PROSTAGLANDINS AND CANCER

D Wang, R N DuBois

115

Gut 2006;55:115–122. doi: 10.1136/gut.2004.047100

SUMMARY

Chemoprevention has been considered as a possible approach for cancer prevention. A significant effort has been made in the development of novel drugs for both cancer prevention and treatment over the past decade. Recent epidemiological studies and clinical trials indicate that long term use of aspirin and similar agents, also called non-steroidal anti-inflammatory drugs (NSAIDs), can decrease the incidence of certain malignancies, including colorectal, oesophageal, breast, lung, and bladder cancers. The best known targets of NSAIDs are cyclooxygenase (COX) enzymes, which convert arachidonic acid to prostaglandins (PGs) and thromboxane. COX-2 derived prostaglandin E₂ (PGE₂) can promote tumour growth by binding its receptors and activating signalling pathways which control cell proliferation, migration, apoptosis, and/or angiogenesis. However, the prolonged use of high dosages of COX-2 selective inhibitors (COXIBs) is associated with unacceptable cardiovascular side effects. Thus it is crucial to develop more effective chemopreventive agents with minimal toxicity. Recent efforts to identify the molecular mechanisms by which PGE₂ promotes tumour growth and metastasis may provide opportunities for the development of safer strategies for cancer prevention and treatment.

INTRODUCTION

The most effective treatments for cancer, including various combinations of surgical resection, radiation, and/or chemotherapy, depend on the detection of cancer at a very early stage. Unfortunately, it has not been possible to identify all individuals who are at the highest risk for developing cancer. Most patients present to their physician with advanced cancer when standard treatment regimens for solid malignancies result in a much lower five year survival. It is generally agreed that an effective way to control cancer is to find better ways of preventing it. Chemopreventive approaches are definitely worth considering for healthy persons who have a strong family history of cancer or those who are particularly susceptible for other reasons. One promising group of compounds with cancer preventive activity includes NSAIDs.

A large body of evidence from population based studies, case control studies, and clinical trials indicate that regular use of NSAIDs over a 10–15 year period reduces the relative risk of developing colorectal cancer by 40–50%.¹ Furthermore, use of NSAIDs leads to regression of pre-existing adenomas in patients with familial adenomatous polyposis (FAP).² As many other human cancers are reported to have elevated levels of COX-2 and overproduce PGs, there is great interest in evaluating the role of NSAIDs for prevention and treatment strategies for other cancers such as breast, stomach, pancreas, urinary tract, lung, and prostate. However, the prolonged use of NSAIDs is associated with side effects such as nausea, dyspepsia, gastritis, abdominal pain, peptic ulcer, gastrointestinal bleeding, and/or perforation of gastroduodenal ulcers.³ It was hypothesised that NSAIDs exert their anti-inflammatory and antitumour effects through inhibition of the inducible COX-2,^{4,5} while unwanted side effects of these drugs such as damage to the gastric mucosa and gastrointestinal bleeding are thought to arise from the inhibition of the constitutive COX-1.⁶ This hypothesis led to the development of COXIBs, such as celecoxib, rofecoxib, and valdecoxib. Indeed, highly selective COX-2 inhibitors retain the anti-inflammatory and antitumour effects of the NSAIDs while not interfering with COX-1 responsible for protection of the gastroduodenal mucosa from the effects of acid from the stomach.^{5,7–9} Therefore, these drugs were approved by the FDA, and as novel anti-inflammatory agents their use is associated with about a 50% reduction in gastrointestinal toxicity. Moreover, these agents have some potential for use as chemopreventives.^{10–12} One of them, celecoxib, was approved in December 1999 by the FDA for use in the prevention of colorectal polyp formation of patients with FAP. Unexpectedly, the prolonged use of higher doses of COX-2 selective inhibitors is associated with increased thrombotic events in humans.^{13–15} We know that this original hypothesis was overly simplistic because of our lack of knowledge of the importance of signalling pathways which are

See end of article for authors' affiliations

Correspondence to:
Professor R N DuBois, The
Vanderbilt-Ingram Cancer
Center, Room 698, Preston
Research Building, 2300
Pierce Ave, Nashville,
Tennessee 37232-6838, USA;
raymond.dubois@
vanderbilt.edu

NSAID TARGETS Inflammation and cancer

NSAIDs are chemically distinct compounds that share a common therapeutic action. The best known of these are aspirin, ibuprofen, piroxicam, indomethacin, and sulindac. NSAIDs are generally prescribed to ameliorate symptoms associated with acute pain and chronic inflammatory diseases such as arthritis. Chronic inflammation caused by infections or autoimmune diseases is clearly associated with an increased cancer risk in a number of instances. For example, it has long been known that patients with persistent hepatitis B, *Helicobacter pylori* infections, or an immune disorder such as ulcerative colitis have a higher risk for the development of liver or gastrointestinal tract cancer. It has been estimated that chronic inflammation contributes to the development of approximately 15% of malignancies worldwide.¹⁶ The connection between inflammation and cancer further supports the concept that anti-inflammatory drugs, NSAIDs, have antineoplastic activity.

COX dependent targets

NSAIDs were shown to exert their anti-inflammatory, analgesic, and antipyretic effects mainly by inhibiting COX, a key enzyme responsible for the biosynthesis of PGs from arachidonic acid.⁶ Aspirin is an irreversible inhibitor of COX through blocking the approach of arachidonic acid, while indomethacin, piroxicam, ibuprofen, and sulindac are competitive inhibitors for substrate binding. When COX converts arachidonic acid to PGs, the key regulatory step in this process is the enzymatic conversion of arachidonate to PGG₂, which is then reduced to an unstable endoperoxide intermediate, PGH₂. Specific PG synthases in turn metabolise PGH₂ to at least five structurally related bioactive lipid molecules, including PGE₂, PGD₂, PGF_{2α}, PGI₂, and thromboxane A₂ (TxA₂)¹⁷ (fig 1).

COX exists in two isoforms commonly referred to as COX-1 and COX-2. Although both COX-1 and COX-2 are upregulated in a variety of circumstances, normally COX-1 is constitutively expressed in a broad range of cells and tissues. COX-1 expression remains constant under most physiological or pathological conditions and its constitutive enzymatic activity is linked to renal function, gastric mucosal maintenance, stimulation of platelet aggregation, and vasoconstriction. For example, COX-1 derived prostaglandins play a central role in many normal physiological processes. In the gut, COX-1 derived PGI₂ (also called prostacyclin) produced by epithelial and stromal cells in subepithelial tissues plays a key role in the cytoprotection of gastric mucosal surfaces and the normal vasculature.¹⁸ COX-1 has been shown to play an essential role in gastrulation in zebrafish and a reduction in COX-1 results in posterior mesodermal defects during zebrafish development.^{19–20} In contrast, COX-2 is an immediate early response gene and its expression is normally absent in most cells and tissues but it is highly induced in response to proinflammatory cytokines, hormones, and tumour promoters.²¹ Furthermore, COX-2 derived PGE₂ is a proinflammatory bioactive lipid and is the major prostaglandin produced in many human solid tumours, including cancer of the colon,²² stomach,²³ and breast.²⁴ Recent research has

indicated that COX-2 derived PGE₂ is a key mediator of acute inflammatory responses,^{25–26} arthritis,^{27–28} and inflammatory bowel disease.^{29–30} Direct evidence supporting the notion that PGE₂ promotes tumour growth comes from the following observations. PGE₂ reversed NSAID induced adenoma regression in *Apc^{Min}* mice.³¹ PGE₂ significantly enhanced colon carcinogen induced tumour incidence and multiplicity in rats.³² Furthermore, our group recently reported that PGE₂ accelerates intestinal adenoma growth in *Apc^{Min}* mice.³³

Although COX-2 selective inhibitors suppress PGE₂ production, the potential inhibition of endothelial cell derived COX-2 activity and subsequent PGI₂ production may promote platelet aggregation and lead to an increased risk of coronary thrombosis and stroke.³⁴ As PGI₂ antagonises thromboxane produced by platelets, inhibition of PGI₂ may shift the homeostatic balance towards more TxA₂ effects. In addition, PGI₂ appears to be important in protecting cardiomyocytes from oxidative stress.³⁵ Therefore, it will be important to develop chemopreventive agents that do not inhibit production of other prostanoids, such as the antithrombotic PGI₂. Given that only PGE₂ appears to be procarcinogenic, more selective pharmacological inhibition of PGE₂ production downstream of COX-2 may be superior and result in fewer side effects. To achieve this goal, researchers have been investigating precisely how PGE₂ promotes tumour growth and its signalling pathways.

COX independent targets

Another explanation for the antitumour effects of NSAIDs recently emerged, based on studies showing that high doses of NSAIDs inhibit tumour cell growth and induce apoptosis through COX independent mechanisms by regulating several different targets,³⁶ such as 15-LOX-1,³⁷ a proapoptotic gene Par-4,³⁸ antiapoptotic gene Bcl-X_L,³⁹ and nuclear factor κB (NFκB) signalling.^{40–41} In the above studies, higher NSAID concentrations may be impossible to achieve in vivo without significant toxicity. For example, the proapoptotic effects of NSAIDs seen at concentrations above 50 μM are most likely modulated through COX-2 independent pathways. However, the best characterised biochemical target of NSAIDs at therapeutic concentrations remains the COX enzymes. It is likely that many of the chemopreventive effects of NSAIDs are carried out via inhibition of COX-2.

PGE₂ AND CANCER

PGE₂ and its receptors play a predominant role in promoting cancer progression. The only other COX-2 derived prostaglandin implicated in oncogenesis is TxA₂, which was reported to promote angiogenesis.⁴² NSAIDs have been shown to inhibit PGE₂ mediated processes that play essential roles in tumour progression, such as tumour cell proliferation, invasion, angiogenesis, and immunosuppression. Therefore, we will focus on modulation of PGE₂ and its downstream targets that control these processes.

PGE₂ regulation and its receptors

Steady state cellular levels of PGE₂ depend on the relative rates of COX-2/PGE synthase dependent biosynthesis and 15-hydroxyprostaglandin dehydrogenase (15-PGDH) dependent degradation (fig 1). For example, cytosolic or microsomal PGE₂ synthases can convert PGH₂ to PGE₂. Two cytosolic PGE₂ synthases called cytosolic glutathione transferases (GSTM2-2 and GSTM3-3) catalyse the conversion of PGH₂ to PGE₂ in the human brain.⁴³ The two microsomal PGE₂ synthases characterised to date are mPGES1 and

Figure 1 Overview of prostaglandin (PG) synthesis and main functions. Arachidonic acid can be metabolised through three major pathways. In the cyclooxygenase (COX) pathway, each COX-2 derived prostaglandin (PGI_2 , PGE_2 , PGD_2 , $\text{PGF}_{2\alpha}$) or thromboxane A₂ (TxA_2) has its unique functions. NSAIDs, non-steroidal anti-inflammatory drugs; COXIBs, COX-2 selective inhibitors.

mPGES2. mPGES1 exhibits a higher catalytic activity than other PGES isomerases, indicating that it probably plays a key role in the synthesis of PGE_2 from PGH_2 .^{44–45}

15-PGDH, a prostaglandin degrading enzyme, catalyses oxidation of the 15(S)-hydroxyl group of PGE_2 to yield an inactive 15-keto PGE_2 .⁴⁶ Genetic deletion of *15-Pgdh* in mice leads to increased tissue levels of PGE_2 .⁴⁷ Although 15-PGDH may promote certain androgen sensitive prostate cancers,⁴⁸ we and others recently reported that loss of expression of 15-PGDH correlates with tumour formation, including colorectal cancer,^{49–50} lung,⁵¹ and transitional bladder cancer.⁵² Interestingly, NSAIDs have been shown to upregulate 15-PGDH expression in colorectal and medullary thyroid cancers.^{49–53} Taken together, these studies suggest that loss expression of 15-PGDH may contribute to tumour progression. The functional role of 15-PGDH in promoting tumour growth is currently under investigation in several laboratories.

PGE_2 exerts its cellular effects by binding to its cognate receptors (EP1–4) that belong to the family of seven transmembrane G protein coupled rhodopsin-type receptors. The central role of PGE_2 in tumorigenesis has been further confirmed through homozygous deletion of its receptors. Mice with homozygous deletions in EP1 and EP4 receptors, but not EP3, were partially resistant to colon carcinogen mediated induction of aberrant crypt foci.^{54–55} EP2 disruption decreases the number and size of intestinal polyps in $\text{APC}^{\Delta 716}$ knockout mice.⁵⁶ Moreover, in carcinogen treated wild-type mice, an EP1 receptor antagonist also decreased the incidence of aberrant crypt foci whereas Apc^{Min} mice treated with the same EP1 receptor antagonist and an EP4 receptor antagonist developed 57% and 69% fewer intestinal polyps, respectively,

than untreated mice.^{54–55} In addition to colorectal cancer, it has been shown that EP1, 2, and 4 receptors were elevated whereas EP3 receptor levels were decreased in mammary tumours in COX-2-MMTV mice.⁵⁷ Furthermore, an EP1 receptor antagonist was shown to reduce tumour burden in a carcinogen induced rat mammary model.⁵⁸ However, Amano and colleagues recently reported that EP3 receptor activation is required for lung tumour associated angiogenesis and tumour growth.⁵⁹ In the future, it will be important to carefully determine the EP receptor profile in human cancers and to examine whether NSAIDs can modulate PGE_2 receptor expression. Taken together, these findings may provide a rationale for the development of EP receptor antagonists which may offer an alternative to COX-2 selective inhibitors.

PGE_2 signalling pathways and its downstream targets

An increasingly large body of evidence indicates that PGE_2 promotes tumour growth by stimulating EP receptor signalling with subsequent enhancement of cellular proliferation, promotion of angiogenesis, inhibition of apoptosis, stimulation of invasion/motility, and suppression of immune responses. These findings prompted research to elucidate PGE_2 signalling pathways and identify PGE_2 downstream targets that are involved in promoting tumour growth (fig 2).

EGFR pathway

Both COX-2 and epidermal growth factor receptor (EGFR) pathways are activated in most human cancers.⁶⁰ The observation that forced expression of COX-2 in human colorectal cancer (CRC) cells stimulates cellular proliferation through induction of EGFR⁶¹ indicated the likelihood of

Figure 2 Prostaglandin (PG) E₂ in carcinogenesis. PGE₂ promotes tumour growth by stimulating EP receptor downstream signalling and subsequent enhancement of cellular proliferation, promotion of angiogenesis, inhibition of apoptosis, stimulation of invasion/motility, and suppression of immune responses. NSAIDs, non-steroidal anti-inflammatory drugs; COXIBs, COX-2 selective inhibitors; AA, arachidonic acid; EGFR, epidermal growth factor receptor; IL-10, interleukin 10; DAF, decay accelerating factor; VEGF, vascular endothelial growth factor; bFGF, basic fibroblast growth factor; PPARδ, peroxisome proliferator activated receptor δ.

crosstalk between these two pathways. We and others have demonstrated that PGE₂ can transactivate EGFR, which results in stimulation of cell migration through increased PI3K-Akt signalling in CRC cells.⁶²⁻⁶⁴ As expression of EGFR directly correlates with the ability of human CRC cells to metastasise to the liver,⁶⁵ it is possible that inhibiting both the EGFR tyrosine kinase and COX-2 at lower doses could yield additive effects, blocking the spread of metastatic disease. Moreover, preclinical studies support the notion that combined treatment of NSAIDs plus EGFR tyrosine kinase inhibitors is more effective than either single agent alone in several models. In colorectal carcinoma cells, blocking both COX-2 and the HER-2/neu pathways synergistically reduced tumour growth.⁶⁶ In soft agar and xenograft assays, the combination of a COX-2 selective inhibitor, an EGFR tyrosine kinase inhibitor, and a protein kinase A antisense construct markedly reduced proliferation and angiogenesis of human colon and breast cancer cells.⁶⁷ Similarly, combined treatment with inhibitors of both pathways significantly decreases polyp formation in *Apc*^{Min} mice,⁶⁸ as *Apc* deficiency is associated with increased EGFR activity in the intestinal enterocytes.⁶⁹ Hence we feel that it will be essential to examine the use of COX-2 selective inhibitors as potential agents in combination with EGFR tyrosine kinase inhibitors in clinical trials.

Nuclear receptor pathways

Peroxisome proliferator activated receptors δ (PPARδ, also referred to as PPARβ) is a ligand activated nuclear transcription factor that is a member of the nuclear hormone receptor superfamily. Published data indicate that the PPARδ/β is important for regulating fat metabolism,⁷⁰ inhibiting obesity induced by either genetic or high-fat-diet,⁷⁰ and decreasing weight gain and insulin resistance in mice fed high fat diets.⁷¹ However, PPARδ has been identified as a direct transcriptional target of the APC/β-catenin/Tcf pathway.⁷² Our recent findings show that PGE₂ transactivates PPARδ which in turn promotes tumour cell survival.³³ PGE₂ activates PPARδ via a PI3K-Akt pathway. Most importantly, we demonstrated that PGE₂ promotes intestinal epithelial cell survival and colorectal adenoma growth in *Apc*^{Min} mice, but not in *PPAR*^{δ-/-}/*Apc*^{Min} mice,³³ indicating that PPARδ is a critical downstream mediator in PGE₂ stimulated tumour growth. Consistent with this result, a selective PPARδ agonist also accelerates intestinal polyp growth in *Apc*^{Min} mice via inhibition of tumour cell apoptosis.⁷³ These results support the rationale for considering the development of PPARδ antagonists for use in cancer prevention and/or treatment and raise caution for developing PPARδ agonists for clinical use to treat dyslipidaemia, obesity, and insulin resistance.

Ras-MAPK pathway

Ras is an oncogene and its activation is found in a wide variety of human malignancies. Ras induces cell transformation, proliferation, and survival by triggering downstream signalling pathways such as the Raf/MEK/ERKs and PI3K/Akt pathways. The Ras-MAP kinase cascade is one of the major intracellular signalling pathways responsible for cell proliferation. Forced expression of constitutively active Ras (mutant Ras) or MEK upregulates COX-2 expression and enhances cell proliferation in a variety of cell culture models, respectively.^{74–77} Non-selective NSAIDs and COX-2 selective inhibitors inhibit cell proliferation and transformation by blocking the Ras-MAPK signalling pathway.^{78–80} Our group recently reported that PGE₂ activates a Ras-MAPK pathway which in turn upregulates COX-2 expression and stimulates colorectal cancer cell proliferation.⁸¹ This finding supports a novel mechanism by which COX-2 derived PGE₂ promotes human cancer cell growth by autoregulation of COX-2 expression, which depends primarily on PGE₂ induced activation of the Ras-MAPK pathway.

PGE₂ downstream targets: angiogenic factors

Angiogenic factors are important for the growth and survival of endothelial cells and to stimulate vascular endothelial cell migration and capillary formation.⁸² Overexpression of COX-2 in CRC cells induces the production of angiogenic factors such as vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF),⁸³ and NSAIDs block the production of these angiogenic factors leading to inhibition of proliferation, migration, and vascular tube formation.^{83–86} The observations that PGE₂ can reverse the antiangiogenic activity of NSAIDs⁸⁷ and homozygous deletion of EP2 completely abrogated induction of VEGF in APC^{Δ716} mouse polyps⁸⁶ support the concept that PGE₂ mediates the major role of COX-2 in inducing expression of proangiogenic factors. Several reports demonstrated that PGE₂ upregulates VEGF in cultured human fibroblasts⁸⁸ and increases VEGF and bFGF expression through stimulation of ERK2/JNK1 signalling pathways in endothelial cells.⁸⁹ EP2/EP4 mediate PGE₂ induction of VEGF in ovarian cancer cells⁹⁰ and human airway smooth muscle cells.⁹¹ Interestingly, VEGF and bFGF induce COX-2 and subsequent PG production in endothelial cells, suggesting that the effects of PGE₂ on regulation of VEGF and bFGF are likely amplified through a positive feedback loop.⁹² However, PGE₂ induction of VEGF may provide another explanation for the undesired side effect of COX-2 selective inhibitors on cardiovascular complications. VEGF is implicated in cardiovascular homeostasis. For example, treatment with Avastin (bevacizumab), a humanised anti-VEGF antibody, has only marginal improvement in survival for colorectal cancer patients and is also associated with an increased risk of hypertension. Therefore, it is important to identify new molecular targets that drive colorectal tumour associated angiogenesis. Preliminary data from our laboratory indicate that PGE₂ induces a proangiogenic chemokine expression in CRC cells (unpublished data).

PGE₂ downstream targets: antiapoptotic factors

Overexpression of COX-2 in rat intestinal epithelial cells increased their resistance to undergo apoptosis and resulted in increased Bcl-2 protein expression.⁹³ The role of COX-2 in preventing apoptosis is likely mediated by COX-2 derived PGE₂, which attenuates cell death induced by the COX-2 selective inhibitor SC-58125.⁹⁴ These findings have stimulated

great interest in identifying PGE₂ downstream targets responsible for modulating apoptosis. PGE₂ induces anti-apoptotic protein expression such as Bcl-2⁹⁴ and increases NFκB transcriptional activity,⁹⁵ which is a key antiapoptotic mediator. As chemotherapeutic agents and radiation therapy enhance COX-2 protein expression as well as PGE₂ synthesis in human cancer cells, elevated PGE₂ production may increase resistance to therapy by giving cells a survival advantage. It will be important to determine whether patients treated with combinations of chemotherapy or/and radiation therapy with NSAIDs respond better than those not treated with NSAIDs. Preliminary study suggests that COX-2 selective inhibitors may increase the beneficial effects of radiotherapy.⁹⁶

PGE₂ downstream targets: chemokines and their receptors

Although chemokines play a crucial role in immune and inflammatory reactions, recent studies indicate that they have an equally important role in the cancer.⁹⁷ PGE₂ has been shown to inhibit production of CC chemokines, macrophage inflammatory protein (MIP)-1α and MIP-1β, in dendritic cells via binding to the EP2 receptor,^{98,99} and suppress CC chemokine RANTES (regulated upon activation normal T cell expressed and secreted) production in human macrophages through the EP4 receptor.¹⁰⁰ These CC chemokines are crucial for macrophage and lymphocyte infiltration in human breast, cervix, pancreas, and gliomas cancers.^{101–102} Moreover, a recent study showed that PGE₂ mediates VEGF and bFGF induced CXCR4 dependent neovessel assembly *in vivo*.¹⁰³ The important role of CXCR4 for cancer pharmacology is based on findings that activation of CXCR4 is involved in stimulating cancer cell migration and increasing invasion in breast, prostate, bladder, and pancreatic cancers.^{104–107} These pre-clinical data indicate that chemokines and their receptors are potential drug targets of PGE₂ downstream signalling for cancer prevention and treatment.

PGE₂ downstream targets: immunosuppressive mediators

The tumour microenvironment is predominantly shifted from a Th1 to a Th2 dominant response (immunosuppressive immune responses). COX-2 selective inhibitors restore the tumour induced imbalance between Th1 and Th2 and promote antineoplastic responses in lung cancer¹⁰⁸ and metastatic spread of colorectal cancer.¹⁰⁹ These findings led to extensive efforts to understand how PGE₂ can regulate immunosuppression. PGE₂ has been shown to downregulate Th1 cytokines (tumour necrosis factor α, interferon γ, and interleukin (IL)-2)¹¹⁰ and upregulate Th2 cytokines such as IL-4, IL-10, and IL-6.^{111–113} IL-10 is an immunosuppressive cytokine. Moreover, PGE₂ can modulate immune function through inhibiting dendritic cell differentiation and T cell proliferation and suppressing the antitumour activity of natural killer cells and macrophages.^{114–115} In addition, our group showed that PGE₂ upregulates the complement regulatory protein decay accelerating factor which results in blocking the complement C3 into two active compounds, C3a and C3b in CRC cells.¹¹⁶ Thus the effects of PGE₂ on the immune system may allow neoplastic cells to evade attack.

CONCLUSIONS

Chemoprevention is being carefully evaluated on several fronts as an effective measure to insure cancer control.

NSAIDs and COX-2 selective inhibitors have been touted for their possible use as chemopreventive agents. However, concerns over the safety of COX-2 selective inhibitors have prompted researchers to develop more effective chemopreventive agents with minimal toxicity. Understanding the molecular mechanisms of COX-2 and its downstream targets will help to identify specific molecular targets for developing more safe agents which target this pathway. Preclinical studies provide evidence to demonstrate that non-selective NSAIDs and COX-2 selective inhibitors decrease the risk of colorectal cancer as well as other cancers through reduction of COX-2 derived PGE₂ synthesis. Significant progress has been made in the elucidation of PGE₂ downstream signalling pathways which mediate the chemopreventive effect of NSAIDs. These cumulative data indicate that the development of agents that lower cellular levels of PGE₂ or that specifically inhibit the PGE₂ downstream signalling pathway might be useful for cancer prevention. PGE synthases, 15-PDGH, and/or PGE₂ receptors may also serve as rational targets for lowering cellular levels of PGE₂ and EGFR, MAPK, and chemokines, and chemokine receptors are molecular targets for specific inhibition of PGE₂ downstream signalling pathways. Moreover, another approach for decreasing the undesired side effects of COXIBs may be to lower the drug dose used. The combined use of multiple agents may allow for a lower dose of drug to be used. Taken together, efforts to develop novel chemopreventive agents with minimal toxicity and to design strategies for combinations of different agents targeting multiple pathways may yield significant benefits for cancer patients.

ACKNOWLEDGEMENTS

We acknowledge support from the United States Public Health Services Grants RO1-DK-62112 and PO1-CA77839. RND is the Hortense B Ingram Professor of Molecular Oncology and the recipient of an NIH MERIT award (R37-DK47297). We also thank the TJ Martell Foundation and the National Colorectal Cancer Research Alliance (NCCRA) for generous support.

Authors' affiliations

D Wang, Department of Medicine, Vanderbilt University Medical Center, Nashville, TN, USA

R N DuBois, Departments of Medicine, Cancer Biology, and Cell and Developmental Biology, Vanderbilt University Medical Center, Nashville, TN, USA, and The Vanderbilt-Ingram Cancer Center, Nashville, TN, USA

Conflict of interest: None declared.

REFERENCES

- Smalley W, DuBois RN. Colorectal cancer and non steroidal anti-inflammatory drugs. *Adv Pharmacol* 1997;**39**:1–20.
- Gupta RA, Dubois RN. Colorectal cancer prevention and treatment by inhibition of cyclooxygenase-2. *Nat Rev Cancer* 2001;**1**:11–21.
- Wolfe MM, Lichtenstein DR, Singh G. Gastrointestinal toxicity of nonsteroidal antiinflammatory drugs. *N Engl J Med* 1999;**340**:1888–99.
- Vane JR, Bakhle YS, Botting RM. Cyclooxygenases 1 and 2. *Annu Rev Pharmacol Toxicol* 1998;**38**:97–120.
- Grover JK, Yadav S, Vats V, et al. Cyclo-oxygenase 2 inhibitors: emerging roles in the gut. *Int J Colorectal Dis* 2003;**18**:279–91.
- Vane JR. Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nature* 1971;**231**:232–5.
- Laine L, Harper S, Simon T, et al. A randomized trial comparing the effect of rofecoxib, a cyclooxygenase 2-specific inhibitor, with that of ibuprofen on the gastroduodenal mucosa of patients with osteoarthritis. *Gastroenterology* 1999;**117**:776–83.
- Bombardier C, Laine L, Reicin A, et al. Comparison of upper gastrointestinal toxicity of rofecoxib and naproxen in patients with rheumatoid arthritis. VIGOR Study Group. *N Engl J Med* 2000;**343**:1520–8.
- Silverman DG, Halaszynski T, Sinatra R, et al. Rofecoxib does not compromise platelet aggregation during anesthesia and surgery. *Can J Anaesth* 2003;**50**:1004–8.
- Anderson WF, Umar A, Viner JL, et al. The role of cyclooxygenase inhibitors in cancer prevention. *Curr Pharm Des* 2002;**8**:1035–62.
- Thun MJ, Henley SJ, Patrono C. Nonsteroidal anti-inflammatory drugs as anticancer agents: mechanistic, pharmacologic, and clinical issues. *J Natl Cancer Inst* 2002;**94**:252–66.
- Steinbach G, Lynch PM, Phillips RK, et al. The effect of celecoxib, a cyclooxygenase-2 inhibitor, in familial adenomatous polyposis. *N Engl J Med* 2000;**342**:1946–52.
- Solomon SD, McMurray JJ, Pfeffer MA, et al. Cardiovascular risk associated with celecoxib in a clinical trial for colorectal adenoma prevention. *N Engl J Med* 2005;**352**:1071–80.
- Bresalier RS, Sandler RS, Quan H, et al. Cardiovascular events associated with rofecoxib in a colorectal adenoma chemoprevention trial. *N Engl J Med* 2005;**352**:1092–102.
- Nussmeier NA, Whelton AA, Brown MT, et al. Complications of the COX-2 inhibitors parecoxib and valdecoxib after cardiac surgery. *N Engl J Med* 2005;**352**:1081–91.
- Kuper H, Adami HO, Trichopoulos D. Infections as a major preventable cause of human cancer. *J Intern Med* 2000;**248**:171–83.
- Wang D, Mann JR, DuBois RN. The role of prostaglandins and other eicosanoids in the gastrointestinal tract. *Gastroenterology* 2005;**128**:1445–61.
- McAdam BF, Catella-Lawson F, Mardini IA, et al. Systemic biosynthesis of prostacyclin by cyclooxygenase (COX)-2: the human pharmacology of a selective inhibitor of COX-2. *Proc Natl Acad Sci U S A* 1999;**96**:272–7.
- Grosser T, Yusuf S, Cheskis E, et al. Developmental expression of functional cyclooxygenases in zebrafish. *Proc Natl Acad Sci U S A* 2002;**99**:8418–23.
- Chai YI, Kim SH, Solnica-Krezel L, et al. Cyclooxygenase-1 signaling is required for vascular tube formation during development. *Dev Biol* 2005;**282**:274–83.
- DuBois RN, Abramson SB, Crofford L, et al. Cyclooxygenase in biology and disease. *FASEB J* 1998;**12**:1063–73.
- Rigas B, Goldman IS, Levine L. Altered eicosanoid levels in human colon cancer. *J Lab Clin Med* 1993;**122**:518–23.
- Uefuji K, Ichikura T, Mochizuki H. Cyclooxygenase-2 expression is related to prostaglandin biosynthesis and angiogenesis in human gastric cancer. *Clin Cancer Res* 2000;**6**:135–8.
- Rolland PH, Martin PM, Jacquemier J, et al. Prostaglandin in human breast cancer: Evidence suggesting that an elevated prostaglandin production is a marker of high metastatic potential for neoplastic cells. *J Natl Cancer Inst* 1980;**64**:1061–70.
- Needleman P, Isakson PC. The discovery and function of COX-2. *J Rheumatol* 1997;**24**(suppl 49):6–8.
- Portanova JP, Zhang Y, Anderson GD, et al. Selective neutralization of prostaglandin E2 blocks inflammation, hyperalgesia, and interleukin 6 production in vivo. *J Exp Med* 1996;**184**:883–91.
- Anderson GD, Hauser SD, McGarity KL, et al. Selective inhibition of cyclooxygenase (COX)-2 reverses inflammation and expression of COX-2 and interleukin 6 in rat adjuvant arthritis. *J Clin Invest* 1996;**97**:2672–9.
- Amin AR, Attur M, Abramson SB. Nitric oxide synthase and cyclooxygenases: distribution, regulation, and intervention in arthritis. *Curr Opin Rheumatol* 1999;**11**:202–9.
- MacDermott RP. Alterations in the mucosal immune system in ulcerative colitis and Crohn's disease. *Med Clin North Am* 1994;**78**:1207–31.
- Gould SR, Brash AR, Conolly ME, et al. Studies of prostaglandins and sulphasalazine in ulcerative colitis. *Prostaglandins Med* 1981;**6**:165–82.
- Hansen-Petrik MB, McEntee MF, Jull B, et al. Prostaglandin E(2) protects intestinal tumors from nonsteroidal anti-inflammatory drug-induced regression in Apc(Min/+) mice. *Cancer Res* 2002;**62**:403–8.
- Kawamori T, Uchiya N, Sugimura T, et al. Enhancement of colon carcinogenesis by prostaglandin E2 administration. *Carcinogenesis* 2003;**24**:985–90.
- Wang D, Wang H, Shi Q, et al. Prostaglandin E(2) promotes colorectal adenoma growth via transactivation of the nuclear peroxisome proliferator-activated receptor delta. *Cancer Cell* 2004;**6**:285–95.
- Mukherjee D, Nissen SE, Topol EJ. Risk of cardiovascular events associated with selective COX-2 inhibitors. *JAMA* 2001;**286**:954–9.
- Adderley SR, Fitzgerald DJ. Oxidative damage of cardiomyocytes is limited by extracellular regulated kinases 1/2-mediated induction of cyclooxygenase-2. *J Biol Chem* 1999;**274**:5038–46.
- Kashfi K, Rigas B. Non-COX-2 targets and cancer: Expanding the molecular target repertoire of chemoprevention. *Biochem Pharmacol* 2005;**70**:969–86.
- Shureiqi I, Chen D, Lee JJ, et al. 15-LOX-1: a novel molecular target of nonsteroidal anti-inflammatory drug-induced apoptosis in colorectal cancer cells. *J Natl Cancer Inst* 2000;**92**:1136–42.
- Zhang Z, DuBois RN. Par-4, a proapoptotic gene, is regulated by NSAIDs in human colon carcinoma cells. *Gastroenterology* 2000;**118**:1012–17.
- Zhang L, Yu J, Park BH, et al. Role of BAX in the apoptotic response to anticancer agents. *Science* 2000;**290**:989–92.
- Yamamoto Y, Yin MJ, Lin KM, et al. Sulindac inhibits activation of the NF-kappaB pathway. *J Biol Chem* 1999;**274**:27307–14.
- Grilli M, Pizzi M, Memo M, et al. Neuroprotection by aspirin and sodium salicylate through blockade of NF-kappaB activation. *Science* 1996;**274**:1383–5.
- Pradono P, Tazawa R, Maemondo M, et al. Gene transfer of thromboxane A(2) synthase and prostaglandin I(2) synthase antithetically altered tumor angiogenesis and tumor growth. *Cancer Res* 2002;**62**:63–6.
- Beuckmann CT, Fujimori K, Urade Y, et al. Identification of mu-class glutathione transferases M2-2 and M3-3 as cytosolic prostaglandin E synthases in the human brain. *Neurochem Res* 2000;**25**:733–8.

- 44 Jakobsson PJ, Thoren S, Morgenstern R, et al. Identification of human prostaglandin E synthase: a microsomal, glutathione-dependent, inducible enzyme, constituting a potential novel drug target. *Proc Natl Acad Sci U S A* 1999;22:7220-5.
- 45 Lazarus M, Munday CJ, Eguchi N, et al. Immunohistochemical localization of microsomal PGE synthase-1 and cyclooxygenases in male mouse reproductive organs. *Endocrinology* 2002;143:2410-19.
- 46 Tai HH, Ensor CM, Tong M, et al. Prostaglandin catabolizing enzymes. *Prostaglandins Other Lipid Mediat* 2002;68-69:483-93.
- 47 Coggins KG, Latour A, Nguyen MS, et al. Metabolism of PGE2 by prostaglandin dehydrogenase is essential for remodeling the ductus arteriosus. *Nat Med* 2002;8:91-2.
- 48 Tong M, Tai HH. Synergistic induction of the nicotinamide adenine dinucleotide-linked 15-hydroxyprostaglandin dehydrogenase by an androgen and interleukin-6 or forskolin in human prostate cancer cells. *Endocrinology* 2004;145:2141-7.
- 49 Backlund MG, Mann JR, Holla VR, et al. 15-Hydroxyprostaglandin dehydrogenase is down-regulated in colorectal cancer. *J Biol Chem* 2005;280:3217-23.
- 50 Yan M, Rerko RM, Platzer P, et al. 15-Hydroxyprostaglandin dehydrogenase, a COX-2 oncogene antagonist, is a TGF-beta-induced suppressor of human gastrointestinal cancers. *Proc Natl Acad Sci U S A* 2004;101:17468-73.
- 51 Ding Y, Tong M, Liu S, et al. NAD+-linked 15-hydroxyprostaglandin dehydrogenase (15-PGDH) behaves as a tumor suppressor in lung cancer. *Carcinogenesis* 2005;26:65-72.
- 52 Gee JR, Montoya RG, Khaled HM, et al. Cytokeratin 20, AN43, PGDH, and COX-2 expression in transitional and squamous cell carcinoma of the bladder. *Urol Oncol* 2003;21:266-70.
- 53 Quidville V, Segond N, Pidoux E, et al. Tumor growth inhibition by indomethacin in a mouse model of human medullary thyroid cancer: implication of cyclooxygenases and 15-hydroxyprostaglandin dehydrogenase. *Endocrinology* 2004;145:2561-71.
- 54 Watanabe K, Kawamori T, Nakatsugi S, et al. Role of the prostaglandin E receptor subtype EP1 in colon carcinogenesis. *Cancer Res* 1999;59:5093-6.
- 55 Mutoh M, Watanabe K, Kitamura T, et al. Involvement of prostaglandin E receptor subtype EP4 in colon carcinogenesis. *Cancer Res* 2002;62:28-32.
- 56 Sonoshita M, Takaku K, Sasaki N, et al. Acceleration of intestinal polyposis through prostaglandin receptor EP2 in ApcDelta716 knockout mice. *Nat Med* 2001;7:1048-51.
- 57 Chang SH, Liu CH, Conway R, et al. Role of prostaglandin E2-dependent angiogenic switch in cyclooxygenase 2-induced breast cancer progression. *Proc Natl Acad Sci U S A* 2004;101:591-6.
- 58 Kawamori T, Uchiya N, Nakatsugi S, et al. Chemopreventive effects of ONO-8711, a selective prostaglandin E receptor EP1 antagonist, on breast cancer development. *Carcinogenesis* 2001;22:2001-4.
- 59 Amano H, Hayashi I, Endo H, et al. Host prostaglandin E(2)-EP3 signaling regulates tumor-associated angiogenesis and tumor growth. *J Exp Med* 2003;197:221-32.
- 60 Kellogg F, Fay J, Steele V, et al. Epidermal growth factor receptor tyrosine kinase inhibitors as potential cancer chemopreventives. *Cancer Epidemiol Biomarkers Prev* 1996;5:657-66.
- 61 Yoshimoto T, Takahashi Y, Kinoshita T, et al. Growth stimulation and epidermal growth factor receptor induction in cyclooxygenase-overexpressing human colon carcinoma cells. *Adv Exp Med Biol* 2002;507:403-7.
- 62 Sheng H, Shao J, Washington MK, et al. Prostaglandin E2 increases growth and motility of colorectal carcinoma cells. *J Biol Chem* 2001;276:18075-81.
- 63 Buchanan FG, Wang D, Bargiacchi F, et al. Prostaglandin E2 regulates cell migration via the intracellular activation of the epidermal growth factor receptor. *J Biol Chem* 2003;278:35451-7.
- 64 Pai R, Soreghan B, Szabo IL, et al. Prostaglandin E2 transactivates EGF receptor: A novel mechanism for promoting colon cancer growth and gastrointestinal hypertrophy. *Nat Med* 2002;8:289-93.
- 65 Radinsky R. Modulation of tumor cell gene expression and phenotype by the organ-specific metastatic environment. *Cancer Metastasis Rev* 1995;14:323-38.
- 66 Mann M, Sheng H, Shao J, et al. Targeting cyclooxygenase 2 and her-2/neu pathways inhibits colorectal carcinoma growth. *Gastroenterology* 2001;120:1713-19.
- 67 Tortora G, Caputo R, Damiano V, et al. Combination of a selective cyclooxygenase-2 inhibitor with epidermal growth factor receptor tyrosine kinase inhibitor ZD1839 and protein kinase A antisense causes cooperative antitumor and antiangiogenic effect. *Clin Cancer Res* 2003;9:1566-72.
- 68 Torrance CJ, Jackson PE, Montgomery E, et al. Combinatorial chemoprevention of intestinal neoplasia. *Nat Med* 2000;6:1024-8.
- 69 Moran AE, Hunt DH, Javid SH, et al. Apc deficiency is associated with increased Egr activity in the intestinal enterocytes and adenomas of C57BL/6J-Min/+ mice. *J Biol Chem* 2004;279:43261-72.
- 70 Wang YX, Lee CH, Tiep S, et al. Peroxisome-proliferator-activated receptor delta activates fat metabolism to prevent obesity. *Cell* 2003;113:159-70.
- 71 Tanaka T, Yamamoto J, Iwasaki S, et al. Activation of peroxisome proliferator-activated receptor delta induces fatty acid beta-oxidation in skeletal muscle and attenuates metabolic syndrome. *Proc Natl Acad Sci U S A* 2003;100:15924-9.
- 72 He TC, Chan TA, Vogelstein B, et al. PPARdelta is an APC-regulated target of nonsteroidal anti-inflammatory drugs. *Cell* 1999;99:335-45.
- 73 Gupta RA, Wang D, Katkuri S, et al. Activation of nuclear hormone receptor peroxisome proliferator-activated receptor-delta accelerates intestinal adenoma growth. *Nat Med* 2004;10:245-7.
- 74 Sheng GG, Shao J, Sheng H, et al. A selective cyclooxygenase 2 inhibitor suppresses the growth of H-ras-transformed rat intestinal epithelial cells. *Gastroenterology* 1997;113:1883-91.
- 75 Sheng H, Williams CS, Shao J, et al. Induction of cyclooxygenase-2 by activated Ha-ras oncogene in Rat-1 fibroblasts and the role of mitogen-activated protein kinase pathway. *J Biol Chem* 1998;273:22120-7.
- 76 Subbaramaiah K, Telang N, Ramonetti JT, et al. Transcription of cyclooxygenase-2 is enhanced in transformed mammary epithelial cells. *Cancer Res* 1996;56:4424-9.
- 77 Heasley LE, Thaler S, Nicks M, et al. Induction of cytosolic phospholipase A2 by oncogenic Ras in human non-small cell lung cancer. *J Biol Chem* 1997;272:14501-4.
- 78 Wong BC, Jiang XH, Lin MC, et al. Cyclooxygenase-2 inhibitor (SC-236) suppresses activator protein-1 through c-Jun NH2-terminal kinase. *Gastroenterology* 2004;126:136-47.
- 79 Husain SS, Szabo IL, Pai R, et al. MAPK (ERK2) kinase—a key target for NSAIDs-induced inhibition of gastric cancer cell proliferation and growth. *Life Sci* 2001;69:3045-54.
- 80 Gala M, Sun R, Yang VW. Inhibition of cell transformation by sulindac sulfide is confined to specific oncogenic pathways. *Cancer Lett* 2002;175:89-94.
- 81 Wang D, Buchanan FG, Wang H, et al. Prostaglandin E2 enhances intestinal adenoma growth via activation of the Ras-mitogen-activated protein kinase cascade. *Cancer Res* 2005;65:1822-9.
- 82 Iniguez MA, Rodriguez A, Volpert OV, et al. Cyclooxygenase-2: a therapeutic target in angiogenesis. *Trends Mol Med* 2003;9:73-8.
- 83 Tsujii M, Kawano S, Tsuji S, et al. Cyclooxygenase regulates angiogenesis induced by colon cancer cells. *Cell* 1998;93:705-16.
- 84 Masferrer JL, Leahy KM, Koki AT, et al. Antiangiogenic and antitumor activities of cyclooxygenase-2 inhibitors. *Cancer Res* 2000;60:1306-11.
- 85 Skopinska-Rozewska E, Piazza GA, Sommer E, et al. Inhibition of angiogenesis by sulindac and its sulfone metabolite (FGN-1): a potential mechanism for their antineoplastic properties. *Int J Tissue React* 1998;20:85-9.
- 86 Sawaoka H, Tsuji S, Tsujii M, et al. Cyclooxygenase inhibitors suppress angiogenesis and reduce tumor growth in vivo. *Lab Invest* 1999;79:1469-77.
- 87 Hernandez GL, Volpert OV, Iniguez MA, et al. Selective inhibition of vascular endothelial growth factor-mediated angiogenesis by cyclosporin A: roles of the nuclear factor of activated T cells and cyclooxygenase 2. *J Exp Med* 2001;193:607-20.
- 88 Trompezinski S, Pernet I, Schmitt D, et al. UV radiation and prostaglandin E2 up-regulate vascular endothelial growth factor (VEGF) in cultured human fibroblasts. *Inflamm Res* 2001;50:422-7.
- 89 Pai R, Szabo IL, Soreghan BA, et al. PGE2 stimulates VEGF expression in endothelial cells via ERK2/JNK1 signaling pathways. *Biochem Biophys Res Commun* 2001;286:923-8.
- 90 Spinella F, Rosano L, Di Castro V, et al. Endothelin-1-induced prostaglandin E2-EP2, EP4 signaling regulates vascular endothelial growth factor production and ovarian carcinoma cell invasion. *J Biol Chem* 2004;279:46700-5.
- 91 Bradbury DA, Clarke DL, Seedhouse C, et al. Vascular endothelial growth factor induction by prostaglandin E2 in human airway smooth muscle cells is mediated by E prostanoic acid EP2/EP4 receptors and SP-1 transcription factor binding sites. *J Biol Chem* 2005;280:29993-30000.
- 92 Kage K, Fujita N, Oh-hara T, et al. Basic fibroblast growth factor induces cyclooxygenase-2 expression in endothelial cells derived from bone. *Biochem Biophys Res Commun* 1999;254:259-63.
- 93 Tsujii M, DuBois RN. Alterations in cellular adhesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase-2. *Cell* 1995;83:493-501.
- 94 Sheng H, Shao J, Morrow J, et al. Modulation of apoptosis by prostaglandin treatment in human colon cancer cells. *Cancer Res* 1998;58:362-6.
- 95 Poligone B, Baldwin AS. Positive and negative regulation of NF-kappaB by COX-2: roles of different prostaglandins. *J Biol Chem* 2001;276:38658-64.
- 96 Davis TW, O'Neal JM, Pagel MD, et al. Synergy between celecoxib and radiotherapy results from inhibition of cyclooxygenase-2-derived prostaglandin E2, a survival factor for tumor and associated vasculature. *Cancer Res* 2004;64:279-85.
- 97 Balkwill F. Cancer and the chemokine network. *Nat Rev Cancer* 2004;4:540-50.
- 98 Jing H, Vassiliou E, Ganea D. Prostaglandin E2 inhibits production of the inflammatory chemokines CCL3 and CCL4 in dendritic cells. *J Leukoc Biol* 2003;74:868-79.
- 99 Jing H, Yen JH, Ganea D. A novel signaling pathway mediates the inhibition of CCL3/4 expression by PGE2. *J Biol Chem* 2004;279:55176-86.
- 100 Takayama K, Garcia-Cardena G, Sukhova GK, et al. Prostaglandin E2 suppresses chemokine production in human macrophages through the EP4 receptor. *J Biol Chem* 2002;277:44147-54.
- 101 Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet* 2001;357:539-45.
- 102 Bottazzi B, Polentarutti N, Acero R, et al. Regulation of the macrophage content of neoplasms by chemoattractants. *Science* 1983;220:210-2.
- 103 Salcedo R, Zhang X, Young HA, et al. Angiogenic effects of prostaglandin E2 are mediated by up-regulation of CXCR4 on human microvascular endothelial cells. *Blood* 2003;102:1966-77.
- 104 Liang Z, Yoon Y, Votaw J, et al. Silencing of CXCR4 blocks breast cancer metastasis. *Cancer Res* 2005;65:967-71.
- 105 Hart CA, Brown M, Bagley S, et al. Invasive characteristics of human prostatic epithelial cells: understanding the metastatic process. *Br J Cancer* 2005;92:503-12.

- 106 Eisenhardt A, Frey U, Tack M, *et al.* Expression analysis and potential functional role of the CXCR4 chemokine receptor in bladder cancer. *Eur Urol* 2005;**47**:111–17.
- 107 Marchesi F, Monti P, Leone BE, *et al.* Increased survival, proliferation, and migration in metastatic human pancreatic tumor cells expressing functional CXCR4. *Cancer Res* 2004;**64**:8420–7.
- 108 Stolina M, Sharma S, Lin Y, *et al.* Specific inhibition of cyclooxygenase 2 restores antitumor reactivity by altering the balance of IL-10 and IL-12 synthesis. *J Immunol* 2000;**164**:361–70.
- 109 Yao M, Kargman S, Lam EC, *et al.* Inhibition of cyclooxygenase-2 by rofecoxib attenuates the growth and metastatic potential of colorectal carcinoma in mice. *Cancer Res* 2003;**63**:586–92.
- 110 Harris SG, Padilla J, Koumas L, *et al.* Prostaglandins as modulators of immunity. *Trends Immunol* 2002;**23**:144–50.
- 111 Shreedhar V, Giese T, Sung VW, *et al.* A cytokine cascade including prostaglandin E2, IL-4, and IL-10 is responsible for UV-induced systemic immune suppression. *J Immunol* 1998;**160**:3783–9.
- 112 Huang M, Stolina M, Sharma S, *et al.* Non-small cell lung cancer cyclooxygenase-2-dependent regulation of cytokine balance in lymphocytes and macrophages: up-regulation of interleukin 10 and down-regulation of interleukin 12 production. *Cancer Res* 1998;**58**:1208–16.
- 113 Della Bella S, Molteni M, Compasso S, *et al.* Differential effects of cyclo-oxygenase pathway metabolites on cytokine production by T lymphocytes. *Prostaglandins Leukot Essent Fatty Acids* 1997;**56**:177–84.
- 114 Yang L, Yamagata N, Yadav R, *et al.* Cancer-associated immunodeficiency and dendritic cell abnormalities mediated by the prostaglandin EP2 receptor. *J Clin Invest* 2003;**111**:727–35.
- 115 Goodwin JS, Ceuppens J. Regulation of the immune response by prostaglandins. *J Clin Immunol* 1983;**3**:295–315.
- 116 Holla VR, Wang D, Brown JR, *et al.* Prostaglandin E2 regulates the complement inhibitor CD55/decay-accelerating factor in colorectal cancer. *J Biol Chem* 2005;**280**:476–83.

EDITOR'S QUIZ: GI SNAPSHOT

Hypervascular pancreatic mass

Robin Spiller, *Editor*

Clinical presentation

A 58 year old man presented with mild cholestasis, with serum alkaline phosphatase 155 U/l (normal 30–120 U/l), serum gamma glutamyl transferase 75 U/l (normal 12–55 U/l), and serum bilirubin 1.45 mg/dl (normal 0.2–1.3 mg/dl). During the previous three months he had lost 2–3 kg in weight but his general state was good with a body mass index of 26 kg/m².

The patient underwent appendicectomy in his childhood and cholecystectomy for gall bladder stone disease 12 years previously. Eight years before being referred to our institution, he had been nephrectomised for renal cell cancer of the right kidney. The tumour measured 5 cm and microscopically did not infiltrate either the renal capsule or blood vessels (stage T1b, N0, M0). No recurrence was detected during regular follow ups. Five years later in the course of bacterial pneumonia the patient developed acute renal failure which required single haemodialysis. At presentation the patient was not anaemic and his serum creatinine level was 1.36 mg/dl (normal 0.7–1.2 mg/dl).

Ultrasound examination revealed a hypoechogenic mass of the pancreatic head, measuring 4 cm in diameter. On biphasic computed tomography (CT) the pancreatic mass was clearly hypervascular in the arterial phase (fig 1) but in the venous phase its contrast enhancement returned to normal. Radiographically, no other masses were detected, and the peripancreatic lymph nodes were not enlarged. The duodenal wall and peripancreatic tissue were not infiltrated by the tumour. The left kidney was intact on both CT and intravenous urography. Serum concentrations of cancer carcinoembryonic antigen, insulin, serotonin, and gastrin were within the normal range. Fine needle biopsy from the tumour was diagnostically not contributory, showing only desmoplastic fibres and normal glandular cells.

Question

What is the presurgical diagnosis and what is the prognosis in this case?

See page 141 for answer

This case is submitted by:

Figure 1 At computed tomography, the pancreatic mass showed increased contrast enhancement in the arterial phase.

M Hartleb

A Nowak

M Błaszczyńska

Department of Gastroenterology, Silesian Medical School, Katowice, Poland

P Lampe

Department of Digestive Tract Surgery, Silesian Medical School, Katowice, Poland

M Kajor

Department of Pathomorphology, Silesian Medical School, Katowice, Poland

Correspondence to: Dr M Hartleb, Klinika Gastroenterologii, CSK, ul Medyków 14, 40-752 Katowice, Poland; lakmus@poczta.wprosl.pl

doi: 10.1136/gut.2005.064410