antibody values were lower but had not yet returned to the normal range. At the end of the year, however, five patients had normal IgA anti-*H pylori* values, and one had a normal IgG value as well.

We conclude that determination of specific

IgA and IgG antibodies against *H pylori* is a suitable method of monitoring treatment against *H pylori* and probably in diagnosing recurrent infection as well. Follow up of longer than 12 months is needed to establish whether a true serological cure of *H pylori* infection by triple therapy is possible, and whether it will result in normalisation of specific IgA and IgG anti-*H pylori* antibody values in all patients. Longer follow up is also needed to determine whether low antibody values against *H pylori* will eventually result in reinfection.

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