

Histocompatibility antigens in patients with alcoholic liver disease in Scotland and northeastern England: failure to show an association

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SUMMARY A study of HLA-A and B antigens in 248 patients with biopsy diagnosed alcoholic liver disease was conducted to examine for a genetic predisposition to alcohol related liver injury. No statistically significant differences were established for 8 HLA-A and 16 HLA-B antigens between normal healthy controls (n=342) and patients with alcoholic fatty liver (n=86), alcoholic hepatitis (n=63), active alcoholic cirrhosis (n=64) and inactive alcoholic cirrhosis (n=35). It is concluded that no HLA-A or B locus genetic susceptibility to alcoholic related injury could be shown.

Epidemiological surveys have indicated that the risk of developing alcoholic cirrhosis is related both to the amount and duration of alcohol abuse.¹ On the basis of numerous histological surveys of liver disease among chronic alcoholics, however, it is estimated that only 30% of heavy drinkers will develop alcoholic hepatitis and 10-15% cirrhosis.²⁻⁴ Clearly, other constitutional or environmental factors must, therefore, play a role in determining individual susceptibility to alcoholic liver disease. Genetic influences have been explored by studying major histocompatibility antigens in patients with established alcoholic liver disease. Several small studies were reported in the late 1970s showing isolated and conflicting associations of HLA-A and B antigens with alcoholic liver disease. An extensive review of the literature in 1982 concluded that no single HLA antigen was associated with a predisposition to alcoholic liver disease and that most of the studies reported on rather small numbers of patients with variable criteria for selection.⁵ Simultaneously, in 1982 a large single centre study of 170 British chronic alcoholics with and without liver disease failed to show any relationship between HLA antigens and susceptibility to alcoholic cirrhosis.⁶

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The controversy was reawakened in 1986 by the publication of two studies; one from France which showed an increased prevalence of HLA-B15 and reduced prevalence of B13 in patients with alcoholic cirrhosis⁷ and one from the USA which showed an increased prevalence of B8 in whites with moderately severe alcoholic hepatitis.⁸

Therefore, we have been stimulated to report our own findings. As part of an extensive epidemiological study of 510 patients with histologically proven alcoholic liver disease in Scotland and northern England,⁹ class I histocompatibility antigens were typed in a randomly selected cohort of 327 patients.

Methods

PATIENTS

Patients were accessed at seven centres in Scotland and one in Newcastle as part of a collaborative study of the Caledonian Society of Gastroenterology.⁹ All patients were British, caucasian, had a history of regular alcohol consumption in excess of 60 g daily and histological evidence of alcoholic liver disease. Patients were classified histologically as showing fatty liver, alcoholic hepatitis, active alcoholic cirrhosis (together with alcoholic hepatitis) or inactive alcoholic cirrhosis.

All samples for HLA typing were transferred to

Table HLA-A and HLA-B antigens in patients with alcoholic liver disease and controls

	Controls		Alcoholic fatty liver		Alcoholic hepatitis		Active alcoholic cirrhosis		Inactive alcoholic cirrhosis	
	Total 342 (n)	Freq (%)	Total 86 (n)	Freq (%)	Total 63 (n)	Freq (%)	Total 64 (n)	Freq (%)	Total 35 (n)	Freq (%)
A1	138	40.3	35	40.7	26	41.3	22	34.4	16	45.7
A2	163	47.6	38	44.2	35	55.5	35	54.7	16	45.7
A3	78	22.8	22	25.6	15	23.8	19	29.7	3	8.6
A9	60	17.5	15	17.4	10	15.9	11	17.2	3	8.6
A10	21	6.1	7	8.1	0	0	5	7.8	3	8.6
A11	26	7.6	10	11.6	8	12.7	5	7.8	3	8.6
A28	31	9.0	6	6.9	2	3.2	3	4.7	3	8.6
A29	16	4.6	3	3.5	2	3.2	6	9.4	3	8.6
Blank	155	44.2	35	40.7	28	44.4	22	34.4	20	57.1
B5	21	6.1	5	5.8	7	11.1	3	4.7	2	5.7
B7	93	27.2	23	26.7	18	28.6	18	26.5	14	40.0
B8	105	30.7	23	26.7	23	36.5	19	29.7	10	28.6
B12	114	33.3	30	34.9	26	41.3	16	25.0	11	31.4
B13	15	4.3	2	2.3	2	3.2	2	3.1	3	8.6
B14	29	8.5	7	8.1	4	6.3	8	12.5	2	5.7
B15	29	8.5	6	6.9	8	12.7	3	4.7	1	2.8
B16	10	2.9	4	4.6	0	0	3	4.7	1	2.8
B17	26	7.6	11	12.8	4	6.3	2	3.1	2	5.7
B18	19	5.5	6	6.9	1	1.6	4	6.2	0	0
Bw21	7	2.0	1	1.2	2	3.2	4	6.2	0	0
Bw22	13	3.8	2	2.3	1	1.6	3	4.7	2	5.7
B27	27	7.9	8	9.3	5	7.9	6	9.4	6	17.1
Bw35	29	8.5	7	8.1	3	4.8	9	14.1	2	5.7
B37	*NT		6	6.9	1	1.6	4	6.2	0	0
B40	30	8.8	8	9.3	6	9.5	12	18.7	6	17.1
Blank	117	34.2	23	26.7	15	23.8	12	18.7	8	22.9

*NT=not tested.

the Tissue Typing Laboratory at Glasgow Royal Infirmary. Twenty millilitres of blood were collected in heparinised tissue culture medium and lymphocytes separated over Ficoll-Hypaque using a differential centrifugation technique.¹⁰ Separated lymphocytes from patients were then stored in liquid nitrogen to enable the same panel of HLA typing sera to be used throughout the entire study. Histocompatibility locus antigens of A and B loci were then determined by lymphocyte microcytotoxicity test¹¹ using a panel of over 100 sera which were standardised against an International Histocompatibility Workshop cell panel. Histocompatibility locus antigen control data were derived from a population of 342 healthy unrelated individuals from the Glasgow area.¹² When only a single antigen was found in any locus a blank was recorded rather than regarding the patients as being homozygous. For some antigens, it was not possible to provide sufficient typing serum and these were omitted from the analysis.

Statistical differences between patients and controls were examined using the χ^2 test or Fisher's exact test where appropriate. Type I errors were avoided

by multiplying the p value obtained by the number of antigens tested to obtain a correct p value.¹³

Results

Blood samples were collected from 327 patients but in only 248 (76%) samples were there sufficient number of viable lymphocytes for full HLA typing. Results of 8 HLA-A antigens and 16 HLA-B antigens in the 248 patients are shown in the Table. No statistical differences were found between the patients with alcoholic liver disease and controls.

Discussion

This large study has therefore failed to confirm any evidence of an HLA associated genetic predisposition to alcoholic liver disease in Scotland and Newcastle. In particular, the frequencies of HLA-B8, B13, B15, and B40 were little different between any of the patient groups or controls, thus confirming the other well documented negative report from Liverpool in the UK.⁶

Histocompatibility locus antigen associations with

alcoholic liver disease that have been reported include an increased frequency of B5,¹⁴ B8,^{8,15} B13,¹⁶, B15,⁷ B40,¹⁷ CW3,¹⁸ DR2,¹⁴, DR3,¹⁹, and DRW9²⁰ and reduced frequencies of B13⁷ and B40.⁶ Clearly, these results are confusing and contradictory. Most of the reports are small single studies with no confirmation from other sites. Failure to allow for the increased chance of obtaining a statistically significant result unless the number of antigens tested is taken into consideration accounts for several of these spurious reports. If the probability value obtained is multiplied by the number of antigens tested then only the following associations still hold true: B8, B13, B15, B40, CW3, DR2, and DRW9. HLA-B8 and CW3 are reported to be increased in alcoholic hepatitis whereas the remaining antigens are increased in alcoholic cirrhosis, B13 being reported as raised and reduced in prevalence.

Other factors which can produce misleading results are failure to select an adequate and representative control population,¹⁵ ethnic mismatching between populations,^{8,14} lack of histological proof of the stage of liver disease in all cases⁸ and failure to examine differences between HLA antigens in all three histological stages of alcoholic liver disease.^{14,16,20}

Overall, we would conclude that there is little evidence to support the concept that any subset of HLA-A and B loci grouped individuals have an increased susceptibility to alcohol induced liver damage. The relationship with DR antigen frequency should probably be further explored in the future.

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