Effect of calcitonin and calcitonin gene-related peptide on pancreatic functions in man

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SUMMARY Calcitonin gene-related peptide (CGRP) has recently been identified in central and peripheral nerve fibres, including those of blood vessels supplying the exocrine pancreas, and in pancreatic islet cells. Moreover, receptors have been characterised in the same tissue. The present study examined the effects of human CGRP and of calcitonin on exocrine pancreatic secretion and on islet cell function in nine healthy volunteers. CGRP (300 ng/kg/h) caused, respectively, a 25% and 31% inhibition of caerulein stimulated trypsin and amylase output which was similar to that seen with calcitonin (300 ng/kg/h). Arginine stimulated insulin and glucagon release was unaffected by either CGRP, or calcitonin. Calcitonin gene-related peptide caused cutaneous flushing, but did not affect the pulse rate or arterial blood pressure in the doses tested. Calcitonin gene-related peptide inhibits exocrine pancreatic secretion in vivo in man, but does not affect islet cell hormone release.

Calcitonin gene-related peptide (CGRP) is a unique 37 amino acid peptide, which is encoded by the calcitonin gene. The peptide has been identified in the central and peripheral nervous system such as in the heart and blood vessels and in the gastrointestinal tract, and in the pituitary and thyroid glands. Little is known so far about the physiological roles of CGRP in man. Recently, CGRP receptors linked to activation of adenylate cyclase have been recognised in the cardiovascular system, and the peptide was shown to exert potent effects in man and rat which include positive chronotropic and inotropic effects on the heart and vasodilation. Other effects involve the gastrointestinal tract which include inhibition of gastric acid secretion in rats, dogs, and man, and dose dependent contraction of guinea pig ileal and colonic smooth muscles.

The recent discovery of a specific CGRP receptor on dispersed acini from the guinea pig pancreas and the stimulation of amylase release are consistent with a regulatory role of the peptide on exocrine pancreatic function. To this end, CGRP has been localised immunohistochemically in nerve terminals of the islets of Langerhans and in islet cells. An action of CGRP on the human pancreas remains to be elucidated. Thus, effects of CGRP have been studied in normal human subjects in vivo on exocrine pancreatic secretion and on the release of islet cell hormones, and compared with those of calcitonin.

Methods

SUBJECTS
Nine healthy fasting male volunteers (mean age 26, range 23–34 years; average weight 78, range 62–91 kg) without history of gastrointestinal and endocrine disorders and not receiving any medication have been studied. All subjects gave written and informed consent to the studies undertaken. They were approved by the local Ethical Human Research Committee.

PEPTIDES
Synthetic human CGRP-1 (alpha) was purchased from Peninsula Laboratories (Belmont, CA). The
peptide was dissolved in 0.9% saline containing 0.1% human serum albumin and prepared under aseptic conditions by the University of Basel Hospital Pharmacy. Vials were stored in ampoules of 25 μg CGRP at -20°C. Synthetic human calcitonin (Cibacalcin®) was donated by the Ciba-Geigy Co., Basel, Switzerland, synthetic secretin (Sekretolin®) by Hoechst-Pharma, Zurich, Switzerland, and caerulein (Takus®) by Farmitalia Carlo Erba, Berne, Switzerland.

**EXPERIMENTAL PROCEDURE**

Each subject was studied on different days and in random order.

**EXOCRINE PANCREATIC SECRETION STUDIES**

After an overnight fast, gastric and duodenal secretions were collected as described previously. Briefly, a multilumen tube was positioned under fluoroscopic guidance with the tip lying at the ligamentum of Treitz. Gastric and duodenal secretions were collected separately and continuously on ice in 15 minute aliquots. Polyethylene glycol (PEG) 4000 was perfused into the duodenum as a non-absorbable marker to correct for intestinal volume losses. After an equilibration period of 30 minutes, exocrine pancreatic secretion was stimulated with iv secretin (50 ng/kg/h=16.4 pmol/kg/h) for 150 minutes. After 60 minutes, 10 ng/kg/h caerulein (7.4 pmol/kg/h) were added to the infusion for the remaining 90 minutes of the experiments. The peptides were dissolved in 0.9% NaCl containing 0.1% human serum albumin (wt/vol). The doses used have previously been shown to submaximally stimulate pancreatic secretion. Seventy five and 300 ng/kg/h (20 and 79 pmol/kg/h) CGRP, 300 ng/kg/h (88 pmol/kg/h) calcitonin, and control solutions (0.9% NaCl) were infused through indwelling catheters positioned in a forearm for the duration of the experiments (150 minutes). Blood samples were obtained in regular intervals at baseline and at 15 minute intervals during the experiments. Blood was collected in ice chilled lithium heparinised tubes containing 5000 KIU aprotinin/5 ml blood. Samples were immediately centrifuged at 4°C and the plasma stored at -20°C until assayed for pancreatic polypeptide (PP) and somatostatin by specific radioimmunoassays (RIs).

Secretory volumes were measured to the next millilitre; bicarbonate concentration of the duodenal juice was determined by the back titration method, trypsin by the method of Wiggins, amylase according to Rick and Stegbauer, and PEG turbidimetrically. Polyethylene glycol served as a non-absorbable marker to calculate the duodenal volume for a given period by the following equation: \( V = F \times (PEG)_{perf} \times 15) / (PEG\ meas) \), where \( V \) is the calculated duodenal volume (ml/min); \( F \) is the flow rate of the PEG solution perfused (2 ml/min); (PEG) perf is the concentration of the perfusate (2 g/l); and PEG meas corresponds to the concentration of PEG in the duodenal juice collected during 15 minutes. The recovery of the duodenal marker averaged 77% (mean [SE]) whereas the average percentage of PEG recovered from the stomach was below 4%. The last three 15 minute aliquots during secretin alone and the six periods under combined stimulation with secretin and caerulein were added in each volunteer to calculate summated outputs per minute. All results are expressed as mean (SEM), if not indicated otherwise. Logarithmic transformation of data was applied when indicated.

**Fig. 1** Effects of intravenous human CGRP and calcitonin on arginine stimulated plasma glucose and insulin release. n=6, data are mean (SE). Glucagon results are not shown as they do not add any pertinent information.
Role of CGRP in pancreatic function

ISLET CELL FUNCTION TESTS
Pancreatic islet cell secretion was stimulated with iv arginine.24 After an overnight fast, indwelling catheters were inserted into both cubital veins for blood sampling and for infusions, respectively. Thirty minutes later, blood samples were drawn at 15 minute intervals before, during, and after the administration of arginine-HCl (2-8 mmol/kg) diluted in 400 ml 0-9% NaCl as indicated in Figure 1. On four different days, the subjects received either 0-9% NaCl (control), 75 or 300 ng/kg/h CGRP or 300 ng/kg/h calcitonin. Blood was collected as described above and assayed for insulin and glucagon by specific RIAs.24 During both, the exocrine pancreatic secretion studies and the islet cell function tests, heart rate and arterial blood pressure were monitored continuously.8

STATISTICAL ANALYSIS
Treatment effects in exocrine pancreatic secretory studies were analysed by paired t test. The significance level was set at p less than 0.05. Plasma concentrations of insulin, glucagon, and glucose concentrations were compared in a multi-variate analysis of variance using a commercial SAS program.

RESULTS

EFFECTS OF CGRP AND CALCITONIN ON EXOCRINE PANCREATIC SECRETION
Combined administration of secretin and caerulein significantly stimulated exocrine pancreatic secretion in all nine subjects and the secretion was maintained with the exception of trypsin output, which tended to decline towards the end of the experiments (not shown). The lower dose of CGRP (75 ng/kg/h)

Table 1 Effects of CGRP and calcitonin on caerulein-stimulated pancreatic polypeptide (PP) release and on somatostatin concentrations (SLI)

<table>
<thead>
<tr>
<th>Peptides</th>
<th>PP (pM)</th>
<th>SLI (pM)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Fasting</td>
<td>Caerulein</td>
</tr>
<tr>
<td></td>
<td></td>
<td>stimulation</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(increment</td>
</tr>
<tr>
<td></td>
<td></td>
<td>over fasting)</td>
</tr>
<tr>
<td>NaCl (control)</td>
<td>4-8 (0-7)</td>
<td>19 2 (5-0)</td>
</tr>
<tr>
<td>CGRP (300 ng/kg/h)</td>
<td>5-3 (0-8)</td>
<td>15 5 (5-6)</td>
</tr>
<tr>
<td>Calcitonin (300 ng/kg/h)</td>
<td>5 7 (0-7)</td>
<td>4-0 (2-0)*</td>
</tr>
</tbody>
</table>

Data are mean (SE) in six healthy volunteers; *indicates p<0.01 vs control.
slightly decreased pancreatic bicarbonate, trypsin and amylase output, but the difference did not reach statistical significance (p<0.1, Figure 2). The upper dose of CGRP (300 ng/h) and calcitonin, however, significantly reduced pancreatic trypsin and amylase secretion in response to the combined stimulation with secretin and caerulein (p<0.05 and p<0.01 respectively, Fig. 2). The decrease of pancreatic enzyme output brought about with 300 ng/kg/h calcitonin was of the same order than with 300 ng/kg/h CGRP.

**EFFECTS OF CGRP AND CALCITONIN ON ISLET CELL FUNCTION**

Basal PP concentrations were comparable and ranged from 4-8 to 7-2 pmol/l (Table 1). Caerulein alone stimulated the release of PP (p<0.05). Calcitonin gene-related peptide and calcitonin did not affect basal PP concentrations. Stimulation of PP by caerulein was suppressed with calcitonin (p<0.01), whereas CGRP produced a minimal inhibition of PP concentrations which did not reach statistical significance (p<0.1).

Basal somatostatin concentrations were comparable in the different experiments (Table 1). Neither CGRP nor calcitonin affected plasma somatostatin concentrations.

In the arginine infusion tests, basal insulin concentrations tended to decrease during the administration of calcitonin (p<0.1) and remained unchanged during CGRP infusions. Arginine-stimulated insulin and glucagon release was unaffected by the administration of CGRP and calcitonin (Fig. 1).

Fasting plasma glucose concentrations ranged from 3.8–4.8 mmol/l. In response to arginine infusions, plasma glucose increased significantly with the higher dose of CGRP and with calcitonin (p<0.05, Fig. 1), but the area under the curve of glucose concentrations did not show any significant difference from control experiments.

**CARDIOVASCULAR EFFECTS OF CGRP AND CALCITONIN**

Intravenous administration of CGRP did not change the pulse rate and the arterial blood pressure in the dose range tested in any of the nine subjects (Table 2). Cutaneous flushing of the face, the proximal thorax and occasionally of the palmar side of the hands was observed in all subjects with the higher dose of CGRP (300 ng/kg/h). Flushing disappeared within 30 minutes after termination of the CGRP infusions. Flushing was not recognised with 75 ng/kg/h of CGRP and no other side effects were noticed. Calcitonin did not produce changes in the pulse rate, arterial blood pressure, and no flushing was noticed.

### Table 2 Comparison of heart rate and arterial blood pressure responses to CGRP, calcitonin or saline (control) in nine healthy volunteers

<table>
<thead>
<tr>
<th>Peptide</th>
<th>Time (min)</th>
<th>−15</th>
<th>0</th>
<th>30</th>
<th>60</th>
<th>90</th>
</tr>
</thead>
<tbody>
<tr>
<td>Heart rate</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CGRP (300 ng/kg/h)</td>
<td>69 (3)</td>
<td>68 (2)</td>
<td>68 (2)</td>
<td>69 (2)</td>
<td>69 (2)</td>
<td></td>
</tr>
<tr>
<td>Calcitonin</td>
<td>68 (4)</td>
<td>68 (1)</td>
<td>69 (2)</td>
<td>69 (1)</td>
<td>66 (3)</td>
<td></td>
</tr>
<tr>
<td>NaCl (control)</td>
<td>67 (2)</td>
<td>67 (4)</td>
<td>67 (3)</td>
<td>68 (2)</td>
<td>66 (2)</td>
<td></td>
</tr>
<tr>
<td>Arterial pressure, systolic (mmHg)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CGRP (300 ng/kg/h)</td>
<td>117 (3)</td>
<td>116 (4)</td>
<td>116 (3)</td>
<td>115 (3)</td>
<td>118 (4)</td>
<td></td>
</tr>
<tr>
<td>Calcitonin</td>
<td>114 (5)</td>
<td>117 (3)</td>
<td>117 (3)</td>
<td>116 (2)</td>
<td>116 (3)</td>
<td></td>
</tr>
<tr>
<td>NaCl (control)</td>
<td>113 (3)</td>
<td>115 (2)</td>
<td>116 (3)</td>
<td>117 (2)</td>
<td>117 (2)</td>
<td></td>
</tr>
<tr>
<td>Arterial pressure, diastolic (mmHg)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CGRP (300 ng/kg/h)</td>
<td>73 (4)</td>
<td>73 (3)</td>
<td>72 (3)</td>
<td>72 (2)</td>
<td>73 (3)</td>
<td></td>
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<tr>
<td>Calcitonin</td>
<td>75 (3)</td>
<td>72 (2)</td>
<td>71 (2)</td>
<td>72 (3)</td>
<td>72 (2)</td>
<td></td>
</tr>
<tr>
<td>NaCl (control)</td>
<td>72 (2)</td>
<td>71 (3)</td>
<td>72 (3)</td>
<td>73 (4)</td>
<td>73 (2)</td>
<td></td>
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</tbody>
</table>

Data are mean (SE).

### Discussion

The wide spread presence of CGRP in peripheral nerves – for example, around the blood vessels of the gastrointestinal tract and the pancreatic islets of Langerhans suggests physiological roles of the peptide.\(^5\)\(^\text{-}^\text{12}\)\(^\text{14}\)\(^\text{-}^\text{15}\) The exocrine pancreas has only few CGRP fibres stained by immunohistochemistry, but the peptide may reach the acinar cells through the insuloacinar portal system and act as a paracrine agent.\(^1^\text{2}\) Moreover, stimulation of amylase secretion in guinea pig acinar cells by CGRP is presumably linked to a CGRP receptor and stimulation of cyclic AMP production.\(^1^\text{4}\) Furthermore, in this *in vitro* system CGRP enhanced amylase release stimulated by agents which act through mobilisation of intracellular calcium such as cholecystokinin or acetylcholine.\(^1^\text{4}\) Calcitonin gene-related peptide, however, did not influence the pancreatic secretory response to an intragastric meal of liver extracts in conscious dogs.\(^1^\text{1}\)

Here we describe for the first time effects of human CGRP on pancreatic function in normal human subjects. Infusion of 300 µg/kg/h CGRP resulted in a small significant decrease of caerulein stimulated pancreatic enzyme secretion and was of the same magnitude as the calcitonin induced inhibition of pancreatic enzyme release. Previous studies have shown inhibition of exocrine pancreatic function with human calcitonin in man, but have been obtained with far higher amounts of the peptide.\(^2^\text{6}\)\(^\text{-}^\text{27}\) At the doses of CGRP used in the present study, marked facial flushing occurred giving further evidence for the cardiovascular properties and presumably pharmacological potency of the peptide used. The
CGRP and calcitonin induced inhibition of the exocrine secretory response was not caused by somatostatin release, as neither calcitonin nor CGRP affected circulating plasma somatostatin concentrations.

Calcitonin has been shown to inhibit glucose and arginine stimulated islet cell functions in man. The doses used, however, were at least 15 times higher than in the present study. Here neither human CGRP nor calcitonin affected plasma insulin and glucagon concentrations, but calcitonin suppressed caerulein stimulated PP release. It remains to be established, if this finding has physiological relevance. Several gastrointestinal hormones stimulate calcitonin secretion such as gastrin and the cholecystokinin peptides. Here again, the effects were obtained with pharmacological rather than physiological amounts. Thus, physiological effects of calcitonin on islet cell function are unlikely to be important in man.

In view of the cutaneous flushing seen here and earlier, transcutaneous Doppler ultrasound measurements of the blood flow in the superior mesenteric artery and for comparison in the common carotid artery were obtained in healthy volunteers. While the blood flow of the superior mesenteric artery was slightly reduced by the same doses of CGRP, a marked increase in carotid blood flow was observed especially with the higher dose of CGRP (manuscript submitted).

In conclusion, the present study has revealed that iv human CGRP, unlike in vitro, induces a small inhibition of exocrine pancreatic enzyme secretion in healthy volunteers, whereas the endocrine pancreas does not appear to be an important target for the peptide. Cutaneous flushing observed here and in previous studies suggests important cardiovascular properties. We conclude that CGRP might be a regulatory peptide in the human gastrointestinal tract, but more studies are required to define its action and establish a physiological role.

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