PANCREAS

Tumour suppression induced by the macrophage activating lipopeptide MALP-2 in an ultrasound guided pancreatic carcinoma mouse model

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Background and aim: Carcinoma of the exocrine pancreas has a particularly poor prognosis. Therefore, novel therapeutic strategies such as immunotherapy are required. Here we investigated the immunomodulatory capacity of macrophage activating lipopeptide 2 (MALP-2), which binds to toll-like receptors 2 and 6 and induces activation of nuclear factor κ B in monocytes. This causes the release of early stage leucocyte attracting chemokines and proinflammatory cytokines.

Methods: MALP-2 was tested in a new orthotopic ultrasound guided pancreatic cancer mouse model. This model is close to the biological situation and avoids the stress and immunostimulation caused by laparotomy. Cells from the syngeneic, highly aggressive, and metastatic cell line Panc 02 were administered orthotopically, by ultrasound guidance, to C57bl/6 mice. MALP-2 was administered intratumorally or intraperitoneally and tumour growth, immune status, and leucocyte infiltration at the tumour site were determined.

Results: We showed a tumour suppressive effect induced by a single injection of MALP-2. Median survival increased from 21 to 30 days ($p < 0.002$). Combining chemotherapy (gemcitabine) with MALP-2 treatment caused further prolonged survival (median survival 27 days with chemotherapy alone v 37 days for combined treatment; $p < 0.0002$). The life prolonging effect was paralleled by a significant increase in cytotoxic T cells, restoration of $\beta 2$ integrin expression on lymphocytes, and high expression of CD45RB on T helper cells. Immunohistochemical stains showed strong cytotoxic T lymphocyte and natural killer cell infiltration.

Conclusions: In conclusion, in a model of orthotopic pancreatic cancer in mice, we induced a tumour suppressive effect by treatment with a synthetic lipopeptide. Treatment with MALP-2 could be an option for immunotherapy in pancreatic cancer.

Ductal adenocarcinoma of the pancreas has a particularly poor prognosis. Five year survival is only 3–5%. Radical pancreaticoduodenectomy, especially for minimal disease, is currently the only chance of cure. Although chemotherapy has led to improvements in survival in patients with locally advanced disease, the overall effect is small.¹ Also, surgery or radiotherapy of locally advanced unresectable pancreatic cancer does not lead to significantly prolonged survival. Thus exocrine pancreatic carcinoma is a neoplasm where various therapies have had demonstrable although small effects.

In this respect, immunotherapy could be another strategy for the treatment of cancer of the pancreas. Immune surveillance is the mechanism by which the immune system monitors and destroys developing malignancies. Although T cells seem to be the main effectors of immune surveillance, the innate immune system (which includes natural killer (NK) cells, macrophages, monocytes, and mast cells) is also involved. Malignant cells evade immunosuppression by downregulating intrinsic immunogenicity.² Stimulation of the immune system could help to overcome the unresponsiveness and anergy in cancer patients. Jaffee *et al* showed in preclinical studies in a murine tumour model that tumour cell vaccines engineered to secrete GM-CSF in a paracrine fashion elicited systemic immune responses that were capable of eliminating established pancreatic tumours as long as they were small.³ These authors were further able to

show safety and antitumour immunity using this approach in a phase I trial.⁴

Some immunomodulators show adjuvant activity by inducing cytokine secretion from monocytes or macrophages (for example, interferon α , interleukin (IL)-12, and tumour necrosis factor α (TNF- α)). Adjuvants can be the danger signals which are necessary to stimulate dendritic cells. Dendritic cells, if appropriately induced, ultimately lead to Th1 dominance resulting in specific cell mediated immunity.

Such approaches may be especially helpful when tumour masses are still low or in combination with other strategies. Thus Weisenthal *et al* showed that adjuvant supported tumour kill was dramatically more effective when applied to tumours obtained from patients who had undergone previous chemotherapy. The authors speculated that response to chemotherapy produces massive release and processing of tumour antigens, and that this response leads to a state in which the immune system is primed (via *in situ* vaccination) to respond to exogenous macrophage activation signals with potent specific antitumour effects.⁵

Abbreviations: MALP-2, macrophage activating lipopeptide 2; NK, natural killer; iNOS, inducible nitric oxide synthase; TLR, toll-like receptor; IL, interleukin; TNF- α , tumour necrosis factor α ; PBS, phosphate buffered saline

One plausible candidate for successful application as a strong immunomodulating substance in this context is the macrophage activating lipopeptide of 2 kDa molecular weight (MALP-2).⁶ MALP-2 is a synthetic lipopeptide with two long chain fatty acid ester residues which signals through toll-like receptors 2 (TLR-2) and 6 (TLR-6),^{7,8} activates nuclear transcription factor κ B,⁹ induces the synthesis of a number of cytokines and chemokines, depending on its target cell,¹⁰ and also induces maturation of dendritic cells.¹¹ Moreover, MALP-2, formerly called MDHM, induces in vitro tumoricidal activity of macrophages. It is also highly active in vivo as it induces leucocyte infiltration after intraperitoneal administration in mice or after intratracheal administration in rats.¹² In this system it reduces formation of metastases in the lung.¹³ MALP-2 was shown to exhibit adjuvant properties when applied intranasally.¹⁴ Studies in BALB/c mice bearing methyl cholanthrene induced fibrosarcoma showed strong haemorrhagic necrosis with complete healing 10 days after MALP-2 application.¹⁵

In this study, the adjuvant and tumour suppressive capacity of MALP-2 was investigated in a syngeneic pancreatic cancer mouse model.

MATERIALS AND METHODS

MALP-2

MALP-2 was synthesised as described previously⁶ and kept as a stock solution of 1 mg/ml in water-2 propanol (1:1) at 4°C. The exact peptide content was determined by amino acid analysis. Stock solutions were first diluted with 25 mM octyl glucoside in saline to provide a carrier and optimal solubilisation, and were then further diluted in several steps with culture medium. The biological activity of MALP-2 was tested by nitric oxide release, as described elsewhere¹⁶: 1 U of activity corresponds to approximately 3 pg of MALP-2.

Mice and cell lines

Panc 02 is a murine ductal pancreas adenocarcinoma cell line which was established in 1984 in female C57BL/6 mice, as described previously. After 3-methylcholanthrene induction, a ductal pancreatic adenocarcinoma was isolated and subcutaneously passaged in the syngeneic host.¹⁷ Panc 02 cells are positive for Ki-Ras, p53, Her2NEU, and CDK, as determined by real time polymerase chain reaction. The cells were a kind gift from V Schmitz (University of Bonn, Germany). Mycoplasma free Panc 02 cells were cultured in RPMI 1640 with 10% fetal calf serum and incubated at 37°C with 5% CO₂ in a humidified atmosphere.

Male and female C57BL/6 mice (Charles River, Sulzbach, Germany) were kept and treated in accordance with the principles laid down in the European Community's Council Directives, and approved by the local administration (reference 50.203.2-BN22). Eight week old mice were used for the experiments.

Subcutaneous tumour model

Panc 02 cells (5×10^5) in 50 μ l phosphate buffered saline (PBS) were administered subcutaneously in the back of eight week old C57BL/6 mice. The animals received a single intraperitoneal injection of 8×10^5 U (approximately 3 μ g) of MALP-2 in saline two days before, on the same day or two days after tumour inoculation. Control mice received injections containing control vehicle without MALP-2. Each group consisted of six mice. Growth of the subcutaneous tumours was determined using a vernier calliper. Mice were sacrificed when tumours were ulcerating or exceeded 150 mm².

Orthotopic ultrasound guided tumour model

For ultrasound imaging, a commercially available ultrasound machine was used (HDI 5000; Philips-Ultrasound, Bothell,

Washington, USA) equipped with a high frequency linear array transducer designed for intraoperative use (CL 15–7). Imaging was performed in a real time compound imaging mode (SonoCT) operating at 10 MHz, emitting at 10 MHz, and receiving at 10 MHz. In the SonoCT mode, the digital beamformer electronically steers the transducer array about nine steering angles. For artefact reduction, frames acquired at each angle were averaged online, allowing real time imaging at frame rates of up to 10 Hz. Line density was set to high. Persistence was disabled. The dynamic range was set to 170 dB and compression was 4. After anaesthesia by isoflurane, animals were placed in the supine position. The abdomen was carefully shaved. Transverse horizontal imaging planes were obtained and modified to allow visualisation of the long axis of the pancreas. To achieve the greatest benefit from real time compound imaging, potential focal lesions were placed in the central region of the image to take advantage of the complete component frame overlap (see fig 1). Images were digitally stored for offline analysis as raw data files prior to scan conversion. Image analysis was performed using a calibrated software tool (HDI-Lab, Version 1.91, Philips-Ultrasound).

We injected 5×10^5 Panc-02 cells in 50 μ l PBS orthotopically in eight week old C57BL/6 mice. Mice received a single injection of 8×10^5 U MALP-2 intraperitoneally or near the pancreas (intratumorally) at different time points after tumour inoculation (12 mice were treated on day +2 and six mice were treated on day +9). Control mice were left untreated (18 animals) or received injections with vehicle control without MALP-2 (16 animals). Tumour growth was determined every second day by ultrasound in anaesthetised animals. Twelve mice were treated intraperitoneally with 2.5 mg/kg gemcitabine five times a week for two weeks starting on day 3 after tumour inoculation. Six mice additionally received 0.3 μ g MALP-2 on days +7 and +14.

Mice were sacrificed according to our local ethics considerations when two of three criteria were met: tumour larger than 6 mm, abdominal metastasis, or ascites formation.

Preparation of spleen leucocytes and flow cytometric analysis

Spleens from sacrificed mice were collected and single cells were passaged through a 40 μ m nylon mesh. They were centrifuged, resuspended in 10 ml of erythrocyte lysis buffer (8.29 g/l NH₄Cl, 1 g/l KHCO₃, 0.0371 g/l EDTA in Aqua dest.),

Figure 1 Ultrasound guided tumour cell inoculation. Tumour cells were inoculated into the pancreatic tail of C57BL/6 mice using SonoCT ultrasound guidance. The tip of the needle in the tail of the pancreas prior to implantation of Panc 02 cells into the pancreatic tail can be seen. The spleen was used for guidance.

and incubated at room temperature for 15 minutes. Cells were incubated on ice for 15 minutes with the appropriate antibodies and then washed with PBS/1% bovine serum albumin. Dual colour flow cytometric analysis was performed on a Coulter Epics XL Cytometer. Data from 30 000 cells were collected and analysed. Leucocytes were phenotyped with the following monoclonal markers: CD4, CD8, CD18, CD45RB, NK1.1, and a marker for granulocytes (all from Pharmingen, Hamburg, Germany). Negative controls consisted of leucocytes labelled with rat IgG (PE/FITC). Mean fluorescence was normalised to mean fluorescence of the control antibodies.

Histological staining

Specimens were fixed immediately in 4% paraformaldehyde (for 24 hours at 4°C) and then embedded in paraffin. Sections (4 µm) were cut from paraffin blocks, mounted on positively charged slides (Superfrost), and air dried at 42°C overnight. For immunohistochemistry, fresh paraffin sections were deparaffinised in xylene, rehydrated in graded alcohols, and washed in Tris buffer. Polyclonal antimouse rabbit antibodies were used for detection of antigens (CD-4, CD-8, NCAM, and NOS2). Primary antibodies were added (dilution 1:50; all antibodies from Santa Cruz, Heidelberg, Germany) and slides incubated overnight at 4°C. Slides were then processed on an immunostainer (TechMate 500; Dako, Hamburg, Germany).

Antibodies were detected using the ABC method (Dako) with 3-amino-9-ethylcarbazol as a visualising reagent. Sections were finally counterstained with haematoxylin, mounted in aqueous mounting media, and analysed by standard light microscopy. Replacement of the first antibody by PBS was used as a negative control to assess the specificity of the antibodies. Additionally, haematoxylin-eosin and Giemsa staining were performed.

Statistical analysis

Statistical significance of Kaplan-Meier plots was analysed using log rank test (SPSS version 10.0 for windows). Significance of tumour growth was determined by the Mann-Whitney U test. All other parameters were tested with paired *t* tests. Differences were considered statistically significant when *p* values were less than 0.05. Data are expressed as mean (SEM).

RESULTS

MALP-2 treatment of mice with subcutaneous tumours

In order to test the capacity of MALP-2 to reduce tumour load or growth, 5×10^5 Panc 02 cells were administered subcutaneously. Tumours grew in all animals. Groups of eight mice received approximately 0.3 µg of MALP-2 intraperitoneally on day +2 of tumour inoculation. This treatment was well tolerated. We observed significantly reduced tumour growth in mice which were treated with MALP-2 two days after tumour inoculation ($p < 0.03$; data not shown). These mice also showed prolonged survival compared with mice that had received vehicle control. Median survival was 23 days compared with 18–19 days in control mice.

Tumour growth in the orthotopic model

We then attempted to treat pancreatic tumours developing at their natural site. To this end we used SonoCT, a non-invasive model which has the advantage of performing real time assessment of injections (fig 1). Data were more consistent compared with measurements in the subcutaneous model (fig 2). Growth of pancreatic cancer was seen in all mice, with abdominal metastases in most cases. Liver metastasis was observed in 50% of all mice (fig 3A, B).

Figure 2 Growth kinetics of subcutaneous (SC) and orthotopic tumours in C57Bl/6 mice. Panc 02 cells (5×10^5) in 50 µl phosphate buffered saline were administered subcutaneously in the back or near the pancreas (via ultrasound guidance) of eight week old C57Bl/6 mice, respectively. Tumour size was determined at three day intervals using a vernier calliper or by ultrasound, respectively. Control mice received injections containing control vehicle on day +2. Results show data from 16–18 mice for the orthotopic model and from six mice in the subcutaneous system. Data are mean (SEM).

MALP-2 treatment of mice with orthotopic tumours

After establishment of the orthotopic model, we investigated the tumour suppressive effect of MALP-2 intraperitoneally, administered two days after tumour implantation. We found prolonged survival of MALP-2 treated mice. Median survival was 16 days in control mice versus 20 days after intraperitoneal treatment ($p < 0.02$, data not shown). This effect was enhanced by intratumoral injection of MALP-2. Median survival was 19 days in control mice versus 26 days after intratumoral treatment ($p < 0.004$) (fig 4A).

Interestingly, treatment with MALP-2 nine days after tumour inoculation instead of two days produced even better results. When we treated mice with a detectable tumour burden (approximately 7.6 (1.6) mm²) with the same amount of MALP-2, median survival increased from 21 days for mice treated on day +2 to 30 days (fig 4B) ($p < 0.002$).

For further optimisation of the MALP-2 regimen, we treated mice once a week intratumorally with the lipopeptide. We observed no life prolonging effect compared with single administration (median survival 23 days, data not shown). There was no significant reduction in the number of metastases in MALP-2-treated animals. Most animals (27/29) had to be sacrificed because of tumour sizes exceeding 6 mm and concomitant formation of ascites.

Combination of MALP-2 with gemcitabine treatment

We next examined whether MALP-2 had a beneficial effect on chemotherapy treated tumour bearing mice. Therefore, mice were treated intraperitoneally with 2.5 mg/kg gemcitabine five times a week for two weeks starting on day 3 after tumour inoculation. One group of mice also received 0.3 µg MALP-2 on days +7 and +14.

Chemotherapy treated mice showed prolonged survival, as shown in fig 5. Median survival increased from 18 days for untreated animals to 29 days ($p < 0.0001$). Mice who had also

Figure 3 Growth and metastasis of orthotopically implanted Panc 02 cells. Mice were killed 21 days after orthotopic injection of Panc 02 cells. Growth of pancreatic cancer (A) was seen in all mice, with abdominal metastases in most cases. Liver metastasis (B) was observed in 50% of all mice.

received the lipopeptide survived a median of 37 days (fig 5). This increase in survival time was highly significant ($p < 0.0002$) compared with the control group who received gemcitabine and vehicle control alone.

Flow cytometric analysis

In order to gain insight into the possible mechanisms that could explain the above findings, we examined leucocyte composition in the spleens of healthy, tumour bearing, treated, and untreated mice.

Generally, we observed immunosuppression in tumour bearing mice. The number of B lymphocytes was significantly lower in tumour mice ($p < 0.001$) (fig 6A) and the $\beta 2$ integrin CD18, important for lymphocyte adhesion, was also significantly downregulated in mice with pancreatic tumours ($p < 0.02$). As a marker of regulatory T cells, we measured CD4⁺CD45RB^{low} cells in both groups. Mean expression of CD45RB on T helper cells was found to be strongly reduced in tumour mice ($p < 0.002$). Furthermore, there was a trend towards reduction of NK cells, CD4⁺ cells, and CD8⁺ cells in tumour bearing mice but this was not statistically significant and we could not rule out the possibility that we had some contamination with other cell types (for example, litoral cells) in our spleen preparations.

Flow cytometric analysis of murine spleen cells from animals treated intraperitoneally or intratumorally with MALP-2 on day +2 or day +9 showed a significant increase in CD18 expression ($p < 0.024$) (fig 6B). CD8⁺ lymphocytes increased significantly in all groups of MALP-2 treated mice, and particularly in the group with orthotopically implanted tumours treated on day +9 with MALP-2 ($p < 0.00003$)

Figure 4 Macrophage activating lipopeptide 2 (MALP-2) treatment of mice with orthotopic tumours. (A) Survival curve of C57Bl/6 mice with orthotopic syngeneic pancreatic carcinoma. Mice were treated with 8×10^5 U MALP-2, or with the corresponding control, intratumorally on day +2 after tumour inoculation. Results show data from at least 12 mice. Mice were sacrificed when tumour size exceeded 6 mm. (B) Survival curve of C57Bl/6 mice after MALP-2 treatment on day +9. Mice were treated with 8×10^5 U MALP-2, or with the corresponding control, intratumorally on day +9 after tumour inoculation. Results show data from six mice. Mice were sacrificed when tumour size exceeded 6 mm. * $p < 0.05$.

(fig 6B). Similar results were observed regarding the number of NK cells. A significant increase in NK cells was found in mice which were treated nine days after tumour inoculation ($p < 0.006$) (fig 6B). Mean fluorescence of CD45RB on T helper cells, an indicator of Th1/Th2 balance,¹⁸⁻²³ was significantly suppressed in tumour bearing mice and normalised or overexpressed in MALP-2 treated mice ($p < 0.02$ for intraperitoneal treatment) (fig 6b). The increase in CD45RB levels on T helper cells in MALP-2 intratumorally treated mice (on day +2) compared with the control group was also significant ($p < 0.0003$). MALP-2 treatment on day +9 did not cause an increase in CD45RB mean fluorescence, a phenomenon we cannot explain.

Mice having undergone chemotherapy alone showed a similar expression profile as untreated tumour bearing mice but there was strong immunomodulation in mice treated with gemcitabine plus MALP-2. In this group, we observed a statistically significant increase in the percentage of CD18⁺CD3⁺ (97.6% v 70.2%; $p < 0.02$), CD4⁺ (12.2% v 9.3%; $p < 0.05$), NK 1.1⁺ (18.1% v 4.2%; $p < 0.0001$), and CD19⁺ cells

Figure 5 Combined treatment with gemcitabine and macrophage activating lipopeptide 2 (MALP-2) intratumorally (it). Survival curve of C57Bl/6 mice with orthotopic syngeneic pancreatic carcinoma. Mice were treated with 2.5 mg/kg gemcitabine intraperitoneally five times a week for two weeks starting on day 3 after tumour inoculation. Additionally, one group of mice received 0.3 µg MALP-2 on day +7 and day +14. Results show data from at least six mice. Mice were sacrificed when tumour size exceeded 6 mm. * $p < 0.05$.

(78.5% *v* 30.9%; $p < 0.0005$) as well as in mean expression of CD45RB on T helper cells (207.7 *v* 20.3; $p < 0.00001$), all compared with the respective vehicle control (data not shown).

Histological staining of tumours

In general, tumours were mostly poorly differentiated with an almost spindle cell-like morphology. In contrast with the control group, MALP-2 treated mice showed progressive, sometimes confluent, tumour cell necrosis with obvious infiltration of lymphocytes.

Lymphocytes were immunohistochemically characterised as T lymphocytes and NK cells by expression of CD4, CD8, and NCAM, respectively (fig 7A, B). The zones in the tumours with a high density of leucocytes had a correspondingly high expression of inducible nitric oxide synthase (iNOS). Infiltration of lymphocytes was more pronounced in the group treated with MALP-2 intratumorally compared with mice treated intraperitoneally.

DISCUSSION

Animal cancer models can only approximate the situation in a human cancer patient. Each model has its specific limitations and pitfalls. We used a murine *in vivo* model with syngeneic, highly aggressive, metastatic tumour cells carrying genetic aberrations typical of a ductal pancreatic carcinoma. We studied the effects of the immunomodulator MALP-2 alone or in combination with chemotherapy in two experimental setups. Tumour cells were either implanted into the skin or orthotopically injected into or near the pancreas. Orthotopic tumour implantation and observations were done under minimally invasive conditions, allowing us to investigate the influence of immunotherapy on tumour growth without interfering with the immune system from surgical stress or ensuing infections. Although the skin model as an experimental system is easier to handle, and treatment effects are more easily observable, the orthotopic model more closely mimics the complex situation in an affected patient. Compared with studies requiring repeated laparotomies, this

Figure 6 Flow cytometric analysis of spleen derived leucocytes. Mice with orthotopic tumours were sacrificed and the spleens were immediately removed. Leucocytes were stained with the indicated antibodies. (A) Expression of various markers in healthy and tumour bearing mice. (B) Percentage of $\beta 2$ integrin positive cells, cytotoxic T lymphocytes, natural killer (NK) cells, and mean expression of CD45RB on T helper cells from tumour bearing mice after macrophage activating lipopeptide 2 (MALP-2) treatment intraperitoneal (ip) or intratumorally (it). Results show data from at least six mice. Data are mean (SEM). * $p < 0.05$.

method allows us to survey *in situ* tumour growth and to gather highly reproducible data compared with conservative measurement (Ziske, submitted).

MALP-2 treatment was beneficial in both experimental systems and significantly prolonged the survival time of tumour bearing mice. Combined therapy of MALP-2 with the ribonucleotide reductase inhibitor gemcitabine was particularly effective. Intraperitoneal as well as intratumoral treatment with MALP-2 were well tolerated but not equally effective. The life prolonging effect was more pronounced in the orthotopic than in the subcutaneous model, especially when animals were treated locally. Better circulation in intrapancreatic tumours and a higher initial local MALP-2 concentration may be responsible. However, even when MALP-2 was injected into the peritoneum, there were effects on the subcutaneous tumours at the back, suggesting a systemic MALP-2 effect.

MALP-2, as its acronym implies, is a lipopeptide. This class of immunomodulators signals through TLR, setting off a signal transduction cascade which results in activation of nuclear factor κB and release of proinflammatory cytokines. Specifically for MALP-2, the formation of nitric oxide, and secretion of CXC chemokines such as IL-8 and GRO-, leucocyte attracting CC chemokines such as MIP-1, MIP-2, and MCP-1, and proinflammatory proteins such as IL-6 and TNF- by activated monocytes and macrophages have been reported.^{9 10 24} MALP-2 differs from lipopeptides derived from or analogous to those from Gram negative bacteria in that it

Figure 7 Immunohistochemical staining of orthotopic tumours. Histological sections from a mouse treated with macrophage activating lipopeptide 2 (MALP-2) were stained with anti-CD8 (A) and anti-NCAM (B) antibodies. Photographs were made with 100 $\times$ magnitude.

carries only two fatty acid chains.⁶ As a consequence, MALP-2 signals through interactions between TLR-2 and TLR-6⁷ whereas “conventional” lipopeptides and their analogues signal via TLR-2 and TLR-1. TLR-1 is expressed in all leucocytes. In contrast with TLR-1, TLR-2 is expressed in myelomonocytic elements, including dendritic cells.²⁵ TLR-6 seems to be broadly expressed in the same manner as TLR-1 but the response to immunological agents after functional interaction with TLR-2 is different to the response that results after interaction of TLR-1 and TLR-2.²⁶ Different receptor specificity is expected to result in different responder cell populations as expression of TLRs is subject to regulation (see for example, Underhill²⁷).

Tumour suppressive effects of MALP-2 in different in vivo models were described earlier. Thus Shingu *et al* showed in a rat mammary adenocarcinoma model that intratracheal administration of MALP-2 reduced the number of lung metastases,¹³ and Galanos *et al* observed in Meth-A tumour bearing mice a strong haemorrhagic necrosis of subcutaneously growing fibrosarcomas after MALP-2 injection.¹⁵ This latter tumour is known to be TNF sensitive, and as TNF is induced by MALP-2, it is reasonable to ascribe the MALP-2 effect in this model primarily to circulating TNF. Fidler’s group described in a series of studies the effects of synthetic lipopeptides bearing three fatty acids (CGP 31362 or JBT 3002) in various tumour models.^{28–34} The observed tumour suppressive results correlated well with high levels of iNOS in the tumour leading to autolysis (for example, see Xie and Fidler³²). We do not know whether iNOS is upregulated by MALP-2 in Panc 02 cells used in this study but believe this is rather unlikely as Panc 02 cells do not express TLR-2 which is necessary for MALP-2 signalling (Märten, unpublished data). Similarly, direct cytotoxicity of MALP-2 on tumour cells can be ruled out as no apoptotic or cytolytic effect of MALP-2 could be observed in vitro (Märten, unpublished data). However, involvement of nitric oxide generated by effector cells such as infiltrating macrophages is likely. The following

observations indicate involvement of the innate as well as specific immune system.

- (i) MALP-2 caused infiltration of NK cells and cytotoxic T lymphocytes 16–37 days after MALP-2 injection, and these cells were localised close to and within the tumour (fig 7).
- (ii) After MALP-2 treatment, normalisation of expression of the β 2 integrin CD18 of leucocytes was observed (fig 6). Integrins are important molecules in contact dependent tumoricidal activity. Thus inhibition of macrophage mediated cytotoxicity was observed after blocking CD18.³⁵
- (iii) Similarly, an increase in expression of CD45RB was noted after MALP-2 (fig 6). High expression of CD45RB on lymphocytes seems to be associated with a Th1 secretion pattern while reduced expression is associated with regulatory T cells and Th2 cytokines.^{18–23} Our data thus suggest that MALP-2 regenerates Th2 shifted non-adhesive leucocytes.
- (iv) Although not demonstrated here, it was shown in cell cultures that MALP-2 causes activation and maturation of dendritic cells.¹¹ Such dendritic cells could take up apoptotic or necrotic tumour cells and prime specific effector cells.

CONCLUSION

Immunotherapy with MALP, particularly in combination with mild chemotherapy, could be an option for treatment of pancreatic cancer after surgical intervention. Even after resection of a ductal adenocarcinoma of the exocrine pancreas patients have an especially poor prognosis, minimal residual disease being the likely source of relapse. Another possible application for MALP-2 could be in the prevention of metastasis during resection. Further studies are required for optimisation of dosage and treatment schedules.

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