Gut-hormone profile in totally pancreatectomised patients

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SUMMARY In eight totally pancreatectomised patients the release of the relevant gut hormones was determined after a standard test meal. Plasma levels of pancreatic glucagon were not significantly different from zero in our series of pancreatectomised patients. Pancreatic polypeptide was undetectable. These findings imply the absence of a significant number of normally functioning alpha cells and pancreatic polypeptide cells in extrapancreatic sites in man. Consistent with the antrectomy, duodenectomy, and resection of the upper jejunum that are performed in conjunction with a total pancreatectomy the gastrin release was significantly impaired. In contrast there was a striking postprandial rise in enteroglucagon probably induced by the rapid intestinal transit time often seen after partial gastrectomy. In contrast plasma motilin and GIP levels were normal. Pancreatectomised man thus presents an interesting model of total deficiency of endogenous insulin, pancreatic polypeptide, and pancreatic glucagon and, in addition, greatly diminished gastrin. The considerable derangement of metabolic and intestinal function that follows total pancreatectomy may, in part, be explained by this gross disturbance of the normal physiology of gut hormone.

The main indications for total pancreatectomy are carcinoma (ampullary, periampullary and pancreatic) and end-stage chronic pancreatitis. The major clinical problems that occur after total pancreatectomy arise from the pancreatic exocrine and endocrine insufficiencies that invariably follow the procedure. The resulting diabetes can be difficult to control in this clinical setting. The greatest clinical hazard in the management of apancreatic diabetes is insulin shock. Difficulty can be encountered in controlling the steatorrhoea despite an adequate replacement dose of pancreatic enzymes. A considerable number of patients fail to gain, and many even lose, weight after total pancreatectomy. Many factors may be relevant to this and it is difficult to single out whether it is the accompanying partial gastrectomy, the pancreatectomy, insufficient enzyme replacement, or the level of caloric intake which is responsible.

Both pancreatic exocrine and endocrine function are profoundly influenced by gut hormones and feedback mechanisms may exist. Thus the degree and clinical severity of the metabolic sequelae and the altered intestinal function that follow total pancreatectomy may well be influenced by abnormalities in the release of gastrointestinal hormones. The partial gastrectomy, duodenectomy, and resection of the upper jejunum that are usually performed in conjunction with a total pancreatectomy may well result in at least partial removal of the source of some gut hormones. We have therefore compared the release of the relevant gut hormones in eight totally pancreatectomised patients, and in 11 age- and sex-matched normal control subjects.

Methods

Patients The stimulus for release of gut hormones was a standard test-breakfast consisting of two medium-sized boiled eggs, 60 g bread as toast, 10 g butter, 35 g marmalade, and 150 ml unsweetened orange juice (containing a total of 18 g protein, 22 g fat, and 66 g carbohydrate equivalent to 530 Calories). Eight totally pancreatectomised patients (Table 1) were studied before their normal morning dose of insulin, without pancreatic enzyme supplement, and also 11 healthy controls (six males and five females), with a mean age of 43 (range 22–59) years and an average weight of 107% of ideal, none of whom had a present or past history of gastrointestinal disease. The efficacy of total pancreatectomy was confirmed

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Table 1  Clinical data on eight totally pancreatectomised patients

<table>
<thead>
<tr>
<th>Patient no.</th>
<th>Sex and age (yr)</th>
<th>Weight (kg)</th>
<th>Height (cm)</th>
<th>Insulin (units/day)</th>
<th>Total pancrectomy: reason and date</th>
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<tbody>
<tr>
<td>1</td>
<td>F54</td>
<td>45.5</td>
<td>164</td>
<td>20</td>
<td>Chronic pancreatitis 1976</td>
</tr>
<tr>
<td>2</td>
<td>F65</td>
<td>50.9</td>
<td>163</td>
<td>20</td>
<td>Adenocarcinoma 1965</td>
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<tr>
<td>3</td>
<td>M39</td>
<td>49.3</td>
<td>163</td>
<td>18</td>
<td>Chronic pancreatitis 1977</td>
</tr>
<tr>
<td>4</td>
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<td>71.1</td>
<td>169</td>
<td>32</td>
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<tr>
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<td>F38</td>
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<tr>
<td>6</td>
<td>M38</td>
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<td>184</td>
<td>44</td>
<td>Chronic pancreatitis 1977</td>
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<tr>
<td>7</td>
<td>M35</td>
<td>68.0</td>
<td>172</td>
<td>40</td>
<td>Chronic pancreatitis 1976</td>
</tr>
<tr>
<td>8</td>
<td>M36</td>
<td>75.0</td>
<td>180</td>
<td>44</td>
<td>Adenocarcinoma 1977</td>
</tr>
</tbody>
</table>

Table 2  Hormone release and blood glucose in pancreatectomised patients

<table>
<thead>
<tr>
<th>Hormone</th>
<th>Controls</th>
<th>Total pancrectomy</th>
<th>P†‡</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gastrin*</td>
<td>B (pmol/l)</td>
<td>5±0.5</td>
<td>2±0.2</td>
</tr>
<tr>
<td></td>
<td>PR (pmol/l)</td>
<td>17±4</td>
<td>3±0.6</td>
</tr>
<tr>
<td></td>
<td>IIR (pmol/l)</td>
<td>1.7±0.3</td>
<td>0.2±0.1</td>
</tr>
<tr>
<td>GIP*</td>
<td>B (pmol/l)</td>
<td>9±1</td>
<td>16±3</td>
</tr>
<tr>
<td></td>
<td>PR (pmol/l)</td>
<td>33±7</td>
<td>46±12</td>
</tr>
<tr>
<td></td>
<td>IIR (pmol/l)</td>
<td>3.9±0.7</td>
<td>5.3±2.1</td>
</tr>
<tr>
<td>HPP*</td>
<td>B (pmol/l)</td>
<td>15±3</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td>PR (pmol/l)</td>
<td>116±19</td>
<td>—</td>
</tr>
<tr>
<td></td>
<td>IIR (pmol/l)</td>
<td>12.9±2.4</td>
<td>—</td>
</tr>
<tr>
<td>Pancreatic glucagon†</td>
<td>B (pmol/l)</td>
<td>9±2</td>
<td>5±0.6</td>
</tr>
<tr>
<td></td>
<td>PR (pmol/l)</td>
<td>3±2</td>
<td>3±0.9</td>
</tr>
<tr>
<td></td>
<td>IIR (pmol/l)</td>
<td>0.3±0.3</td>
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<td>Motilin*</td>
<td>B (pmol/l)</td>
<td>63±8</td>
<td>41±10</td>
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<td></td>
<td>PR (pmol/l)</td>
<td>16±6</td>
<td>48±6</td>
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<tr>
<td></td>
<td>IIR (pmol/l)</td>
<td>-1.1±0.9</td>
<td>49±0.9</td>
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<tr>
<td>Enteroglucagon*</td>
<td>B (pmol/l)</td>
<td>32±7</td>
<td>38±10</td>
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<tr>
<td></td>
<td>PR (pmol/l)</td>
<td>26±7</td>
<td>89±23</td>
</tr>
<tr>
<td></td>
<td>IIR (pmol/l)</td>
<td>2.6±1.0</td>
<td>8.9±1.9</td>
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<tr>
<td>Neurotensin*</td>
<td>B (pmol/l)</td>
<td>50±6</td>
<td>49±7</td>
</tr>
<tr>
<td></td>
<td>PR (pmol/l)</td>
<td>22±3</td>
<td>42±14</td>
</tr>
<tr>
<td></td>
<td>IIR (pmol/l)</td>
<td>1.8±0.4</td>
<td>4.3±2.2</td>
</tr>
<tr>
<td>Blood glucose*</td>
<td>B (mmol/l)</td>
<td>4.2±0.2</td>
<td>11.6±2.2</td>
</tr>
<tr>
<td></td>
<td>PR (mmol/l)</td>
<td>1.6±0.3</td>
<td>14.7±1.8</td>
</tr>
<tr>
<td></td>
<td>IIR (mmol/l)</td>
<td>7.1±36</td>
<td>2.0±0.3</td>
</tr>
</tbody>
</table>

All hormone levels (B: basal, PR: peak response) given as pmol/l (mean±SEM); blood glucose levels given as mmol/l (mean±SEM).

*After test-breakfast—integrated incremental response (IIR) in pmol/l (mean±SEM) over 180 minutes; blood glucose IIR in mmol/l (mean±SEM) over 180 minutes.

†P values vs. controls: a: <0.001, b: <0.005, c: <0.01, d: <0.02, e: <0.05. NS: not significant.

by undetectable basal plasma levels of C-peptide. The operation of total pancreatectomy involved block resection of the pancreas, the duodenum, two-thirds of the distal stomach (Billroth II—resection), the upper part of the jejunum, the lower end of the common bile duct, the gall bladder, and the spleen, in conjunction with a gastro-jejunostomy.

In patient no. 1 large parts of the jejunum were bypassed by a distal enterostomosis. After this study, this patient suffered from massive steatorrhea, despite adequate enzyme replacement, and intractable diarrhoea and died from an intercurrent infection.

Blood samples for subsequent hormone assays were taken into heparinised tubes and 400 kallikrein-inactivating units of aprotinin (Trasylol) per ml were added. Plasma was separated within 15 minutes of sampling and stored at -20°C until assay.

Hormone radioimmunoassays were carried out by conventional methods with antisera raised to pure natural human gastrin and pancreatic polypeptide, to porcine gastric inhibitory polypeptide (GIP) total glucagon (N terminal), pancreatic glucagon (C terminal), motilin and to bovine neurotensin.

The assays could detect the following plasma changes with 95% confidence: gastrin 2 pmol/l, pancreatic polypeptide 4 pmol/l, GIP 8 pmol/l, pancreatic glucagon 1.5 pmol/l, enteroglucagon 8 pmol/l, neurotensin 5 pmol/l, and motilin 3 pmol/l. No cross-reaction was detectable between any of the peptides assayed.

Results

Basal (fasting) concentrations, maximum concentrations after the appropriate stimulus, total post-prandial integrated release of each hormone, and blood glucose are given in Table 2.

**Pancreatic Glucagon**

The pancreatectomised patients had a mean fasting pancreatic glucagon concentration of 16±1.5 pmol/l, which was not significantly different from zero. The control group had a significantly greater mean fasting glucagon concentration of 7.6±2.3 pmol/l (p<0.05). In neither group was there a significant change from basal levels in the post meal glucagon concentrations.

**Gastric Inhibitory Polypeptide** (Fig. 1)

Normal subjects had a significant post-prandial rise in plasma GIP to 31.6±6.5 pmol/l at 60 minutes. Patients with total pancreatectomy had an equivalent peak of 45.2±7.0 pmol/l. The total integrated
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Fig. 1 Plasma gastric-inhibitory-polypeptide response to a test-meal in totally pancreatectomised patients.

The release of GIP after the meal was not significantly different from that in the controls.

Patient no. 1 had plasma GIP concentrations considerably higher than the rest of the group, accounting for the large standard errors of the means.

**ENTEROGLUCAGON** (Fig. 2)

Basal plasma-enteroglucagon concentrations were similar in the two groups. After the meal there was a much larger increase to 124.1 ± 22.9 at 60 minutes in the pancreatectomised patients in contrast with 40.9 ± 6.8 pmol/l (P < 0.005) in the normal subjects. The three hour total integrated release of enteroglucagon after the meal was also significantly increased in pancreatectomised patients (15.6 ± 2.1 nmol/l) compared with controls (8.3 ± 1.1 nmol/l, P < 0.01).

**NEUROTENSIN**

Though basal and post-breakfast neurotensin concentrations were not significantly different from normal controls the mean peak rise and total integrated decrease were higher in the pancreatectomised patients (42.0 ± 14.0 pmol/l and 43 ± 2.2 nmol/l) than in the controls (22 ± 3 pmol/l and 1.8 ± 0.4).

**PANCREATIC POLYPEPTIDE**

Pancreatic polypeptide was undetectable in the pancreatectomised patients, whereas the normal group had a maximal post-prandial rise in plasma pancreatic polypeptide of 116 ± 19 pmol/l.

**GASTRIN** (Fig. 3)

The pancreatectomised group of patients had significantly lower basal gastrin concentrations of 1.8 ± 0.2 pmol/l compared with controls (5.0 ± 0.5 pmol/l, P < 0.001) with a greatly reduced post-prandial rise of only 2.1 ± 0.4 pmol/l (controls: 12.3 ± 3.1 pmol/l, P < 0.005).

**MOTILIN**

Plasma-motilin and total integrated release of this hormone after the test-breakfast were similar in the two groups.
Discussion

Totally pancreatectomised man has been previously studied as a useful model for diabetes without glucagon. As in this earlier study with a different group of pancreatectomised patients, use of a highly sensitive and specific antibody showed plasma levels of pancreatic glucagon to be not significantly different from zero. The report of basal glucagon immunoreactivity in a totally pancreatectomised patient may reflect the lesser specificity of the antibody used. Pancreatic polypeptide was undetectable in our series of pancreatectomised patients. This is consistent with previous observations that pancreatic polypeptide is found in specific cells of both the endocrine and exocrine pancreas, and that pancreatic polypeptide cells are very rare in the intestine. In man the plasma concentration of the hormone varies with the prandial state. Ingestion of a protein-rich meal induces a brisk, biphasic secretion pattern comparable with post-prandial release of insulin.

The physiological actions of pancreatic polypeptide are not fully elucidated but experiments in dogs suggested that it affected the biliary tract, the exocrine pancreas and gastric secretion. Recent experiments in man, using physiological doses of infused bovine pancreatic polypeptide, indicated that it inhibited bilirubin and trypsin, but not gastric acid or pepsin. As both known target organs (pancreas and gall bladder) are resected in these patients, the effect of pancreatic polypeptide is difficult to interpret. However, until its physiology and pathophysiology are fully understood, the possible consequences of a total lack of pancreatic polypeptide cannot be determined.

The significantly impaired gastrin release after total pancreatectomy is consistent with antrectomy. In addition, the duodenectomy and resection of the upper jejunum diminish the main sources of extragastric gastrin. The incidence of gastrointestinal bleeding and ulcer disease after total or partial pancreatectomy seems to be low in spite of potentially impaired ability to neutralise acid in the duodenum consequent on impaired bicarbonate secretion. Observations by Plam et al. and Mangold et al. observed no gastrointestinal bleeding or perforated ulcer in a group of 24 totally pancreatectomised and 44 partially pancreatectomised subjects. We have observed 13 pancreatectomised subjects who also developed no ulcer disease (unpublished observations). Others have reported similar findings. Increased antral or extragastric gastrin release after stimulation is believed to play a role in the pathophysiology of duodenal and gastric ulcer disease. Thus, it is tempting to speculate that in many of the cases the low incidence of gastrointestinal bleeding may be at least partly secondary to the impaired post-prandial gastrin-release secondary to the extensive surgery undertaken during pancreatectomy.

There was a striking post-prandial rise in enteroglucagon in the pancreatectomised subjects. Increased release of enteroglucagon has also been reported in coeliac disease and in the dumping syndrome. Enteroglucagon is found in highest concentrations in the jejunum and it is thought to stimulate mucosal growth and to slow intestinal transit. In coeliac disease it has been claimed that the increased mucosal turnover and the slower transit time may be secondary to the raised enteroglucagon concentrations. In our study, the raised enteroglucagon levels were probably stimulated by the rapid intestinal transit time often seen after partial gastrectomy. In patient no. 1 large parts of the jejunum (about 1-5 m) were bypassed by a distal Roux-enteroanastomosis. This patient had basal enteroglucagon values of 327.7 ± 18.8 and a peak of 381 pmol/l. In contrast, however, the release of neurotensin, another hormone produced in the distal part of the small intestine, was normal in the totally pancreatectomised patients.

The totally pancreatectomised patients had plasma motilin levels which were not significantly different from normal. This is in contrast with the raised motilin levels found in patients with both mild and severe pancreatic disease. The latter subjects, however, have intact stomachs and upper small intestine—those areas where motilin exerts its actions, increasing the rate of gastric emptying and initiating the induction of interdigestive myoelectric complexes. The pancreatectomised patients studied here had also undergone duodenectomy and proximal jejunectomy. These areas contain the highest concentration of motilin cells. The finding of normal plasma motilin concentrations in these patients may reflect a compensatory increase in the rate of secretion of the few remaining motilin cells.

Basal levels of plasma GIP and the post-prandial GIP response in the pancreatectomised patients were not significantly different from normal, despite one patient having high levels. The duodenum and jejunum are the areas with the highest concentrations of GIP cells and thus, after surgery, the pancreatectomised patients will have lost a significant portion of the GIP secreting cells. The normal plasma GIP levels in these patients presumably reflects increased secretion by GIP cells further down the small intestine. One mechanism which might stimulate this is the lack of endogenous insulin, as it has been proposed that insulin normally inhibits GIP release by means of a negative feedback mechanism.
Pancreatectomised man thus presents an interesting model of a total deficiency of endogenous insulin, pancreatic polypeptide, and pancreatic glucagon. In addition, there is also a greatly diminished mass of gastrin cells and, to a lesser degree, GIP and motilin cells. The considerable metabolic and intestinal derangements that follow total pancreatectomy may well reflect this gross disturbance of normal gut hormone physiology. Proper treatment of such patients thus depends on the recognition of the comprehensive nature of these abnormalities.

References