Defective opsonisation and complement deficiency in serum from patients with fulminant hepatic failure

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SUMMARY Serum from 23 of 26 patients with fulminant hepatic failure and grade IV encephalopathy had defective opsonisation of E. coli and yeast (S. cerevisiae). No toxic serum factors acting on the polymorphonuclear leucocytes or inactivators of the normal serum opsonisation factors were found. Complement deficiency was shown to be the most likely cause of the defect in opsonisation. The addition of a heat-labile fraction of normal serum at low concentration corrected the defect and factors of both the classical and the alternative pathways of complement were reduced to below 40% of the activity of the control serum. During the early stages of clinical recovery serum opsonisation and complement activity returned to normal with statistically significant correlations between tests of opsonisation and total haemolytic complement CH₅₀, C3 and total alternative pathway activity. Defective serum opsonisation and complement deficiency represent major defects in the body's defences against infection.

Bacterial infection is a common complication of fulminant hepatic failure and has been implicated as the main cause of death in 11% of fatal cases. Normally the removal of bacteria from the blood requires serum factors, immunoglobulin, and complement, which coat (opsonise) the bacteria before they are ingested and killed by the polymorphonuclear leucocytes. Congenital deficiency of individual complement factors or immunoglobulin markedly increases susceptibility to infection. In this paper we describe investigations into opsonisation by serum from patients with fulminant hepatic failure. Two organisms, Escherichia coli and yeast (Saccharomyces cerevisiae), which have different opsonisation requirements were used. We also investigated the relationship between the opsonisation defect measured in vitro and the deficiency in complement factors in the same patients.

Methods

Patients Serum samples were obtained serially from 26 patients with fulminant hepatic failure after they had developed grade IV encephalopathy. The cause of their liver damage was paracetamol self-poisoning in 14, viral hepatitis in nine (two HBsAg-positive), and multiple exposure to halothane in three. Seven patients (two hepatitis, five paracetamol self-poisoning) recovered completely. Full supportive therapy was given including intravenous dextrose, cimetidine, dexamethasone (for cerebral oedema), vitamin supplements, lactulose orally, and magnesium sulphate enemas. Liver support by means of daily periods of four to six hours' haemodialysis with a polyacrylonitrile membrane was needed in all patients, especially in 11 who developed renal failure (creatinine > 200 μmol/l). Three patients developed bacterial septicaemia after admission and a fourth had E. coli septicaemia on admission. Three of the four patients with infection survived and in the other the immediate cause of death was cerebral oedema rather than the infection. Antibiotics were given to three patients with proven infection and to nine with suspected bacterial infection. Venous blood samples were taken daily before haemodialysis, from the time of the first signs of grade IV encephalopathy until death or recovery. Blood was allowed to clot for two hours at room temperature before separation of serum, which was either used immediately or stored in aliquots at -196°C until the time of assay. Blood from 41 healthy medical and laboratory personnel aged between 18 and 40 years without evidence of liver disease or illness known to impair polymorphonuclear leucocyte function acted as controls. A
control serum pool was derived from the sera of six controls, mixed in equal proportions, and stored in aliquots at −196°C.

PREPARATION OF POLYMORPHONUCLEAR LEUCOCYTES FOR \textit{E. coli} AND YEAST OPSONISATION ASSAYS

After sedimentation of erythrocytes with dextran (average molecular weight 26,400), the leucocyte-rich plasma fraction was layered onto a Ficoll-Trisioil gradients and centrifuged at 400–450 g for 20 minutes at 10–15°C. The pellet containing polymorphonuclear leucocytes and erythrocytes was washed twice with a 1:2 mixture of ordinary and modified Hank’s balanced salt solution, separated by centrifugation and the residual erythrocytes ‘shock-lysed’ with distilled water for 10 seconds then made isotonic with equal volumes of 1.8% saline and ordinary Hank’s balanced salt solution. The polymorphonuclear leucocytes were separated by centrifugation and resuspended in modified Hank’s balanced salt solution. At least 90% of the cells isolated in this way were viable as judged by exclusion of trypan blue. The number of polymorphonuclear leucocytes was determined with a Neubauer chamber and a working suspension made with modified Hank’s balanced salt solution.

\textit{Escherichia coli} OPSONISATION ASSAY

Serum opsonisation of \textit{E. coli} was measured by the method of Quie et al. E. coli 089 in the late log-phase of growth, obtained by overnight culture at 30°C in trypticase soy broth, were washed twice with 0.9% saline at 4°C and the number of organisms estimated turbidimetrically. The reaction mixture containing 2.5×10\(^6\) polymorphonuclear leucocytes, 1×10\(^6\) \textit{E. coli}, and 40 µl of test serum in a final volume of 1 ml of Hank’s balanced salt solution was incubated in air in polystyrene tubes on a Matburn rotary mixer for one hour at 37°C. A 50 µl aliquot of reaction mixture was removed at zero and 60 minutes and the polymorphonuclear leucocytes lysed with 5 ml of cold distilled water. The number of viable \textit{E. coli} cells was determined by plating out in duplicate with the Miles-Misra technique onto MacConkey’s agar and the number of colony-forming units (CFU) counted after overnight incubation at 37°C.

Results were expressed as the opsonisation index:

\[
\frac{\text{Mean number of CFU [in duplicates] at 60 min}}{\text{Mean number of CFU [in duplicates] at zero min}} \times 100
\]

Control tubes included in each experiment contained: (a) \textit{E. coli} + buffer; (b) \textit{E. coli} + test serum; (c) \textit{E. coli} + polymorphonuclear leucocytes + normal serum; (d) \textit{E. coli} + polymorphonuclear leucocytes. The numbers of \textit{E. coli} in control tubes (a), (b) and (d) never varied by more than 2% during the hour of incubation. Serum bactericidal and bacteriostatic activity was excluded by controls (b). Control (c) was used both as a standard and to establish the normal range. Serum from patients who received antibiotics was dialysed against Hank’s balanced salt solution for 12 hours at 4°C and was shown to be without bactericidal or bacteriostatic effects by incubation with \textit{E. coli}.

In the presence of polymorphonuclear leucocytes and serum from one control, opsonisation studied on four separate occasions gave a mean opsonisation index of 98.72±SD 1.19. The variance for counting duplicate colony units was 3.16%.

YEAST OPSONISATION ASSAY

Yeast opsonisation was measured by the method of Levinsky et al. with a suspension of heat-inactivated bakers’ yeast (\textit{Saccharomyces cerevisiae}) in Hank’s balanced salt solution mixed with normal polymorphonuclear leucocytes (in a ratio of yeast: polymorphonuclear leucocytes 10:1) and 5% test serum in a final volume of 1 ml. After incubation at 37°C for 30 minutes in polystyrene tubes on a Matburn rotary mixer, the number of residual yeasts was counted with an electronic Coulter counter. Controls included in each experiment were (a) yeast + buffer; (b) yeast + polymorphonuclear leucocytes + buffer; (c) yeast + polymorphonuclear leucocytes + buffer + control serum pool. Control (a) provided a measure of the number of yeasts present in the reaction mixture after 30 minutes’ incubation. Control (b) was used both to confirm the inability of polymorphonuclear leucocytes to ingest unopsonised yeasts and as a measure of the cellular content of the reaction mixture.

The opsonisation index =

\[
\frac{\text{Number of residual yeast cells after 30 min}}{1 - \frac{\text{Number of residual yeast cells in control (b) after 30 min}}{\text{Number of residual yeast cells after 30 min}}} \times 100
\]

Results for test serum were expressed as a percentage of the activity of the control serum pool-control tube (c).

In the presence of polymorphonuclear leucocytes from one control, the control serum pool (studied on eight separate occasions) gave a mean opsonisation index of 77.75±SD 6.25 SEM 1.11. With polymorphonuclear leucocytes from 13 controls the control serum pool opsonised a mean of 78.2±SD 4.03 SEM 1.11 of the yeasts.
Defective opsonisation and complement deficiency in serum

SERUM COMPLEMENT AND IMMUNOGLOBULIN ASSAYS

Functional activity of C4, C5, total haemolytic complement (CH50), total alternative pathway, and factors B and D were determined by haemolytic diffusion in agarose.8 Immunoreactive C3 concentrations were determined by radial immunodiffusion against rabbit antisera (kindly provided by Professor D K Peters). Results were expressed as the percentage activity of the control serum pool included as a standard in each plate. For some experiments, aliquots of serum pool were depleted of active C3 (R3), factor B (RB) and C4 (R4) by the method described by Lachman et al.8 Levels of serum immunoglobulin IgG, and IgM were measured in the serum of 19 of the patients with specific antisera (Behring Diagnostics, Hoechst UK Ltd, Hounslow, UK) and a laser nephelometer.

Results

Serum opsonisation indices for both E. coli and yeasts were reduced in all but three patients. E. coli opsonised by serum from 12 controls were almost all killed by normal polymorphonuclear leucocytes, mean opsonisation index being 98.5±SD 1.2. Opsonisation by serum from 22 patients was significantly reduced (p<0.01, Wilcoxon’s rank sum test), although the results were spread over a wide range, mean 40.7±38.7. The mean yeast opsonisation index was also significantly reduced (p<0.01, Wilcoxon’s rank sum test), although, in common with E. coli opsonisation, results for patients’ serum were spread over a wide range, with a mean of 31.8±28.9 compared with 98.7±7.1 for the control sera (Table 1). No correlation was found between this defect in serum opsonisation index for either organism and the aetiology or clinical outcome of the hepatic necrosis.

The results of the serial measurements in 10 of the fatal cases showed a progressive reduction in opsonisation indices in eight while, in seven of the patients who recovered, serial studies were available during the period of improvement in five. In all cases serum opsonisation indices for both E. coli and yeasts returned to normal, in three patients between one and nine days after all signs of encephalopathy had disappeared (between two and 10 days after admission). There was, however, no correlation with the level of consciousness, some minor signs of encephalopathy still being present in two other patients when opsonisation indices had returned to normal.

Experiments with polymorphonuclear leucocytes from four patients, in the presence of autologous serum, showed the E. coli opsonisation index to be reduced, with a mean value of 31.5. In the presence of control serum pool the cells functioned normally, with a mean opsonisation index of 98.75.

CHARACTERISATION OF SERUM ABNORMALITY

Dialysis in vitro against Hank’s balanced salt solution for 12 hours at 4°C had no effect; mean yeast opsonisation index in the five patients studied was 60.5 before and 53 after dialysis (P>0.5, paired t-test). Similarly, serum obtained immediately before and after treatment by polycrylonitrile membrane dialysis in five patients gave mean opsonisation values for yeast of 56.3 and 39 respectively and for E. coli 88.3 and 76.3 respectively (Table 2). These findings indicate that the serum defect was not caused by a toxic substance of low molecular weight.

To examine for the action of a non-dialysable toxic serum factor, normal polymorphonuclear leucocytes were incubated for one hour at 37°C in a 10% dilution of serum from nine patients known to have defective opsonisation. When normal polymorphonuclear leucocytes treated in this way were incubated with E. coli or yeast cells already opsonised

Table 1 Initial measurements of serum opsonisation (mean±SD) at time of grade IV encephalopathy

<table>
<thead>
<tr>
<th>Opsonisation</th>
<th>Yeast</th>
<th>E. coli</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>%</td>
<td>%</td>
</tr>
<tr>
<td>Total series</td>
<td>37.1±8.9</td>
<td>40.7±8.7</td>
</tr>
<tr>
<td>Survivors</td>
<td>31.8±27</td>
<td>23.2±33.6</td>
</tr>
<tr>
<td>Fatalities</td>
<td>38.8±30.14</td>
<td>47.3±39.3</td>
</tr>
<tr>
<td>Paracetamol self-poisoning</td>
<td>38.6±26.5</td>
<td>31.3±30.3</td>
</tr>
<tr>
<td>Viral hepatitis</td>
<td>29.4±25.7</td>
<td>55.4±42.6</td>
</tr>
<tr>
<td>Controls</td>
<td>98.7±7.1</td>
<td>98.5±1.2</td>
</tr>
</tbody>
</table>

Figures in parentheses show number of patients studied.

Table 2 Measurements of serum opsonisation before and after four hours of polycrylonitrile haemodialysis in five patients

<table>
<thead>
<tr>
<th>Patient</th>
<th>Aetiology</th>
<th>Opsonisation index</th>
<th>Before</th>
<th>After</th>
<th>Before</th>
<th>After</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Yeast</td>
<td>E. coli</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Hepatitis</td>
<td>105</td>
<td>54</td>
<td>92</td>
<td>71</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Halothane</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Hepatitis</td>
<td>72</td>
<td>70</td>
<td>82</td>
<td>91</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Hepatitis</td>
<td>0</td>
<td>0</td>
<td>—</td>
<td>—</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Paracetamol</td>
<td>48</td>
<td>32</td>
<td>—</td>
<td>—</td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>56.3</td>
<td>39</td>
<td>88.3</td>
<td>76.3</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
by preincubation in control serum pool, a mean of 99.3% _E. coli_ and 100% yeast cells were phagocytosed, indicating no impairment of leucocyte function.

To determine whether the defect was due to a deficiency of normal opsonisation factors, serum from patients and from control serum pool were mixed together in different proportions. Severe defects of _E. coli_ opsonisation by serum from three cases were totally corrected by the addition of control serum pool at concentrations of 0.25, 0.33, and 2%. As control serum pool in a concentration of 1% in the absence of patients’ serum was only able to opsonise 35% of _E. coli_, the contribution by patients’ serum in these experiments is greater than immediately apparent. Similar studies of yeast opsonisation by serum from three other patients showed that the defect could be totally corrected by addition of 0.5 and 1% control serum pool (Table 3), while control serum pool at a concentration of 0.5 and 1% in the absence of patients’ serum opsonised only 14 and 53% of yeasts respectively (Table 4). Heating of control serum to 56°C for 30 minutes abolished the beneficial effect even at a concentration as high as 5%.

**Table 3** Effects of dilutions of control serum on yeast opsonisation by serum from three patients

<table>
<thead>
<tr>
<th>Final concentration (%) serum in reaction mixture</th>
<th>Control serum</th>
<th>Patients’ serum</th>
<th>Serum of patient 1</th>
<th>Serum of patient 2</th>
<th>Serum of patient 3</th>
<th>Control serum diluted with buffer</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0.5</td>
<td>1</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>4.5</td>
<td>4</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Serum of patient 1</td>
<td>56</td>
<td>100</td>
<td>98</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Serum of patient 2</td>
<td>75</td>
<td>99</td>
<td>104</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Serum of patient 3</td>
<td>75</td>
<td>98</td>
<td>105</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control serum diluted with buffer</td>
<td>0</td>
<td>14.3</td>
<td>53</td>
<td>100</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Numbers do not denote the same patients as in Table 2.

**Table 4** Initial measurements of serum complement expressed as percentage activity of serum pool (mean ± SD)

<table>
<thead>
<tr>
<th>Clinical groups</th>
<th>Factor B</th>
<th>Factor D</th>
<th>Alternative pathway</th>
<th>Total haemolytic complement (CH50)</th>
<th>CH3</th>
<th>CH4</th>
<th>CH5</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total series</td>
<td>26.4±25.9</td>
<td>29.1±28.8</td>
<td>22.6±17.2 (25)</td>
<td>12.3±15.1 (23)</td>
<td>23.8±11.6 (25)</td>
<td>15.9±13.1 (24)</td>
<td>15.4±17.1 (15)</td>
</tr>
<tr>
<td>Fatalies</td>
<td>25.2±28.8</td>
<td>23.0±30.9</td>
<td>21.6±17.5 (18)</td>
<td>13.5±15.9 (17)</td>
<td>24.1±13.1 (18)</td>
<td>15.9±12.9 (18)</td>
<td>18.3±19.5 (10)</td>
</tr>
<tr>
<td>Survivors</td>
<td>29.7±17.6</td>
<td>39.2±23.8</td>
<td>25.6±17.5 (7)</td>
<td>9.3±13.7 (7)</td>
<td>23.2±7.4 (6)</td>
<td>16.2±14.8 (9)</td>
<td>9.6±10 (5)</td>
</tr>
<tr>
<td>Paracetamol self-poisoning</td>
<td>26.7±24.5</td>
<td>31.1±24.2</td>
<td>18.5±17.1 (14)</td>
<td>6.4±8.4 (14)</td>
<td>20.4±8.6 (14)</td>
<td>11.4±11.8* (15)</td>
<td>8.8±12.1 (4)</td>
</tr>
<tr>
<td>Viral hepatitis</td>
<td>32.1±31.7</td>
<td>37.0±37.7</td>
<td>26.8±20.1 (8)</td>
<td>26.4±18.6 (7)</td>
<td>32.3±14 (8)</td>
<td>23.9±12.1* (7)</td>
<td>33.8±18.9 (4)</td>
</tr>
</tbody>
</table>

Figures in parentheses represent number of patients studied.

*0.05 > p > 0.02 (Wilcoxon's rank sum test).
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GRADE OF ENCEPHALOPATHY:
4 4 4 3 1

DAYS AFTER ADMISSION

OPSONISATION INDEX

Figure Serial values of serum opsonisation and complement levels in a case of paracetamol self-poisoning while in grade IV encephalopathy and during the acute stages of recovery. □□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□□.png
of normal serum needed to correct the opsonisation defect is consistent with complement deficiency rather than immunoglobulin as the cause of the defective opsonisation. Additional evidence is the reduction of complement factors in the sera of patients at a time when opsonisation defects were most markedly affected and the good correlations between improvement in serum opsonisation and complement activity during the acute stages of recovery. The correlation between the recovery of the alternative pathway activity and E. coli opsonisation should be interpreted with caution, in view of the correction achieved by the addition of normal serum depleted of factor B (RB), which suggests a defect in the classical pathway.

The liver is the major site of synthesis of complement factors and the fact that serum complement and opsonisation returned to normal during recovery is evidence that these defects are not congenital but secondary to the hepatic necrosis. Markedly lowered complement levels have been reported in patients with acute hepatic necrosis and less severe defects in viral hepatitis due to reduced synthesis or increased catabolism. In our patients consumption of complement may have been increased especially in cases with acute viral hepatitis in whom circulating immune complexes are raised. Similar defects in complement and opsonisation, however, were present in hepatic necrosis caused by paracetamol self-poisoning (D Vergani, personal communication) in which immune complexes are almost always absent. The complement deficiencies in our patients are severe and affect several factors to a degree associated with impaired host defence against infection in other congenital and acquired complement deficiency states.

The importance of these findings is shown by the occurrence of bacterial infection in four (15%) of the cases at a time when serum opsonisation and complement factors were severely reduced. Reports of congenital deficiency of serum opsonisation suggest that transfusions of fresh blood or freshly prepared plasma provide the most effective replacement therapy. While opsonisation returned to normal in one patient, the failure of small transfusions of fresh frozen plasma in other cases may have been due to the insufficient volume of the transfusion or to the fact that the plasma had been stored and was not freshly separated. As in other diseases, infection can be overwhelming in the presence of such defects of host defence and attempts to correct these in fulminant hepatic failure should prove worthwhile.

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References


Defective opsonisation and complement deficiency in serum


