Synthesis of prostaglandin E₂, thromboxane B₂ and prostaglandin catabolism in gastritis and gastric ulcer

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SUMMARY Because endogenous prostaglandins may protect the gastric mucosa a study was conducted to determine factors influencing (a) the synthesis of immunoreactive prostaglandin (iPG) E₂ and thromboxane (iTx) B₂ as measured by radioimmunoassay and (b) prostaglandin catabolism measured radiometrically, in human gastric mucosa. Gastric mucosa was obtained at endoscopy. Synthesis of iPE₂ and iTxB₂ was inhibited in vitro by indomethacin; iTxB₂ synthesis was also selectively inhibited by the thromboxane synthesis inhibitor dazmegrel. Prostaglandin catabolism was inhibited by carbenoxolone. Multivariate analysis showed that synthesis of iPGE₂ from endogenous precursor during homogenisation was decreased in patients on non-steroidal anti-inflammatory drugs. Mucosal inflammation was associated with significantly increased synthesis of iPGE₂ and decreased prostaglandin catabolism. There were no differences between the mucosa of patients with or without gastric ulcers, nor between the ulcer rim and mucosa 5 cm away. Age, sex, smoking history, and ingestion of antisecretory drugs appeared to exert no influence. In this study gastritis was the major influence on prostaglandin synthesis. It seems unlikely that prostaglandin deficiency is a strong predisposing factor for gastric ulceration.

Reports suggesting gastric mucosal protection by endogenous prostaglandin synthesis¹ ³ raise the possibility that this process might be defective in patients who develop gastric ulcers. Previous evidence on this point has been conflicting. Some studies have found reduced prostaglandin synthesis in gastric ulcer patients,³ ⁵ even in the presence of gastritis: this would be surprising because other inflammatory lesions lead to enhanced prostaglandin synthesis. Another study reported increased prostaglandin synthesis in patients with gastric ulcers⁶ and attributed this to the associated gastritis; however, even where this occurs it remains possible that prostaglandin synthesis could be deficient in relation to the prevailing levels of mucosal inflammation.

Factors other than reduced prostaglandin synthesis might render the mucosa liable to ulcerate. These could include an absolute or relative enhance-ment of thromboxane synthesis as thromboxane synthesis is associated with gastric ulceration in rats and inhibitors of thromboxane synthesis appear to be protective.⁷ ¹¹ Alternatively reduced levels of protective prostaglandins might result from increased prostaglandin catabolism.¹² ¹³

In this study synthesis of immunoreactive prostaglandin (iPG)E₂ and of thromboxane (iTx)B₂, and prostaglandin catabolism in human gastric mucosa have been investigated with reference both to histological appearances and to the presence or absence of gastric ulceration. The relationship of age, sex or smoking or ingestion of non-steroidal anti-inflammatory drugs (NSAIDS) or antisecretory drugs to these parameters has also been investigated in view of the known influence of these factors on the incidence or natural history of gastric ulceration.

Methods

PATIENTS
The following radiolabelled compounds were used: (H₃)PGE₂, 160 Ci/mmol (Amersham International):
Human gastric prostanoids

(\(H_2\)) TxB\(_2\), 139-3 Ci/mmol (New England Nuclear) and (9 \(H_2\))PGF\(_2\), 13-7 Ci/mmol (Amersham International). The following compounds were gifts: indomethacin (Merck Sharp & Dohme), dazmegrel (Pfizer UK), carbenoxolone (Biorex Laboratories). TxB\(_2\) antiserum was kindly donated by Dr Lawrence Levine. PGE\(_2\) antiserum was bought from the Institut Pasteur. Other chemicals were obtained from standard laboratory suppliers.

Patients undergoing upper gastrointestinal endoscopy with an Olympus T endoscope were studied. Where there was a possibility that mucosa might be taken for research patients were asked to give written informed consent before the endoscopy. The study was approved by the South Nottingham Ethical Committee.

In studies used to develop and validate the incubation conditions and radioimmunoassays and to assess reproducibility with repeated study, both normal subjects and patients with a variety of gastroduodenal conditions were studied. During the main study 27 patients with gastric ulcers and 43 control patients were studied. As gastritis was one of the factors which the study was designed to investigate, endoscopic or histological evidence of inflammation was not an exclusion criterion. In all other respects, the control patients had normal endoscopic findings. The characteristics of the patients studied are detailed in Table 1.

**HISTOLOGY**

Four to five mucosal biopsy specimens were taken from the lesser curve between 45 and 50 cm from the incisor teeth and used immediately. A small piece from two of the specimens, or a separate biopsy specimen from the same area was examined histologically (without knowledge of the biochemical results) and graded as uninflamed (normal or only slight increase in inflammatory cell infiltrate) or inflamed (moderate or marked increase in inflammatory cell infiltrate) using standard histological criteria of inflammation.\(^1\)\(^2\) No attempt was made to differentiate between inflammation caused mainly by neutrophils and inflammation caused mainly by lymphocytes. As mucosal atrophy and mucosal inflammation were frequently associated no attempt was made to assess them independently.

**HOMOGENATES AND INCUBATIONS**

The rest of the biopsy specimens were homogenised immediately on ice in Tris HCl 0.05M, pH 7.4 (20 mg wet weight/ml, final proportions) for 15 seconds using an Ulfraturax homogenizer.

**SYNTHESIS FROM ENDOGENOUS STORES**

(ENDOGENOUS iPGE\(_2\) AND iTxB\(_2\))

Ten milligram aliquots in 0.5 ml (final proportions) of the homogenate were extracted immediately into chloroform,\(^13\) for the determination of prostanoids synthesised from endogenous stores (henceforth designated Endogenous iPGE\(_2\) or iTxB\(_2\)).

**INHIBITOR EXPERIMENTS**

Some samples were pre-incubated in Tris HCl containing either indomethacin 50 \(\mu\)g/ml (4 \times 10\(^{-4}\) M) for 15 minutes before being homogenised in fresh Tris HCl also containing indomethacin 50 \(\mu\)g/ml (4 \times 10\(^{-4}\) M). These samples were extracted in the usual way and results compared with those obtained from biopsies from the same patient but not exposed to indomethacin.

**SYNTHESIS AFTER INCUBATION WITH ARACHIDONIC ACID (TOTAL iPGE\(_2\) SYNTHESIS)**

Other aliquots (10 mg, 0.5 ml, final proportions) were incubated with arachidonic acid 2 \(\mu\)g for 30 minutes at 37°C in a rocking water bath before extraction into chloroform.\(^15\) Because this procedure results in synthesis of prostanoids from both endogenous and exogenous arachidonic acid it is designated Total iPGE\(_2\) synthesis.

**INHIBITOR EXPERIMENTS**

In some experiments aliquots of homogenates were incubated with or without indomethacin 1 \(\mu\)g/ml (2.8 \times 10\(^{-7}\)M) or dazmegrel 0.3 \(\mu\)g/ml (10\(^{-6}\)M). The effect of the inhibitor was calculated after allowance for synthesis during homogenisation.

**RADIOIMMUNOASSAYS**

After extraction into chloroform, samples were resuspended in phosphate buffered saline (pH 7.4, 0.15 M), and stored at −50°C for radioimmunoassay, usually within one month (maximum three

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**Table 1 Characteristics of the patients studied**

<table>
<thead>
<tr>
<th>Characteristics of the patients studied</th>
<th>Control</th>
<th>GU patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td>43</td>
<td>27</td>
</tr>
<tr>
<td>Men</td>
<td>29</td>
<td>13</td>
</tr>
<tr>
<td>Smokers</td>
<td>16</td>
<td>17</td>
</tr>
<tr>
<td>Antisecretory drugs</td>
<td>9</td>
<td>17</td>
</tr>
<tr>
<td>NSAIDS(^*)</td>
<td>5</td>
<td>4</td>
</tr>
<tr>
<td>Age &lt;30 years</td>
<td>7</td>
<td>—</td>
</tr>
<tr>
<td>30-50</td>
<td>11</td>
<td>—</td>
</tr>
<tr>
<td>50-65</td>
<td>9</td>
<td>11</td>
</tr>
<tr>
<td>65-75</td>
<td>11</td>
<td>14</td>
</tr>
<tr>
<td>75+</td>
<td>4</td>
<td>2</td>
</tr>
</tbody>
</table>

\(^*\)Patients on regular NSAIDS's, last dose in each case approximately 12 hours before endoscopy; individual drugs as follows: indomethacin: five, aspirin: two, naproxen: one, flurbiprofen: one.
months). iTxB2 was measured by radioimmunoassay using anti-serum kindly donated by Dr Lawrence Levine. For thromboxane assays standard TxB2, TxB2 anti-serum and [\(^{3}H\)] TxB2 tracer (Amersham) were dissolved and samples were diluted in Tris saline pH 7.4, 0.15 M with 1% gelatin. iPGE2 was measured by radioimmunoassay as previously described.16 17

CATABOLISM
Aliquots of homogenate (10 mg 0.5 ml, final proportions) were also incubated with (9\(\beta\)-H\(^3\))PGF\(_{2\alpha}\) (1 mg, 0.1 \(\mu\)Ci) and NAD 1 mm for five minutes at 37\(^\circ\)C. They were extracted into chloroform at pH 3.5 and re-suspended in 75 \(\mu\)l chloroform:ethanol (1:1). PGF\(_{2\alpha}\), and its metabolites were separated by thin layer chromatography using Watman LK6D silica gel chromatography plates and ethyl acetate: acetone:acetic acid (90:10:1) as solvents. Reagents corresponding to authentic PGF\(_{2\alpha}\), 15 keto PGF\(_{2\alpha}\), 13-14 dihydro PGF\(_{2\alpha}\), and 13-14 dihydro 15 keto PGF\(_{2\alpha}\) were scraped and added direct to LKB Optiphase scintillant for quantitation by liquid scintillation counting. Radioactivity cochromatographing with metabolites was expressed as a percent of total radioactivity scraped. A similar value derived using boiled mucosa from the same patient was then deducted to obtain values for the percentage of enzymatic catabolism in samples.

INHIBITOR EXPERIMENTS
In some experiments, aliquots of the homogenate were preincubated on ice for 15 minutes with carbenoxolone 57 \(\mu\)g/ml (10\(^{-4}\) M) or 5.7 \(\mu\)g/ml 10\(^{-5}\) M) and then incubated with (9\(\beta\)-H\(^3\))PGF\(_{2\alpha}\) (1 \(\mu\)g, 25 nCi) and NAD 1 mm for 15 minutes at 37\(^\circ\)C.

STATISTICAL METHODS
The catabolism data approximated to a normal distribution but all synthesis data were transformed logarithmically to obtain a normal distribution for analysis. The influence of gastritis, gastric ulceration, age, sex, smoking, drug ingestion, and study number (to allow for any changes in experimental technique with time) upon prostaglandin synthesis and catabolism were investigated by multivariate analysis (Statistical Package for Social Sciences Programme). Paired data were evaluated using the paired \(t\) test.

Results
VALIDITY OF RADIOIMMUNOASSAYS
(i) iPGE2
In these experiments assay of samples at two-fold dilutions gave similar results (value at higher dilution 99+3%, SEM, of value at lower dilution, \(n=6\)).

(ii) iTxB2
Under the assay conditions used, the cross reactivity of the TxB2 antiserum with other prostanooids (based on concentrations required to produce 50% displacement of (H\(_1\))TxB2) was as follows: PGD\(_2\): 1.2%; PGF\(_{1\alpha}\): 0.12%; PGE\(_2\): 0.1%; PGF\(_{2\alpha}\): 0.01%; arachidonic acid, PGA\(_2\), 6 keto PGF\(_{1\alpha}\), and 13-14 dihydro 15 keto PGE\(_{2}\): all <0.001%. Assay of samples at two dilutions gave similar results (value at higher dilution=105+5% of value at lower dilution, \(n=15\)). Authentic TxB2 added to samples before or after extraction was assayed as 90+9% (\(n=20\)) of that added. TxB2 re-assayed after one to three months of storage was 109+1% (\(n=53\)) of initial value. TxB2 anti-serum showed selective reactivity with material cochromatographing with authentic PGF\(_{2\alpha}\).

QUALITY CONTROL
Preliminary experiments showed that iPGE2 and iTxB2 (measured by radioimmunoassay) were recovered to a similar extent (69±1% for PGF\(_{2\alpha}\) and 69±2% for TxB2) (\(n=3\) each). In subsequent experiments (H\(_1\))iPGE2 was used to measure recovery: mean recoveries were 72%. Three external standards of PGF\(_{2\alpha}\) or of TxB2 were included in each radioimmunoassay and two samples previously assayed satisfactorily were re-assayed to ensure accuracy and consistency from one assay to another.

HOMOGENATES
EFFECTS OF INHIBITORS IN VITRO
(a) Immediate extraction (Endogenous iPGE\(_2\) and TxB\(_{2}\))
Synthesis of iPGE2 was inhibited by 60±8% (\(n=8\)) if homogenisation was carried out in the presence of indomethacin, 50 \(\mu\)g/ml (1.4×10\(^{-4}\) M). Under these conditions synthesis of iTxB2 was reduced by 68±6% (\(n=8\)).

(b) Incubated synthesis
Synthesis of iPGE2 was inhibited by 72±9% and of iTxB2 by 63±15% (\(n=4\)) by indomethacin 1 \(\mu\)g/ml (2.8×10\(^{-5}\) M). The thromboxane synthesis inhibitor dazmegrel 0.3 \(\mu\)g/ml (10\(^{-6}\) M) inhibited iTxB2 synthesis by 44±21%; synthesis of iPGE2 was 243±69% control (\(n=4\)).
(c) Catabolism
Carbenoxolone inhibited prostaglandin catabolism by 29±9% at 10⁻⁵ M (n=8) and by 79±5% at 10⁻⁴ M (n=6).

REPRODUCIBILITY
Eighteen patients were studied on two occasions after receiving in the intervening period, misoprostol (n=7), trimoprostil (n=4), placebo (n=4), cimetidine (n=2) and ranitidine (n=1). Two way analysis of variance showed that patients differed significantly from each other but that there was no significant change after the period of treatment. Based on the residual variance, the coefficients of variation for repeated study were: for endogenous iPGE₂: 14%; for total iPGE₂: 10%; for iTxB₂: 17% (all logarithmically transformed data) and for prostaglandin catabolism (untransformed data): 33%.

INFLUENCE OF PATIENT CHARACTERISTICS ON SYNTHESIS AND CATABOLISM
Table 2 shows all factors having a statistically significant or near significant influence on the dependent variables studied. Where patients were studied more than once the first data only were used. Inflammation was a major influence and was associated with increased synthesis of iPGE₂ (both endogenous and total) as well as reduced prostaglandin catabolism (Fig. 1). Patients taking non-steroidal anti-inflammatory drugs showed reduced synthesis of prostanoids (Fig. 1): this was statistically significant for endogenous iPGE₂. The study number was a significant influence on prostaglandin catabolism. Total iPGE₂ and iTxB₂: values of all three increased gradually as the study progressed. Finally there were trends towards lower levels of iPGE₂ and of iTxB₂ in patients with gastric ulceration but these did not reach conventional levels of significance. No other factor showed statistically significant influence.

**Table 2 Factors influencing the synthesis and catabolism of prostanoids in human gastric mucosa**

<table>
<thead>
<tr>
<th>Dependent variable</th>
<th>n</th>
<th>Influence</th>
<th>R</th>
<th>F</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ln(Endogenous PGE₂)</td>
<td>64</td>
<td>NSAIDS</td>
<td>-0.31</td>
<td>8.30</td>
<td>&lt;0.01</td>
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<tr>
<td></td>
<td></td>
<td>Inflammation</td>
<td>0.28</td>
<td>8.39</td>
<td>&lt;0.01</td>
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<tr>
<td></td>
<td></td>
<td>GU</td>
<td>-0.15</td>
<td>3.73*</td>
<td>&lt;0.1*</td>
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<tr>
<td>Ln(Total PGE₂)</td>
<td>60</td>
<td>Inflammation</td>
<td>0.37</td>
<td>9.81</td>
<td>&lt;0.01</td>
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<tr>
<td></td>
<td></td>
<td>Study number</td>
<td>0.32</td>
<td>7.015</td>
<td>&lt;0.05</td>
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<tr>
<td>Ln(TXB₂)</td>
<td>45</td>
<td>Study number</td>
<td>0.42</td>
<td>8.39</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td></td>
<td></td>
<td>GU</td>
<td>-0.28</td>
<td>3.85*</td>
<td>&lt;0.1*</td>
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<tr>
<td>Catabolism</td>
<td>70</td>
<td>Study number</td>
<td>0.32</td>
<td>8.68</td>
<td>&lt;0.01</td>
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<tr>
<td></td>
<td></td>
<td>Inflammation</td>
<td>-0.23</td>
<td>5.10</td>
<td>&lt;0.05</td>
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</table>

*Ln=natural logarithm

Discussion

The main conclusions from these data are that iPGE₂ synthesis as measured in the study is increased and that prostaglandin catabolism is reduced in the presence of gastritis. After allowing for the influence of coexisting inflammation, however, there were no significant differences between gastric mucosa from patients with or without gastric ulceration.

Radioimmunoassays were used as the bases of measurement for synthesis. In these studies as with...
Human gastric prostanoids

Endogenous iPGE2  Total iPGE2  TXB2

Synthesis (pg/mg)

0  100  1000

Lesser curve  Rim

Catabolism (%)

100  80  60  40  20

Lesser curve  Rim

Fig. 2  Synthesis and catabolism of prostanoids in gastric ulcer patients: comparison of mucosa away from the ulcer (lesser curve) and from the ulcer rim. Results from lesser curve and from the ulcer rim are joined for each individual patient — show means. Stippled areas show SEM's.

Endogenous iPGE2  Total iPGE2  TXB2

Synthesis (ng/mg)

0  100  1000

No GU  GU  No GU  GU  No GU

Catabolism (%)

100  80  60  40  20

No GU  GU

Fig. 3  Synthesis and catabolism of prostanoids in gastric mucosa: comparison of gastric ulcer patients with matched controls. Each pair of connected points shows results from an individual gastric ulcer patient joined to those obtained from the matched control. — show means. Stippled areas show SEM's.

others using radioimmunoassay it is possible that crossreacting substances were at least in part measured. This is acknowledged by use of the terms immunoreactive PGE2 or TXB2 although it seems unlikely that another substance would cross react with the antiserum, cochromograph with the prostanoid and that its synthesis would be inhibited in an appropriate way by indomethacin or dazmegrel. The rather modest reduction (60–70%) in immunoreactive prostanoids synthesised during homogenisation achieved by high concentration of indomethacin could be taken as evidence that substances other
than cyclo-oxygenase products were measured. Greater inhibition was seen, however, with lower concentration of indomethacin in the incubated homogenates and other explanations for the results obtained during homogenisation are more likely: there may have been incomplete diffusion of indomethacin and consequently only partial cyclo-oxygenase inhibition so that some synthesis occurred during homogenisation. The present results are also consistent with earlier data and the suggestion accompanying them that in gastric mucosa there may in fact be a small resting content of prostanoids, though methodological difficulties make it impossible to resolve this point with certainty.

Prostaglandin catabolism was measured radiometrically. Prostaglandin F$_{2\alpha}$ was used as substrate because of its resistance to non-enzymatic catabolism. The assay is of enzyme activity in a defined system and should not be taken to imply that PGF$_{2\alpha}$ is normally the main prostaglandin catabolised by this pathway. Inhibition of prostaglandin catabolism by carbenoxolone has been reported before. In the present study different methodology has been used to confirm that carbenoxolone inhibits prostaglandin catabolism with a potency similar to that previously reported.

The enhanced synthesis of iPGE$_2$ with gastritis presumably reflects the infiltration of the mucosa by inflammatory cells: human neutrophils and macrophages both have the capacity to synthesise PGE$_2$. These differences between inflamed and uninflamed mucosa are similar to those shown by Schlegel et al. Doubt has been expressed as to the validity of these earlier data as the amounts of PGE$_2$ synthesised appeared to be very high. In the present study the quantities measured were much lower but the pattern of results was the same. The reduction in prostaglandin catabolism in inflamed tissue has not been shown before. Two main explanations are possible. One is that a primary defect in prostaglandin catabolism leads to increased mucosal levels of pro-inflammatory prostaglandins and makes the mucosa more liable to become inflamed. An alternative explanation is that the reduced prostaglandin catabolism is a consequence rather than a cause of the inflammation: gastric mucosa has an exceptionally high capacity for prostaglandin catabolism which might fall because of the mucosal atrophy which accompanies the inflammation in gastritis.

Whatever the mechanism, increased synthesis and reduced catabolism of prostaglandins would both lead to increased tissue levels in gastritis. As with ulcerative colitis, the significance of increased prostaglandin synthesis in gastritis must be viewed in the context of the known properties of prostaglandins. In both situations, it is difficult to ascribe a central role to prostaglandins because they do not have properties which make them likely to cause recruitment of inflammatory cells or to lead to mucosal atrophy. Vasodilator prostaglandins may, however, give rise to mucosal redness sometimes seen in association with acute superficial gastritis. The mediators responsible for the recruitment of inflammatory cells in gastritis however remain unidentified. Obvious candidates which require investigation are leukotriene B$_4$ and other related chemotactic lipoxygenase products.

Although there were large differences between patients the data show reasonable within patient consistency on repeated study, despite intervening treatment with a variety of therapeutic agents. The purpose of the reproducibility study was to validate the methodology rather than investigate the effects of individual drugs and the numbers are too small to exclude the possibility that some of the treatments influenced subsequent prostanoid synthesis. Antisecretory drug treatment, however, was not a significant influence on any of the variable investigated in the main study in contrast with the earlier suggestion that cimetidine enhances gastric mucosal prostaglandin synthesis.

The present results are different from those obtained by Wright et al both in showing enhanced synthesis with gastritis and in failing to find significant differences in patients with gastric ulceration. Trends toward lower levels of endogenous iPGE$_2$ and of iTxB$_2$ in gastric ulcer patients do not reach statistical significance and no difference was seen in the comparison of matched pairs of patients. Moreover, there were no local changes in PGE$_2$ or TxB$_2$ synthesis or prostaglandin catabolism in the ulcer rim. Thus in this study gastric ulceration seems at best to be a much weaker influence than suggested by Wright et al who reported reduced levels of PGE$_2$ in patients with gastric ulcers even when the mucosa was inflamed. There was no evidence of an imbalance between PGE$_2$ and TxB$_2$.

The reasons for the differences between the present data and those of Schlegel et al on the one hand and those reported by Wright et al on the other are not clear. The use of homogenates might be insensitive to subtle changes in cyclooxygenase activity and it remains possible that changes in prostanoid synthesis could occur in vivo because of phospholipase activation, the influence of neural stimulation or changes in endogenous synthesis. Specific evidence for such changes in gastric ulcer patients is lacking, however, and these possibilities could not account for the differences between the three studies as all used homogenates. The controls in the present study were 43 patients in whom no
Human gastric prostansoids

Pathology was found whereas Wright et al used seven normal volunteers. One point that emerged from the use of the patient study number as a quality control measure was that there was a tendency for all parameters measured to rise slightly in value during the course of the study (presumably because of subtle changes in methodology). Because the gastric ulcer patients and their controls were studied concurrently these changes do not influence the comparison between them. Comparable data are not provided in the other studies.

One stimulus for this study was the observation that the incidence of gastric and duodenal ulceration is static or rising in older women in the United Kingdom but falling in other groups.21, 24 Although a balanced group of patients with a wide age distribution was studied there was no evidence that the parameters measured were influenced by age, sex, or smoking. Patients on NSAID’s, however, showed reduced prostaglandin synthesis even though they were studied more than 12 hours after their last dose. Comparable differences between patients not on NSAID’s and controls were not seen suggesting that reduced prostaglandin dependent processes are less important in this group than in patients taking NSAID’s. Wright et al regarded the reduction in synthesis of PGE2 which they observed with gastric ulcer to be a secondary phenomenon. The data reported in the present paper give even fewer grounds for believing that a primary deficiency of prostaglandin synthesis accounts for the development of gastric ulceration in patients not on NSAID’s and thus cast some doubt on the clinical relevance of using prostaglandins for mucosal protection of these patients.

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