PANCREAS

Role of interleukin-6 in mediating the acute phase protein response and potential as an early means of severity assessment in acute pancreatitis

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Abstract
A number of laboratory and clinical studies have shown that interleukin-6 is the principal mediator of the acute phase protein response. In this study the relationship between serum concentrations of interleukin-6 and C-reactive protein in acute pancreatitis are examined and the ability of interleukin-6 to discriminate between severe and mild attacks is assessed. We have studied 24 patients (10 severe and 14 mild). Serum samples were collected on admission, six hourly for 48 hours and then 12 hourly for a further three days. When the areas under the curves of individual patients were compared there was a strong correlation between the total production of interleukin-6 and C-reactive protein ($r=0.73$) (Spearman rank correlation) and peak interleukin-6 and C-reactive protein concentrations ($r=0.75$), suggesting a close relationship between interleukin-6 and C-reactive protein production. Both on admission and peak interleukin-6 concentrations were significantly higher in patients with severe than mild disease. There was no significant difference in on admission C-reactive protein concentrations, although significant differences were seen when peak concentrations were considered. Utilising a peak interleukin-6 concentration of >130 u/ml, we were able to distinguish between severe and mild attacks of acute pancreatitis with a sensitivity of 100% and specificity of 71%. These figures were comparable with those for peak C-reactive protein, a C-reactive protein of

>150 mg/l detecting severe attacks of acute pancreatitis with a sensitivity of 90% and specificity of 79%. In view of the fact that interleukin-6 concentrations peaked earlier than those of C-reactive protein, interleukin-6 is capable of providing comparable, but earlier severity prediction than C-reactive protein. (Gut 1993; 34: 41-45)

Our inability to identify correctly severe acute pancreatitis on admission to hospital using clinical examination alone¹⁻³ has led to the development of a number of more objective means of severity assessment. These include the Ranson⁴ and Glasgow⁵ scoring systems, APACHE II scoring system^{3,6} and a variety of single factors measured in serum.^{7,8} Recently the effectiveness of serum C-reactive protein concentration measurements in determining the severity of acute pancreatitis has been demonstrated.^{3,9-11} In common with the other methods described above, however, C-reactive protein measurements involve a delay of 48 hours or longer before prediction. One possible means of avoiding this delay is by measuring the serum concentrations of the principal mediator of the acute phase protein response: interleukin-6.¹²⁻¹⁴ In this study we examine the ability of interleukin-6 to provide early severity prediction in acute pancreatitis and in addition examine the relationship between interleukin-6 and the acute phase protein, C-reactive protein.

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Accepted for publication
8 June 1992

TABLE 1 The clinical details of 10 patients with severe and 14 with mild acute pancreatitis. GS=gall stones. There are more complications than patients due to the fact that a patient may have more than 1 complication

	Severe pancreatitis	Mild pancreatitis	Difference
Median age (years) (range)	51.5 (23-69)	44.5 (21-77)	95% CI=-13 to 20, $p<0.79$
Ratio of sexes M:F	8:2	11:3	NS $p<0.67$
Aetiology of attack			
Alcohol	6	6	NS $p<0.3$
Gall stones	3	7	
Unknown	1	1	
Median hospital stay (days) (range)	19 (7-25)	7.5 (6-22)	95% CI=9.6 to 1 $p<0.001$
Complications			
Respiratory failure	2	0	-
Renal failure	1	0	-
Hospital stay <14 days	5	0	-
Pancreatic pseudocyst	4	0	-

CI=confidence interval, NS=not significant.

Methods

PATIENTS

Twenty four patients with acute pancreatitis were prospectively entered into the study between December 1987 and July 1989. The diagnosis of acute pancreatitis was made on the basis of a serum amylase greater than 720 IU/l in the presence of a compatible clinical picture of the disease (a value of 720 IU/l being equivalent to a value of 1200 IU/l as measured by the Phadebas method). The clinical and aetiological details of the 24 patients entered into the study (14 mild and 10 severe) are summarised in Table I. The groups were comparable in terms of age, sex, and aetiology with hospital stay being significantly longer in patients with severe disease.

SPECIMEN HANDLING AND ANALYSIS

Samples of blood were taken at the time of admission, six hourly for 48 hours and then twice daily for a further three days. Specimens were centrifuged, separated, aliquoted, and stored at -20°C before analysis for C-reactive protein, and interleukin-6.

Measurements of serum amylase concentrations were made on a Hitachi 737 random access discrete analyser (Boehringer Mannheim, Germany) using an enzymatic colorimetric assay (α amylase PNP (Boehringer Mannheim, Germany)). C-reactive protein concentrations (normal <10 mg/l) were measured using a competitive binding, fluorescence polarisation immunoassay (TDX reagents and analyser, Abbott Diagnostic, England). Interleukin-6 concentrations were measured using the interleukin-6 dependent 7TD1 cell line,¹⁵ the rate of growth of which was calculated using a colorimetric substrate (a tetrazolium salt which was cleaved by dehydrogenase enzymes present in living cells).¹⁶ Standardisation was performed using recombinant interleukin-6¹⁷ which had been assigned a specific activity of 10^6 units per microgram – that is, 1 picogram of interleukin-6 was equivalent to 1 u. A normal serum interleukin-6 concentration was <10 u/ml.

DISEASE SEVERITY

Disease severity was graded retrospectively depending upon the clinical outcome. Patients were considered to have suffered a severe attack of acute pancreatitis if they developed one or more of the following: a pancreatic pseudocyst; pancreatic sepsis; respiratory failure (as judged by a $\text{pO}_2 < 8\text{ kPa}$ requiring oxygen for >5 days or assisted ventilation); renal failure (as evidenced by <400 ml urine per 24 hours after adequate fluid replacement); shock (defined as a systolic

blood pressure <90 mm of Hg in the presence of an adequate circulating fluid volume); a hospital stay of >14 days (except where discharge was delayed for cholecystectomy or social reasons); or death. Where none of the above was present the patient was considered to have suffered a mild attack.

DATA ANALYSIS

In order to examine the time course of interleukin-6 and C-reactive protein production in patients with acute pancreatitis, it was necessary to correlate serum concentrations from the time of onset of symptoms rather than from the time of admission. As it was not possible to control the delay between the onset of symptoms and admission to hospital, it was not possible to obtain samples at exactly corresponding tissue points from different patients. Samples were therefore allocated to one of eight groups. These were <24 hours, 25–36 hours, 37–48 hours, 49–60 hours, 61–72 hours, 73–96 hours, 97–120 hours and >121 hours representing the range of times between the onset of symptoms and the sample being taken.

Data were analysed using medians, the Mann Whitney U test, Fisher's exact test and the Spearman rank correlation test as appropriate. Confidence intervals were used in preference to p values in judging the relevance of differences.¹⁸

The area under the curve of interleukin-6 and C-reactive protein represents the magnitude of the response during the study period. The areas under curves (AUC) were calculated from the formula:-

$$\text{AUC} = \frac{1}{2} \sum_{i=0}^{n-1} (t_{i+1} - t_i) (y_i + y_{i+1})$$

Where t = the time after admission, y = the concentration at time t , i = initial time points and n represents the last time point.

The areas under the curves were divided by the number of hours in order to take account of the different times for which samples had been collected from individual patients.

Results

Serum concentrations of interleukin-6 rose during the first 24 hours after the onset of symptoms in all patients and peaked between 24 and 36 hours (Fig 1). The magnitude of the interleukin-6 response (represented by the area under the response curve) differed between patient groups. In patients with severe disease the median response was 240 u/ml/h (range 72–560 u/ml/h) and in the mild group 38 u/ml/h (range 18–140 u/ml/h). These differences were highly significant (95% confidence interval (CI) = 92 to 232 u/ml/h, $p < 0.001$). Peak interleukin-6 concentrations correlated well with the magnitude of the interleukin-6 response ($r = 0.94$) and can therefore be considered to represent accurately the overall response.

C-reactive protein concentrations rose during the first 24 hours to peak between 36 and 48 hours after the onset of symptoms (Fig 1). The magnitude of the C-reactive protein response

Figure 1 Median interleukin-6 and C-reactive protein concentrations in the serum of 10 patients with severe and 14 with mild acute pancreatitis.

Figure 2 Integrated interleukin-6 and C-reactive protein response in 10 patients with severe and 14 with mild acute pancreatitis.

differed between patient groups. In patients with severe disease the median response was 147.5 mg/l/h (range 69–273 mg/l/h) and in the mild group 47.5 mg/l/h (range 2–120 mg/l/h). These differences were highly significant (95% CI=59 to 147 mg/l/h, $p<0.001$). Peak C-reactive protein concentrations correlated well with the magnitude of the C-reactive protein response ($r=0.93$). There was a significant correlation between the magnitude of the interleukin-6 and C-reactive protein responses ($r=0.73$, Figure 2) and between peak interleukin-6 and C-reactive protein concentrations ($r=0.75$, Figure 3). On admission and peak concentrations of interleukin-6 were significantly greater in patients with severe than mild disease (95% CI=33.9 to 129 u/ml, $p<0.008$ and 95% CI=134 to 593, $p<0.005$) respectively.

On admission concentrations of C-reactive protein were not significantly greater in patients with severe than mild disease (CI= -96 to 4 mg/l, $p<0.125$) although significant differences were

Figure 3 Peak serum interleukin-6 and C-reactive protein concentrations in 10 patients with severe and 14 with mild acute pancreatitis.

noted when peak concentrations were considered (95% CI=74 to 214 mg/l, $p<0.0006$). The sensitivity, specificity, positive and negative predictive values and percentage correct for a peak interleukin-6 concentration of >130 u/ml and a C-reactive protein concentration of >150 mg/l are displayed in Table II.

Discussion

Experimental studies conducted in animals, hepatoma cell lines, and normal human hepatocyte cultures have shown that interleukin-6 is the principal mediator of the acute phase protein response of which C-reactive protein is an important component.¹²⁻¹⁴ In the present study the median delay between the onset of symptoms and peak interleukin-6 concentrations was between 24 and 36 hours and is in agreement with the clinical studies of Van Oers *et al*¹⁹ but is somewhat longer than those of Nijsten *et al*,²⁰ Shenkin *et al*²¹ and Cruickshank *et al*.²² Van Oers *et al*¹⁹ noted peak serum interleukin-6 concentrations on the second day post renal transplantation (presumed delay between transplantation and the measurement of interleukin-6 concentration 36–48 hours) and Nijsten *et al*²⁰ noted peak concentrations on admission in burns patients (presumed delay <six hours). Shenkin *et al*²¹ noted peak concentrations of interleukin-6 between 1.5 and four hours after the skin incision was made in a group of patients undergoing elective cholecystectomy and Cruickshank *et al*²² six to 12 hours after the skin incision for a variety of surgical procedures. As Nijsten *et al*²⁰ tell us little about the severity of the burns (if patients had only mild burns the peak interleukin-6 concentration would be small and short lived) and only took samples on a daily basis, it is probable that peak interleukin-6 concentrations have been missed. Further evidence that peak interleukin-6 concentrations may occur later in inflammatory disease comes from animal work. Geiger *et al*²³ injected one group of rats with human recombinant interleukin-6 and one with turpentine (a means of inducing an acute inflammatory reaction) and then compared the time to maximal mRNA production for a number of acute phase proteins. In those injected with interleukin-6 peak concentrations of acute phase protein mRNAs were seen within four hours, whereas there was a delay of 16–24 hours in those injected with turpentine. While peak mRNA concentrations will precede those of the proteins for which they code these findings are in better agreement with those of the present study and of Van Oers *et al*¹⁹ than of Nijsten *et al*²⁰ and the other workers quoted above. On the basis of this and the studies quoted above the timing of peak interleukin-6 concentrations appears to be somewhat variable. The late peaks probably relate to a continuing inflammatory process, in contrast with operative trauma which was short lived.

Of more importance than the exact time of peak interleukin-6 concentrations is the relationship between peak interleukin-6 and C-reactive protein concentrations. In the present study the delay to peak C-reactive protein concentrations was 36 to 48 hours. In Nijsten *et al*²⁰ study

TABLE II Sensitivity, specificity, positive predictive value, negative predictive value and percentage correct for serum concentrations of interleukin-6 and C-reactive protein in seventy predictions in acute pancreatitis

	Sensitivity %	Specificity %	Positive predictive value %	Negative predictive value %	% Correct
Admission interleukin-6 >120 IU/ml	70	79	88	79	75
Admission C-reactive protein >120 mg/l	30	100	100	59	54
Peak interleukin-6 >130 IU/ml	100	71	71	100	83
Peak C-reactive protein >150 mg/l	90	79	75	92	83
Glasgow score >2	50	92	83	72	75

peak C-reactive protein concentrations were noted on the first post admission day (presumed delay between interleukin-6 and C-reactive protein peak 24 hours) and are in keeping with the results for this study population. Similar results were achieved by Shenkin *et al*²¹ who noted peak C-reactive protein concentrations between 36 and 48 hours and Cruickshank *et al*²² who noted peak C-reactive protein concentrations between 24 and 48 hours. These results are also in keeping with the findings of laboratory based studies. The addition of human recombinant interleukin-6 to hepatoma cell lines has been shown to produce a maximal acute phase protein response between 20 and 36 hours.^{24 25} When interleukin-6 was added to adult rat hepatocyte cultures the delay was 24 hours²⁶ and when injected into rats 16–24 hours.²³ The delay to peak C-reactive protein concentrations in patient with acute pancreatitis in the present study are in agreement with the work of the group from Leeds⁹ and our own previous study.³ Both Buchler *et al*¹¹ and Poulakkainen *et al*,¹⁰ however, reported peak C-reactive protein concentrations at the time of admission. Many of their referrals were tertiary and probably had already undergone a delay of 48 hours or longer before C-reactive protein measurements.

The significant correlation noted between the integrated interleukin-6 and C-reactive protein responses ($r=0.73$) and between peak interleukin-6 and C-reactive protein concentrations ($r=0.75$), coupled with the fact that the median peak interleukin-6 concentrations preceded those of C-reactive protein provides support for the hypothesis that interleukin-6 is acting as a mediator of C-reactive protein production. Interleukin-6 may not be the only mediator of the acute phase protein response, however. When Morrone *et al*²⁷ added recombinant interleukin-6 to Hep 3B cell line they were able to induce partially the gene governing C-reactive protein production. Complete induction was only achieved when monocyte supernatant was added as well. Baumann *et al*²⁸ have demonstrated that the optimal acute phase protein response in Hep G2 human hepatoma cell lines is only produced in the presence of interleukin-1 and dexamethasone and both Castell *et al*^{13 14} and Moshag *et al*¹² have demonstrated that interleukin-1 augments the action of interleukin-6 in producing C-reactive protein in human hepatocyte cultures. It therefore seems likely that at least one of the factors inducing C-reactive protein production in

Morrone *et al*'s monocyte supernatant was interleukin-1.²⁷

C-reactive protein has been widely adopted as a non-specific indicator of inflammation because serum concentrations rise more rapidly and to a greater degree than any other acute phase protein. Its usefulness in this role has been shown in medical²⁹ and surgical³⁰ conditions, including acute pancreatitis.^{3 9-11} It does, however, have a major drawback in that it entails a similar delay to the other methods of severity assessment in acute pancreatitis.^{3 9 10 11} The measurement of interleukin-6 concentrations may overcome this problem and facilitate earlier severity prediction. Serum interleukin-6 concentrations rose more rapidly than those of C-reactive protein. This earlier rise in interleukin-6 concentrations resulted in significant differences in on-admission interleukin-6 concentrations between patients, an on admission serum concentration >120 u/ml separated severe and mild attacks with a sensitivity of 70% and specificity of 79%. These figures are comparable with figures previously reported for the multifactorial scoring systems and provide clinically useful separation between patient groups.^{4 5} Such differences were not seen for serum C-reactive protein on admission, a serum concentration of >120 mg/l being unable to distinguish effectively between patient groups. These findings are in agreement with the studies of Mayer⁹ and Wilson *et al*,⁶ as discussed above. Better separation between the patient groups was noted when peak concentrations were considered, the sensitivity and specificity of interleukin-6 and C-reactive protein being comparable, and somewhat better than previous reports of the multifactorial scoring systems.^{4 5}

Speculation that interleukin-6 may be useful in clinical practice as a severity indicator in acute pancreatitis is dependent on results being available within a few hours of sample collection. The major drawback of the interleukin-6 bioassay used here is the long turn around time of five days. Several commercial manufacturers, however, have recently developed and marketed immunoassays for interleukin-6 (Quantikine Human interleukin-6, British Bio-technology Ltd, Abingdon, Oxon; Biokine interleukin-6 test kit, T Cell Sciences, Cambridge; Co-eliza interleukin-6, Kabi Diagnostica, Sweden). These assays have turnaround times of less than six hours and consequently, the methodology is now available to produce interleukin-6 results within the time scale necessary for them to be of value clinically. It should be noted, however, that we have no direct experience of any of these assays.

We would like to thank Dr J Van Snick for providing 7TD1 cell line and Dr L Aarden for providing recombinant interleukin-6.

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