Interaction of glucagon and pentagastrin on pepsin secretion in healthy subjects*

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SUMMARY The effect of pentagastrin in step-wise increasing doses of 0·02, 0·2, 2·0, and 20 nmol/kg/h (0·01, 0·1, 1·0, and 10·0 μg/kg/h) on pepsin and acid secretion was studied in seven healthy subjects. The study was repeated on another day during infusion of glucagon in a dose of 103 pmol/kg/h (0·36 μg/kg/h) which results in plasma-glucagon concentrations comparable with those seen after a protein-rich meal. Pepsin output was maximal after 0·2 nmol/kg/h (0·1 μg/kg/h) of pentagastrin and 20 nmol/kg/h (10 μg/kg/h) resulted in a marked decrease. The dose of pentagastrin required for half-maximal pepsin output was less than 0·1 nmol/kg/h (0·05 μg/kg/h). When the study was repeated during infusion of glucagon, the dose-response curve was shifted to the right. The highest pepsin output was obtained with 20 nmol/kg/h (10 μg/kg/h) of pentagastrin and D₅₀ increased to well over 1 μg/kg/h. The dose of pentagastrin required for half-maximal acid secretion was about 0·3 nmol/kg/h (0·15 μg/kg/h) indicating that the sensitivity of the chief cells to pentagastrin is more than three times that of the parietal cells.

There is evidence that pancreatic glucagon participates in the physiological inhibition of gastric acid secretion.¹ ² Whether glucagon is involved in the physiological control of pepsin secretion also is unknown, as previous studies on glucagon and pepsin secretion have been performed with supra-physiological doses of glucagon.³ ⁴ ⁵

The present study investigates the dose-response relationship of pentagastrin and pepsin secretion and the effect on the dose-response curve of glucagon administered exogenously in a dose resulting in plasma concentrations comparable with those seen after a protein-rich meal.

Methods

SUBJECTS

Seven healthy volunteers, four men and three women with a median age of 26 years (21–34 years), were studied. Informed consent was obtained in every case.

EXPERIMENTAL PROCEDURE

Each subject was studied twice on separate days. On the first day a step dose-response study of the effect of pentagastrin on gastric acid and pepsin secretion was performed. After an overnight fast, a Levin tube was positioned under fluoroscopic control and the stomach was emptied. Through a thin polyvinyl tube welded to the Levin tube ⁵¹Cr-EDTA dissolved in 0·9% saline was infused with a flow of 30 ml/h in order to determine recovery. Basal secretion was collected for one hour. Pentagastrin was administered by continuous intravenous infusion in doses of 0·02, 0·2, 2·0, and 20 nmol/kg/h (0·01, 0·1, 1·0, and 10·0 μg/kg/h), each dose being administered for one hour. (These doses of pentagastrin were the same as those used in a previous study of pentagastrin glucagon interaction on acid secretion.¹) On the second day the step dose-response study with pentagastrin was repeated but now on a background infusion of glucagon in a dose of 103 pmol/kg/h (0·36 μg/kg/h). Glucagon Novo was diluted in saline containing, in addition, 1% human albumin, which prevents adherence of the hormone to glass syringes and plastic tubes. Infusion of 103 pmol/kg/h (0·36 μg/kg/h) with this technique results in the same plasma glucagon concentrations as infusion of 460 pmol/kg/h (1·6 μg/kg/h) without addition of albumin to the diluent, a dose which in a previous study was
calculated to be $D_{50}$ for inhibition of pentagastrin stimulated acid secretion in healthy subjects and which gave plasma glucagon concentrations comparable with those seen after a protein-rich meal.\(^1\) The glucagon infusion was started one hour before the infusion of pentagastrin. Through a cubital vein blood samples for analysis of pancreatic glucagon and glucose were taken every 15 minutes. All infusions were administered with a flow of 30 ml/h. On the first day isotonic saline containing 1% albumin was administered intravenously as a control infusion.

**LABORATORY ANALYSIS**

The volume of gastric secretion was measured for each 15 minute period and the concentration of $H^+$ was determined by titration to pH 7.0 with an autotitrator (Radiometer, Copenhagen, Denmark). $^{51}$Cr-EDTA was determined in a well counter and the volume of secretion and the output of $H^+$ and pepsin corrected according to the actual recovery. As a control of duodenogastric reflux, the osmolarity of each sample was determined by freezing point reduction.

Pepsin in gastric juice was measured using a modification of the Hunt method for estimating peptic activity in gastric juice.\(^6\,7\) The analysis determines the total peptic activity of the gastric juice at pH 1.9 with freeze dried human citrated plasma as protein substrate. The standard used was crystalline pepsin (Mann Research Lab.).

The plasma concentration of pancreatic glucagon was determined radioimmunoochemically. The glucagon antiserum (4305) is highly specific for pancreatic glucagon and cross-reacts less than 0.1% with high concentrations of enteroglucagon, and does not cross-react with secretin, VIP, GIP, cholecystokinin, or gastrin. Detection limit of the assay is 5 pmol/l (17 pg/ml), the within-assay coefficient of variation is 16%, and measurements of glucagon in dilutions of plasma with added glucagon in different concentrations yielded results which deviated less than 10% from the expected results.\(^8\)

Blood glucose was determined by the hexokinase method.\(^9\) Pepsin and acid output during the last 30 minutes of each hour were plotted against dose of pentagastrin after subtraction of basal secretion — that is, dose 0 corresponds to response 0. The approximate dose of pentagastrin required for half-maximal pepsin and acid secretion ($D_{50}$) was estimated from the dose-response curves. The $D_{50}$ for pentagastrin stimulation of acid secretion was also calculated by linear transformation of the Michaelis-Menten equation according to Dowd-Riggs\(^10\) using the formula: $V = V_{max} - D_{50}$ ($V/D$), where $V$ is the response, $V_{max}$ is the calculated maximal parietal cell response, and $D$ is the administered dose of pentagastrin. These calculations were performed on the individual data after subtraction of basal values by computerised estimation of the regression line according to the method of least squares. For the statistical analysis Student's $t$ test for paired observations was used. Values are given as mean ± SEM.

**Results**

Pepsin and acid output are shown in the Table. Pepsin output relative to maximum output during stepwise increasing doses of pentagastrin is shown in Fig. 1 (upper curve). Maximum output (37±4 U/30 min) was reached during infusion of 0.1 μg/kg/h of pentagastrin and was maintained during infusion of 1 μg/kg/h, while increasing the dose to 10 μg/kg/h resulted in a significant decrease in pepsin output ($p<0.01$). From the dose-response curve $D_{50}$ for pentagastrin was found to be less than 0.05 μg/kg/h. On a background infusion of glucagon the dose-response curve was shifted to the right (Fig. 1, lower curve). The highest pepsin output was the same as during infusion of pentagastrin alone (35±4 U/30 min, $p<0.01$) but was obtained only with the highest dose of pentagastrin (20 nmol/kg/h; 10 μg/kg/h), and no dose reversal (inhibition by larger doses) was observed. $D_{50}$ for pentagastrin during glucagon infusion was found to be well over 2 nmol/kg/h (1 μg/kg/h).

Acid output was significantly reduced during the combined infusion of pentagastrin and glucagon compared with infusion of pentagastrin alone (Fig. 2). Maximum output obtained decreased from 17±5 to 8±2 mmol/30 min ($p<0.01$). $D_{50}$ for acid secretion was approximately 0.3 nmol/kg/h (0.015 μg/kg/h).

Recovery of gastric juice ranged between 81 and 97% in all experiments. During infusion of pentagastrin and saline the osmolarity of gastric juice was 243±8 mmol/l and during infusion of pentagastrin and glucagon 251±7 mmol/l ($p>0.10$).

<table>
<thead>
<tr>
<th>Dose of pentagastrin (μg/kg/h)</th>
<th>Pepsin output (meq H+/30 min)</th>
<th>Acid output (U/30 min)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>−Glucagon +Glucagon</td>
<td>−Glucagon +Glucagon</td>
</tr>
<tr>
<td>0-01</td>
<td>5-1 (1-8) 3-5 (1-3)</td>
<td>15-1 (1-9) 14-2 (2-0)</td>
</tr>
<tr>
<td>0-1</td>
<td>8-2 (1-6) 4-2 (1-2)</td>
<td>37-1 (3-9) 17-4 (2-5)</td>
</tr>
<tr>
<td>0-5</td>
<td>16-8 (2-1) 7-9 (1-6)</td>
<td>35-2 (3-6) 17-8 (2-1)</td>
</tr>
<tr>
<td>10-0</td>
<td>17-0 (5-0) 8-2 (1-7)</td>
<td>21-0 (2-8) 35-0 (2-9)</td>
</tr>
</tbody>
</table>

Table: Pepsin and acid output during increasing doses of pentagastrin with and without infusion of glucagon (0-36 μg/kg/h) (mean (SEM))
Mean plasma glucagon concentration during the combined infusion of pentagastrin and glucagon was 154±17 pmol/l. Mean preinfusion blood glucose was 5.1±0.3 mmol/l and rose during glucagon infusion to 6.3±0.5 mmol/l (p<0.05).

Discussion

The present study shows that intravenous infusion of glucagon in a dose resulting in plasma concentrations comparable with postprandial levels previously reported reduces the effect of low doses of pentagastrin on pepsin secretion, whereas the effect of a high dose (20 nmol/kg/h; 10 μg/kg/h) is unchanged. Analysis of the data according to Michaelis-Menten kinetics is not possible but from the dose response curve D50 for pentagastrin was found to be less than 0.1 nmol/kg/h (0.05 μg/kg/h). During infusion of glucagon the D50 was considerably higher, but it is not possible from the data to give any exact value. The observations indicate, however, that glucagon markedly reduces the potency of pentagastrin for pepsin secretion.

The D50 of pentagastrin for acid secretion was found to 0.3 nmol/kg/h (0.15 μg/kg/h), which is of the same order of magnitude as found in a previous study; this means that the sensitivity of the chief cells to pentagastrin is more than three times that of the parietal cells.

These findings contrast with studies by others, who found the same D50 of pentagastrin for both acid and pepsin secretion in healthy subjects as well as in duodenal ulcer patients. Thodleifsson and Wormsley found maximal pepsin output after a pentagastrin dose of 2 nmol/kg/h (1 μg/kg/h) and dose reversal after 8 nmol/kg/h (4 μg/kg/h), whereas in another study maximum pepsin output was obtained with pentagastrin doses ranging from 0.06-3 nmol/kg/h (0.03-1.5 μg/kg/h).

The present study, combined with evidence from other studies, suggests that examination of pepsin secretion during stimulation with pentagastrin in doses used for maximal acid stimulation may be quite unreliable, as they have been performed in the dose-response range where the dose of pentagastrin is highly supramaximal for pepsin secretion, and where an unpredictable degree of dose reversal occurs.

The effect of glucagon on pentagastrin stimulated pepsin secretion consisted in a shift of the dose response curve to the right and prevention of the dose reversal after 20 nmol/kg/h (10 μg/kg/h) of pentagastrin. How this latter observation should be interpreted is at present obscure.

Bolus injection of glucagon in pharmacological doses (7 nmol/kg; 25 μg/kg) in man had no effect on unstimulated pepsin secretion, whereas histamine stimulated pepsin output was inhibited by 60% after 575 nmol (2 mg) glucagon. If the dose-response relationship for histamine stimulated acid and pepsin secretion is comparable with that of pentagastrin, the validity of the latter data may be doubtful.

The structure of glucagon resembles that of secretin, which, however, stimulates pepsin secretion in man, whereas both hormones inhibit acid secretion. Furthermore, acid perfusion of the duodenum stimulates pepsin secretion, suggesting a physiological role for secretin or related duodenal factors as a stimulator of pepsin secretion.
Glucagon and pepsin secretion

In conclusion, the study has shown that the potency of pentagastrin for pepsin secretion seems to be considerably higher than for acid secretion, and that glucagon reduces this potency without changing the maximal response, which contrasts with the effect on pentagastrin-stimulated acid secretion.

References

1 Christiansen J, Holst JJ, Kalaja E. Inhibition of gastric acid secretion in man by exogenous and endogenous pancreatic glucagon. Gastroenterology 1965; 70: 688–92.