Alteration of H₂ receptor sensitivity in duodenal ulcer patients after maintenance treatment with an H₂ receptor antagonist

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SUMMARY The effects of a specific H₂ receptor agonist impromidine, on gastric acid secretion were measured in six patients with duodenal ulcer in clinical remission before and after three months treatment with ranitidine 150 mg noxte. After treatment basal acid output increased from 1·2 to 2·8 mmol/h and after maximal impromidine stimulation from 36·9 (4·7) to 44·2 (6·2) mmol/h (p<0·02). Intravenous ranitidine 50 mg was given at the end of the impromidine infusion on each study day; the antisecretory effect of intravenous ranitidine was accentuated after the treatment with ranitidine from a trough acid output of 8·5 (1·2) mmol/h before, to 3·8 (1·5) mmol/h (p<0·05) after, treatment. The increased response to the H₂ agonist impromidine and the H₂ antagonist ranitidine after treatment with ranitidine suggests an enhanced sensitivity of the H₂ receptor. This might be explained on the basis of an increase in the number of H₂ receptors (‘up-regulation’).

The discovery of the histamine H₂ receptor on the parietal cell1 initiated the development of H₂ receptor antagonists with potent gastric antisecretory activity and these agents have now become firmly established in the treatment of peptic ulceration.2 After healing of duodenal ulcer by the H₂ receptor antagonists, cimetidine and ranitidine, maintenance treatment with either of these drugs has successfully reduced ulcer relapse rates at 12 months from approximately 80% without therapy to about 30% on continuous maintenance treatment.3 There has been some suggestion that the antisecretory effects of H₂ receptor antagonists may alter with continuous H₂ receptor blockade during prolonged treatment. Previous studies have utilised either pentagastrin or peptone meals as secretagogues in tests of gastric acid secretion.4 5 Because the inter-relationship between histamine, muscarinic and gastrin receptors is complex6 neither pentagastrin nor a peptone meal specifically identifies a potential alteration in H₂ receptor sensitivity after maintenance treatment with a competitive antagonist. Impromidine, however, is a highly potent, and specific agonist for the H₂ receptor7 exhibiting up to 27 times the affinity of histamine.8 In this study we have examined the effect of three months of treatment with ranitidine 150 mg nocte on impromidine stimulated gastric acid secretion in six patients with duodenal ulcer in remission.

Methods

Patients Six male patients, aged 33 to 62 years, participated in the study. All had endoscopically confirmed duodenal ulcer previously healed on conventional doses of H₂ receptor antagonists before entry in the study. All the patients were non-smokers and none took alcohol to excess. None of the patients had concurrent disease or was taking any other medication. The study was approved ethically by the Research Advisory Committee of the Chedoke-McMaster Hospitals and written, informed consent was obtained from all participants.

STUDY DESIGN Patients were pretreated with ranitidine 150 mg nocte for three days before the first study day. The first impromidine stimulated gastric secretion study...
was carried out 10 hours after the last dose of ranitidine. On each study day at 0900 hours, after an overnight fast, a 14 French Salem Sump tube (Sherwood Medical Industries, St Louis, MO, USA) was placed in the most dependent part of the stomach, and its position checked by the water recovery test. Gastric juice was collected by continuous aspiration divided into 10 minute intervals throughout the study. After a one hour basal period, an impromidine (Smith, Kline and French Research Ltd, Welwyn, Herts, UK) step dose intravenous infusion was started at a dose of 2.5 μg/kg/h for 90 minutes. After 90 minutes, the infusion rate was doubled to 5.0 μg/kg/h for a further 90 minutes, and then doubled again to 10 μg/kg/h for 210 minutes. At the end of 90 minutes of impromidine 10 μg/kg/h, ranitidine 50 mg was administered intravenously over five minutes and gastric secretion continuously aspirated for the next 120 minutes during continued high dose impromidine infusion.

The volume of each sample was measured, and acid concentration determined by automatic titration (Radiometer, Copenhagen) to pH 7.00 with 0.1 M NaOH. Acid output was calculated as the product of volume and concentration.

After the impromidine study, all six patients were continued on three months treatment with ranitidine 150 mg nocte. Within one week of completion of the three months course, the impromidine study was repeated in an identical fashion and started 10 hours after the last dose of ranitidine. This approach generated data for each patient before and at the end of a three month course of ranitidine 150 mg/h.

**DATA PRESENTATION**

Basal acid output (BAO) was calculated by summation of the acid outputs over the one hour before impromidine infusion and expressed as mmol/h. Plateau acid outputs during impromidine infusion (PIAO) was calculated by summation of the last six 10 minute aliquots of each 90 minute infusion period, and the results expressed as mmol/h. In order to estimate the maximal effect of ranitidine on impromidine stimulated acid output, acid output measured between 60 and 90 minutes after bolus ranitidine (representing trough acid output) were summated, and expressed as mmol/h.

**Table**  Pre and post ranitidine treatment impromidine dose response infusion results for acid output expressed in mmol/h

<table>
<thead>
<tr>
<th>Basal</th>
<th>Imp 2.5</th>
<th>Imp 5.0</th>
<th>Imp 10</th>
<th>Ran 50 mg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pre</td>
<td>1.91 (0.38)</td>
<td>7.15 (2.80)</td>
<td>21.32 (5.39)</td>
<td>36.90 (4.7)</td>
</tr>
<tr>
<td></td>
<td>2.79 (0.89)</td>
<td>22.49 (6.16)</td>
<td>35.31 (6.38)</td>
<td>44.17 (6.2)</td>
</tr>
<tr>
<td>Post</td>
<td>0.25</td>
<td>0.04</td>
<td>0.05</td>
<td>0.02</td>
</tr>
</tbody>
</table>

**Figure**  Gastric acid secretion, before and after three months maintenance ranitidine 150 mg treatment, expressed as acid output (mmol/h), in basal state (BAO), after three infusion rates of impromidine, and after bolus intravenous ranitidine 50 mg.

**STATISTICAL ANALYSIS**

Data are presented as the mean (SE). Statistical comparisons were made by paired, two tailed, Student’s *t* test," accepting a *p* value of 0.05 as the minimal level of significance.

**Results**

Before and after maintenance ranitidine 150 mg nocte, impromidine resulted in a dose dependent stimulation of gastric acid secretion (Table, Figure) to a maximal acid output of approximately 40 mmol/h during the highest dose. Apart from transient facial flushing, the impromidine infusion was well tolerated with no serious adverse effects. The intravenous bolus of ranitidine 50 mg resulted in a prompt and dramatic decrease in gastric acid output during the continued high dose impromidine infusion (Table, Figure).

Three months of treatment with ranitidine 150 mg nocte resulted in a non-significant increase in basal acid output from 1.2 (0.4) to 2.8 (0.9) mmol/h (*p*=0.25). At each of the three stepwise doses of impromidine, three months ranitidine treatment resulted in a significant (*p*<0.05) increase in impromidine stimulated gastric acid secretion compared with pretreatment values (Table). At the highest dose of impromidine infusion, gastric acid output increased from 36.9 (4.8) mmol/h before ranitidine 150 mg treatment to 44.2 (6.2) after (*p*<0.02).

Similarly, ranitidine treatment resulted in an accentuated response to the intravenous bolus of
ranitidine 50 mg (Figure). The pretreatment trough acid output before maintenance treatment was 8.5 (1.2) mmol/h, and was reduced to 3.8 (1.6) mmol/h (p<0.05) after maintenance treatment (Table).

**Discussion**

A variety of stimuli result in increased gastric acid secretion, including histamine, cholinergic and gastrin stimulation. As a working hypothesis, it is postulated that the parietal cell has distinct receptors for histamine, acetylcholine, and gastrin and that these receptors interact with, and may potentiate each other. Thus, for example, pentagastrin stimulated acid secretion may be blocked by \( \mathrm{H}_2 \) receptor antagonists. This interaction makes study of the \( \mathrm{H}_2 \) receptor difficult *in vivo*. In this study we used the potent, specific secretagogue imipramine which is an agonist with a high degree of specificity for the \( \mathrm{H}_2 \) receptor. It has no direct effect on muscarinic or gastrin receptors. Imipramine has between nine and 48 times the activity of histamine *in vitro* at the \( \mathrm{H}_2 \) receptor, and is up to 27 times more potent than histamine in stimulating gastric acid secretion *in vivo*. In previous studies of human gastric acid secretion, an infusion of 10 \( \mu \)g/kg/h achieved virtually maximal acid output.\(^{35}\)

Interaction of histamine, or an \( \mathrm{H}_2 \) agonist, with the \( \mathrm{H}_2 \) receptor on the parietal cell results in activation of adenyl cyclase. The resultant increase in intracellular cyclic AMP results in activation of type I and II phosphokinases, with possible subsequent mobilisation of calcium ions from intracellular stores.\(^{14}\) Activation of the phosphokinase system results in alteration of the intracellular cytoskeleton with activation of the secretory canaliculus initiating hydrogen ion secretion by the proton pump. The end result is hydrogen ion secretion from the secretory canaliculus into the gastric lumen.\(^{14}\) *In vitro* studies have shown that incubation of human gastric cancer cell lines (HGT-1) with histamine results in desensitisation of the \( \mathrm{H}_2 \) receptor after only three hours. This phenomenon is known as ‘down-regulation’ of the receptor and for any given ligand receptor interaction, the resultant biological event will be reduced. Conversely, it might be expected that continued exposure of the \( \mathrm{H}_2 \) receptor to an antagonist might result in ‘up-regulation’ of the receptor. In previous studies,\(^{44}\) the apparent increase in stimulated gastric acid secretion after continued \( \mathrm{H}_2 \) receptor antagonist treatment could have been attributed to a simple increase in parietal cell mass, to an increase in \( \mathrm{H}_2 \) receptor density, or to true biological ‘up-regulation’ of the \( \mathrm{H}_2 \) receptor. The present study suggests that the phenomenon of true ‘up-regulation’ has occurred after three months maintenance ranitidine treat-

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**References**


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