Familial occurrence of complement dysfunction in Crohn's disease: correlation with intestinal symptoms and hypercatabolism of complement

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Summary

Complement was studied in Crohn's disease probands with early onset and in their first degree relatives. Controls included 24 healthy volunteers and 24 patients with ulcerative colitis or peptic ulcers. Subnormal generation of chemotactic activity by the alternative pathway was shown in eight of 21 probands and in six of 33 relatives, a frequency in both groups significantly different from controls (p<0.005), with a strong connection between findings in patients and relatives. As previously shown in patients with Crohn's disease, the subnormal generation was related to decreased utilisation of complement C3 in relatives. Raised levels of circulating complement C3c split products suggested complement involvement in Crohn's disease probands. In contrast, plasma C3c was normal in all relatives, and none of the six cases with complement dysfunction had gastrointestinal symptoms or a history of inflammatory bowel disease. Our data suggest, that complement abnormality seen in Crohn's disease patients does not simply reflect mucosal inflammation or hypercatabolism of complement.

A significant abnormality of complement has previously been shown in Crohn's disease. Release of chemotactic activity for neutrophils,1 2 elicited via C5a' from complement, is subnormal, the defective function being confined to the alternative pathway.3 Increases of complement C3 catabolism in vivo,4 raised levels of circulating complement C3c split products5 and positive immunoconglutinin titres6 all suggest involvement of complement in Crohn's disease.

In chronic granulomatous disease the primary dysfunction of phagocytic cells produces histologic changes of the gut, closely resembling those seen in Crohn's disease.7 This similarity stresses the potential significance of an inappropriate capacity for complement activation of neutrophils in Crohn's disease.

The aim of the present study was to define the nature of complement abnormality in Crohn's disease by a family study. Familial aggregation of chronic inflammatory bowel disease has been shown in several investigations8 and seems to be associated with early onset of the disease.9 10

Methods

Patients

A total of 21 patients with Crohn's disease, with an onset before the age of 21 years, were chosen as probands from our regional outpatient clinic. First degree relatives, aged 15–60 years were invited to participate.

The study was joined by all of 33 such family members of 16 probands living in the regional area. Five patients had no close relatives or they refused to accept family members' participation in the study.

Normal controls comprised 24 healthy volunteers, nine men and 15 women, aged 23–63 years. Disease controls included 12 consecutive outpatients with ulcerative colitis and 12 with peptic ulcers. The clinical data are given in Table 1. None of the subjects investigated had complicating infections, rheumatic disease, or conditions affecting the immune system such as atopy.

In a preliminary part of the study, optimal experimental conditions were defined through investigations of 12 Crohn's disease patients (Table 1) and 12 healthy volunteers.

All patients, relatives, and normal controls gave informed consent to participate. Our protocol has been accepted by the regional scientific-ethical
committee.

Circulating complement was activated by the alternative pathway with subsequent assessment of both chemotactic activity generated and C3 utilisation. Concentrations of C3c split products were assessed in non-activated plasma.

**COMPLEMENT ACTIVATION**

Freshly prepared heparin plasma (2 IU/ml) and serum were incubated with zymosan A (Sigma, St Louis, USA) at 37°C to activate alternative pathway complement.14 Non-activated samples served as negative controls. Further, spontaneous activation of complement was prevented with disodium EDTA (10 mM) added immediately after incubation. Zymosan particles were removed by centrifugation. All samples were frozen in liquid nitrogen, stored at -80°C, and thawed only once before analysis.

**CHEMOTACTIC ACTIVITY**

Generation of chemotactic activity by complement was quantified by the Boyden filter assay15 and the leading front technique, which has previously been described in detail.16 Leucocyte donors were healthy volunteers of blood type O Rh-negative. Test neutrophils were isolated from peripheral blood drawn in disodium EDTA (10 mM) by a methylcellulose sedimentation of the red cells, gradient centrifugation ofuffy coat leucocytes on Lymphoprep® (Nygaard, Oslo, Norway), and three times washing and resuspension in Gey's solution, including purified human-albumin (20 g/l, Behringwerke, Marburg, FR Germany). The final leucocyte preparation contained ≥97% polymorphonuclear leucocytes, and the median recovery of neutrophils was 45%. Two million cells from this suspension were added to the upper compartment of the modified Boyden chambers and neutrophil chemotaxis was assessed after migration for 45 minutes at 37°C in 3 μm pore size Sarstorius filters. Ten per cent dilutions of the serum and plasma samples in Gey's solution with disodium EDTA (10 mM) were added to the lower compartment of the Boyden chambers for assessment of chemotactic activity.

Controls included neutrophil migration towards casein (5 g/l, Alkalilloslich, Merck, Darmstadt, FRG) and Gey's solution with disodium EDTA (10 mM). The casein was dissolved in modified Gey's solution for 60 minutes at pH 12 with subsequent lowering of pH to 7.3.15 Samples from Crohn's disease patients with an early onset, their relatives and both groups of controls were tested in each of 21 experiments.

In a preliminary series of experiments 12 Crohn's disease patients and 12 healthy volunteers were studied to define optimal conditions for the activation procedure. Results represent the median of five determinations on each of two filters. Coefficient of variation for the double determinations was 4%.

**C3 UTILISATION AND C3C LEVELS**

Corresponding pairs of complement activated and non-activated plasma and serum samples were analysed by crossed immunoelectrophoresis on one immuno-plate. Complete separation of native C3 and its conversion products was achieved by the first run at 20 V/cm for 1.5 hours.18 The second
electrophoretic dimension was performed in antibody to C3c (Dakopatts, Copenhagen, Denmark) at 3 V/cm for 18 hours. Utilisation of C3 during activation was expressed as percentual decrease of the native C3 precipitation area.

Native C3 and the c split product of C3 (C3c) were quantified in plasma samples drawn in disodium EDTA, immediately frozen in liquid nitrogen, stored at -80°C, and thawed only once, before analysis. Native C3 was assessed by immunoelectrophoresis19 in antibody to C3c (Dakopatts) whereas C3c split products were analysed by an intermediate gel technique.20 Precipitation of native C3 and C3 split products with d specificity was achieved by antibody specific for C3d (Dakopatts) in the intermediate gel. C3c rockets were formed in the upper gel by precipitation with antibody specific for C3c. A plasma pool served as standard for quantitation of native C3 whereas standard human serum from Behringwerke, containing C3 in the form of C3c only, served as standard in the intermediate gel technique.

**Statistics**
The result were evaluated by non-parametric statistics: the Mann-Whitney U test, Wilcoxon's test for paired data, the four-fold table test, and Spearman rank correlation test.

**Results**

Eight of 21 Crohn's disease probands, and six of their 33 first degree relatives showed subnormal generation of chemotactic activity by the alternative pathway of circulating complement (Fig. 1). Significance was at the p<0.005 level, both comparing probands (χ²=17, df=1) and comparing relatives (χ²=7, df=1) with total control group. The chemotactic activity of non-activated samples did not differ between probands, relatives and controls (Table 2). Identical results were obtained with plasma and serum complement in this study (Table 3). Means of results obtained with the two sources of complement are given in Fig. 1.

The six Crohn's disease relatives with complement dysfunction had a decreased utilisation of C3 by the alternative pathway of complement (Fig. 2), compared with normal and disease controls, and with 27 Crohn's disease relatives showing normal generation of chemotactic activity (p<0.005). Means of identical results from plasma and serum are given. Normal levels of circulating, native C3 in Crohn's disease relatives (Table 4) permitted the C3 utilisation to be expressed in percent of C3 concentrations.

Levels of plasma C3c split products were normal in Crohn's disease relatives and in disease controls with ulcerative colitis and peptic ulcers (Fig. 3) whereas an increase was observed in Crohn's disease patients, versus both normal and disease controls (p<0.005). Neither C3c levels nor chemotactic

**Table 2** Chemotactic activity in μm/45 minutes of non-activated plasma and serum samples. Medians with ranges in parentheses.

<table>
<thead>
<tr>
<th></th>
<th>Normal controls</th>
<th>Crohn's disease probands</th>
<th>Crohn's disease relatives</th>
<th>Disease controls</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(n=24)</td>
<td>(n=21)</td>
<td>(n=33)</td>
<td>(n=24)</td>
</tr>
<tr>
<td>Non-activated plasma complement</td>
<td>34 (16-56)</td>
<td>26 (6-60)</td>
<td>30 (8-54)</td>
<td>31 (10-58)</td>
</tr>
<tr>
<td>Non-activated serum complement</td>
<td>37 (18-85)</td>
<td>30 (6-95)</td>
<td>36 (6-98)</td>
<td>33 (9-84)</td>
</tr>
</tbody>
</table>

Fig. 1 Generation of chemotactic activity by the alternative pathway of complement in Crohn's disease patients (CD), their first degree relatives (R), normal controls (N), and disease controls with ulcerative colitis (UC) or peptic ulcers (PU). • experimental groups, ○ controls.
Table 3  Generation of chemotactic activity in μm/45 minutes by the alternative pathway of plasma and serum complement. Medians with ranges in parentheses.

<table>
<thead>
<tr>
<th></th>
<th>Normal controls (n=24)</th>
<th>Crohn's disease patients (n=21)</th>
<th>Crohn's disease relatives (n=33)</th>
<th>Disease controls (n=24)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Activated plasma comple</td>
<td>100</td>
<td>68*</td>
<td>86</td>
<td>106</td>
</tr>
<tr>
<td>ment</td>
<td>(72-143)</td>
<td>(20-106)</td>
<td>(19-140)</td>
<td>(70-146)</td>
</tr>
<tr>
<td>Activated serum comple</td>
<td>101</td>
<td>74*</td>
<td>94</td>
<td>110</td>
</tr>
<tr>
<td>ment</td>
<td>(70-146)</td>
<td>(33-111)</td>
<td>(10-155)</td>
<td>(73-150)</td>
</tr>
</tbody>
</table>

* p<0.005.

activity generated by complement related to disease activity (p<0.10, rank correlation test) or to localisation of inflammation (p<0.10, rank sum test).

Complement abnormality in Crohn's disease relatives was confined to families of probands with a release of chemotactic activity by complement below or at the lower border of the reference interval (Table 5). The relatives with dysfunction had no symptoms or history of inflammatory bowel disease. The phenomenon occurred both in parents and siblings, in young and middle aged, and in men and women (Table 5). One Crohn's disease relative with complement dysfunction had a previously diagnosed ankylosing spondylitis (Table 5).

In a preliminary part of the investigation the dose response and the time response for generation of chemotactic factors by complement was investigated (Table 6). The optimal sensitivity in the Boyden filter assay of chemotactic factor generation was obtained with zymosan 2×10⁻¹ g/l and incubation time of 60 minutes. These conditions were chosen for the present study. Declining responses with increases of incubation time and zymosan concentrations are consistent with a relation between decreasing responses and supramaximal concentrations of C5a. Test neutrophils had a median casein chemotaxis of 135, range 105–174 μm/45 minutes in all experiments, and spontaneous migration towards Gey's solution with EDTA was 37, range 5–65 μm/45 minutes.

Discussion

Subnormal generation of chemotactic activity by the alternative pathway of complement has previously been shown in unselected patients with Crohn's disease. This observation was confirmed in the

Fig. 2  Utilisation of complement C3 by the alternative pathway of complement in first degree relatives of Crohn's disease patients (R), normal controls (N), and disease controls with ulcerative colitis (UC) or peptic ulcers (PU).

Fig. 3  Plasma concentrations of C split products of complement C3 (C3c) in Crohn's disease patients (CD), their first degree relatives (R), normal controls (N), and disease controls with ulcerative colitis (UC) or peptic ulcers (PU). ● experimental groups, ○ controls. Values expressed in per cent of a standard human serum containing C3 in the form of C3c only.
Complement dysfunction in Crohn's disease

Table 4 Concentrations of native C3 in per cent, medians with ranges in parentheses.

<table>
<thead>
<tr>
<th>Normal controls (n=24)</th>
<th>Crohn's disease probands (n=21)</th>
<th>Crohn's disease relatives (n=33)</th>
<th>Disease controls (n=24)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma C3</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>96 (65-142)</td>
<td>101 (56-148)</td>
<td>107 (58-153)</td>
<td>93 (57-125)</td>
</tr>
</tbody>
</table>

The present group of probands with early onset Crohn's disease. A similar defect of complement was revealed in six of their 33 first degree relatives. All members of the subgroup of relatives with complement abnormality had a negative case history of intestinal symptoms and, in contrast with patients, no raised plasma C3c, excluding hypercatabolism of complement in Crohn's disease patients as the explanation.5

The unexpected finding of normal C3c split product levels in ulcerative colitis controls, contrasting C3c rise in Crohn's disease probands has previously been discussed.5

Abnormalities of complement function in chronic inflammatory conditions such as systemic lupus erythematosus are believed to reflect excessive consumption of complement by cascade reactions.22 The present results in relatives, indicating an increased risk for later development of overt inflammatory bowel disease, strongly suggest that the demonstrated complement dysfunction can be a central phenomenon, unrelated to mucosal inflammation or hypercatabolism of complement. It must, however, be pointed out that this argument is based essentially on just six abnormal findings among the relatives, although none were found among normal and disease controls. It would seem interesting to assess the incidence of this possible genetic predispositions to Crohn's disease in the normal population by studying a rather large sample. Such studies must await screening

Table 5 Families of Crohn probands ranked according to results of complement function test (release of chemotactic activity). Numbers 1 to 8: subnormal function; numbers 9 to 16: normal function.

<table>
<thead>
<tr>
<th>No</th>
<th>Patient's code</th>
<th>Relatives' code</th>
<th>Release of chemotactic activity</th>
<th>Intestinal symptoms/complicating conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>JO 23♂</td>
<td>P: 56♂ S: 28♂</td>
<td>N</td>
<td>Crohn's disease</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Diarrhoea episodes</td>
</tr>
<tr>
<td>2</td>
<td>AME 17♀</td>
<td>P: 53♂ S: 24♂</td>
<td>D</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>JHH 27♂</td>
<td>P: 50♂ S: 24♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>OFR 21♂</td>
<td>S: 23♂</td>
<td>D</td>
<td>Ankylosing spondylitis</td>
</tr>
<tr>
<td>5</td>
<td>PMB 28♀</td>
<td>P: 53♂ S: 23♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>MT 19♀</td>
<td>P: 48♂ S: 22♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>AW 35♂</td>
<td>S: 39♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>VE 20♀</td>
<td>S: 23♂</td>
<td>D</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>TW 13♂</td>
<td>P: 42♂ S: 38♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>GFJ 37♀</td>
<td>S: 25♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>11</td>
<td>MRH 28♀</td>
<td>P: 52♂ S: 25♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>LBH 28♀</td>
<td>S: 34♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>LSS 19♀</td>
<td>P: 39♂ S: 18♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>14</td>
<td>LSH 45♀</td>
<td>S: 29♂</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>15</td>
<td>LFN 13♀</td>
<td>P: 31♀</td>
<td>N</td>
<td></td>
</tr>
<tr>
<td>16</td>
<td>SNK 26♀</td>
<td>P: 55♂ S: 31♂</td>
<td>N</td>
<td></td>
</tr>
</tbody>
</table>

P = parents; S = siblings; N = normal; D = decreased.
procedures which are less time consuming than the combination of biologic and electrophoretic assays applied in the present paper. An immuno-electrophoretic method for C5a measurements may permit a valuable methodological alternative for large studies.

The possible pathogenetic significance of the complement abnormality remains to be established. Only careful clinical follow up will reveal if relatives considered at risk will ultimately develop overt disease. Complement dysfunction seems to be relatively specific for Crohn’s disease probands and relatives because the present patients with ulcerative colitis and peptic ulcers all showed normal generation of chemotactic activity by complement.

As previously shown for Crohn’s disease patients, complement abnormality in the present subgroup of first degree relatives was associated with a decreased consumption of complement C3 by the cascade reaction. Depressed conversion at both C3 and C5 levels during activation, with resulting subnormal release of the major, complement derived chemotactic factor C5a, is probably the underlying defect. The recent demonstration of increased cell directed inhibitors of chemotaxis and of chemotactic factor inactivators possibly contribute to a defective chemotactic factor generation but not to the abnormality at C3 level in Crohn’s disease.

The molecular basis of this deficient function of the alternative pathway remains unknown. Previous measurements of individual complement components involved and of the control proteins have been negative. Further studies trying to relate complement dysfunction with polymorphism of complement proteins and with established genetic markers such as the HLA-types are being done.

The subgroups of relatives with complement dysfunction comprised six persons only. Both parents/siblings and men/women were represented, but the group is too small for any statistical analysis. Complement abnormality in relatives was confined to families of patients who themselves showed similar changes. Thus, further screening of relatives could probably be confined to families of patients with complement dysfunction. Crohn’s disease patients with early onset may be especially predisposed to acquire the disease by genetic and/or environmental factors. Our figures for complement abnormality in close relatives consequently may not be representative for relatives of unselected patients with Crohn’s disease.

We wish to thank H Furhauge and H Kargaard for excellent technical assistance and L Nielsen for typing the manuscript. The study was supported by Johann and Hanne Weimann née Sedorffs foundation.

References
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