Maximal acid response of Pavlov pouches to food and histamine

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SUMMARY Secretion of gastric acid in response to histamine, insulin, and a meal of liver, with and without 5% bone dust, was studied in three dogs with Pavlov pouches and gastric fistulas. The ratios of insulin-induced to histamine-induced secretion were the same in pouch and main stomach indicating that their innervation was equivalent. Meals of a liver and bone dust mixture in amounts of 11 to 44 g/kg resulted in Pavlov pouch acid outputs that were 34 to 56% higher than the maximal response to histamine (p < 0.05 to < 0.001). Liver without bone dust also produced higher acid outputs than histamine. These results indicate that combinations of endogenously generated stimuli can produce a potentiated gastric response which exceeds that produced by histamine alone.

In dogs with vagally denervated pouches histamine and food in maximal doses produced about the same rate of acid secretion (Thomas and Forrest, 1967). Indirect studies in man using changes in blood base gave similar results although the author recognized that his technique might underestimate food-induced gastric secretion (Rune, 1966). The present investigation compares the gastric acid response to food and histamine in dogs with innervated (Pavlov) pouches.

Methods

Three dogs weighing 12.7 to 16.2 kg were prepared with a Gregory cannula (Gregory, 1958) in a Pavlov pouch (Emás, Swan, and Jacobson, 1967) and a Thomas cannula (Thomas, 1941) in the main stomach. Tests were started two weeks after surgery and were done at intervals of at least 48 hours.

After a fast of 18 hours or more, 15-min collections of gastric juice were obtained from the gastric fistula and Pavlov pouch. In feeding experiments the gastric fistula was closed throughout. Volumes were measured and 0.2 ml samples were titrated to pH 7 on an automatic titrator.1

An intravenous infusion of 154 mM NaCl was delivered at 60 ml per hour by a peristaltic pump2 throughout each study. Histamine was added to the NaCl infusion and insulin was rapidly injected through a three-way stopcock in the tubing of the intravenous infusion.

Four basal 15-min collections were obtained and the gastric stimulant (food, histamine, or insulin) were given. A further 16 15-min collec-

1Radiometer, Copenhagen, Denmark.
Fig. 1  Mean 15 min-Pavlov pouch acid outputs in response to histamine infusion. Each point is the mean of two observations on each of three dogs. The vertical lines are the standard errors of the mean.

Fig. 2  Mean peak 15-min Pavlov pouch acid outputs in response to histamine, liver-bone dust mixture, and liver alone. In histamine and liver-bone dust studies each point represents two observations on each of three dogs. In experiments using liver without bone dust each point is the mean of one observation on each of three dogs.

Fig. 3  Comparison in each dog of mean peak 15-min acid output from Pavlov pouch and gastric fistula in response to insulin (black) 1-0 units/kg and to histamine (shaded).

Fig. 4  Mean peak 15-min insulin-stimulated acid output from Pavlov pouch and gastric fistula expressed as a percentage of histamine response (derived from data of Figure 3).
Histamine produced a sustained acid output from the Pavlov pouch which was approximately equal for the three doses used (Fig. 1). There was no significant difference between the three doses in the peak 15-min acid output (Fig. 2). The responses of the gastric fistula and the Pavlov pouch to histamine and insulin are shown in Figure 3. To compare the response to insulin of the gastric fistula and Pavlov pouch each is expressed as a percentage of its histamine response (and presumably as a percentage of its parietal cell mass) so that absolute differences in the secretory rate of gastric fistula and Pavlov pouch are eliminated (Fig. 4). Insulin-stimulated acid output was 48.2 ± 3.2% of the histamine response from the gastric fistula and 49.7 ± 11.5% of the histamine response from the Pavlov pouch. This suggests that the pouch and main stomach do not differ significantly in the degree of vagal innervation.

The three highest doses of liver-bone dust mixture produced peak acid outputs that were 33.7 to 56.2% greater than the peak histamine response (Fig. 2, p < 0.05 to < 0.001). The acid output induced by smaller meals was less sustained than with larger meals (Fig. 5). Liver without bone dust produced peak acid outputs that were 29.5 to 41.4% higher than the peak histamine output (p < 0.05 to < 0.01).

Discussion

We have been unable to find previous reports of the maximal secretory response to food in vagally innervated pouches. Code and Watkinson (1955) found that a standard meal of horse meat given as a single feed of 170 g or repeated feed of 15 g every 15 to 30 minutes produced about the same level of acid output as was achieved with maximal histamine infusion in Pavlov pouch dogs.

In a systematic study of dogs with vagally denervated (Heidenhain) pouches, Thomas and Forrest (1967) showed that the maximal acid output to a meal of meat and bone dust was about equal to the maximum elicited by histamine. Bone dust was added to meat because this mixture produced higher levels of gastric secretion than meat alone in dogs with denervated pouches (Öbrink, 1954). Large meals were required to duplicate the maximal response to histamine, approximately 80 g/kg as a single feeding or 5 to 10 g/kg every 15 minutes. Pentagastrin, a gastrin analogue, produced lower rates of secretion than histamine or food.

Our dogs with innervated pouches were fed a meal of liver and 5% bone dust similar to that given in the experiments of Thomas and Forrest (1967). These pouches, in response to considerably smaller meals (as low as 11 g/kg),
It is difficult to quantitate the gastric secretory response to feeding in man. Aspiration of the stomach contents after feeding, as in the Ewald test meal, is limited by solid food, alkaline duodenal reflux, buffering of acid by the food, and gastric emptying. Rune (1966) has attempted to estimate gastric acid secretion indirectly by measuring the elevation in blood base which accompanies the discharge of H+ ions into the gastric lumen. He found equal elevations in blood base in man during a meal and during histamine stimulation with simultaneous gastric aspiration. As he noted, food may have produced a greater output of acid than histamine since pancreatic secretion of bicarbonate during a meal would suppress the blood base elevation and lead to an underestimation of the acid secretory response to food.

There is controversy about the occurrence of potentiation of gastric acid secretion in various species. Johnson and Grossman (1969) have conclusively demonstrated in the dog that combinations of histamine and gastrin clearly exceed the highest rate of gastric acid secretion that can be achieved with either stimulant alone. Konturek and Olekzy (1967) found slight potentiation between histamine and pentagastrin in man. Checketts, Gillespie, and Kay (1968) showed that in 32 of 67 subjects insulin appeared to potentiate histamine. In a single subject studied by Makhlof, McManus, and Knill (1968) gastrin combined with histamine or mecothane produced the same calculated maximum as gastrin alone. This study has demonstrated that combinations of hormonal and neural stimuli, endogenously initiated by food, are capable of producing a higher rate of gastric secretion in dogs than can be achieved by a single exogenous stimulant (histamine) alone.

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References
Gillespie, I. E., and Grossman, M. I. (1964). Potentiation between bethanecol and histamine in Heidenhain pouch dogs. Andersson and Grossman (1965) showed potentiation between bethanecol and histamine in Pavlov pouch dogs. In all of these studies acid from the main stomach was entering the duodenum and it must be assumed that potentiation occurred despite duodenal acidification. Our histamine studies were conducted with acid diverted to the exterior through the open gastric fistula to prevent the possible release of inhibitors of histamine stimulated secretion (Code and Watkinson, 1955).
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