Ileal and colonic fatty acid profiles in patients with active Crohn’s disease

S Bühner, E Nagel, J Körber, H Vogelsang, T Linn, R Pichlmayr

Abstract
In patients with active Crohn’s disease and in a control group the fatty acid profiles in the whole lipid fraction of ileal and colonic mucosal biopsy specimens were determined by capillary gas chromatography. The biopsy specimens in Crohn’s disease patients were taken from the inflamed terminal ileum as well as from the inflamed and macroscopically normal colon. Compared with controls the fatty acid distribution in the inflamed ileal mucosa was significantly characterised by (a) a decrease of 18:2 n6 and 18:3 n3 accompanied by a substantial increase of the highly polyunsaturated fatty acids 20:4 n6, 22:4 n6, and 22:6 n3 and (b) a higher unsaturation index of total fatty acids compared with controls. These changes were similar in the inflamed colon. Additionally, both the inflamed and the macroscopically normal colonic mucosa showed an increase of saturated (18:0) and a decrease of monounsaturated fatty acids (18:1 n9). Fatty acid profiles of ileum and colon showed side variations in controls, but not in the Crohn’s disease group. These data suggest that in Crohn’s disease changes in the distribution of polyunsaturated fatty acids seem to be the general feature of inflamed mucosa in small and large intestine. Results further suggest that colonic fatty acid metabolism in Crohn’s disease is altered by degrees, showing changes in saturated and mono-unsaturated fatty acids as an additional, primary event.

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In inflammatory bowel disease (IBD) particular eicosanoids are increased in inflamed intestine.1–3 These substances have proinflammatory properties and derivate from an n6 fatty acid, the arachidonic acid (20:4 n6). Taking advantage of the competitive inhibition between n6 and n3 fatty acids, one therapeutic approach focuses on the modification of the eicosanoid synthesis by dietary supplementation with fish oils, which are rich in long chain highly polyunsaturated n3 fatty acids.4–5 Essential functions of polyunsaturated fatty acids in the gut, however, depend on their balance between n6 and n3 fatty acids5 and on their balance between saturated and unsaturated fatty acids.7 An important example is the control of membrane properties.6,7 This is one reason why the therapeutic use of high doses of n3 fatty acids is not undisputed. Furthermore, for unknown reasons, clinical benefit seems to be confined to ulcerative colitis,8 which affects only the colon. In Crohn’s disease, which may involve both small and large intestine, local fatty acid metabolism might be different.

Basic knowledge of fatty acid metabolism in patients with IBD is limited. Three recent studies showed changes in plasma fatty acid profile in patients with inactive9,10 and active IBD.11 Studies concerning the intestinal mucosal fatty acid profile of the terminal ileum and the colon in patients with Crohn’s disease and in a control group. We were basically interested in characteristics and site variations of changes in inflamed areas. As another study shows that the metabolism of the ‘quiescent’ colon in IBD might already be altered,12 we additionally assessed the fatty acid profile of macroscopically normal colonic mucosa in patients with Crohn’s disease.

Methods

PATIENTS
Studies were carried out on mucosal biopsy specimens, which were obtained from patients undergoing routine endoscopic examinations of the small and large bowel. Each patient gave informed consent.

Diagnosis for Crohn’s disease (n = 15) was based on full clinical examination including endoscopy, histological examination of mucosal biopsy specimens, and radiographic examination; the first diagnosis was 2–19 years before the current examination. All patients had acute symptoms. Routine endoscopy showed inflammatory activity in the mucosa of the colon (n = 6), the terminal ileum (n = 4), or both (n = 5). Inflamed mucosa was reddened, friable, oedematous, and characterised by spontaneous bleeding, focal ulcerations, and aphthous lesions. Non of the patients had been surgically treated; 9 of 15 were receiving drug treatment (Table I).

The control group consisted of 16 women and men undergoing routine endoscopy for various diseases of the gastrointestinal tract (Table I). An acute or (present or past) chronic inflammation of the bowel was excluded on the basis of the complete anamnesis and the clinical parameters listed above. Non of the patients were suffering from acute diarrhoea.
There was no surgical pretreatment or current drug treatment in this group. At the time of this study all patients lived at home and were receiving a normal, Western diet. The nutritional habits were obtained from a questionnaire. Subjects with: (a) greatly reduced food intake, (b) dietary regimens, such as diabetic or vegetarian diets, and (c) special dietary treatments including vitamin and fish oil supplements during the previous six months, were ruled out and not examined in this study. None of the patients excluded a particular dietary fat (such as a particular vegetable fat or animal fat) from their daily food.

### Analysis of Fatty Acids

For fatty acid analysis one mucosal biopsy specimen was taken from each patient either at the terminal ileum (5–10 cm proximal to the ileocaecal valve) or at the colon (left flexura) or at both sites (Table I). Control specimens had a normal mucosal appearance; in Crohn’s disease these areas were inflamed. Additionally, specimens of endoscopically normal colonic mucosa were obtained from nine patients with Crohn’s disease. A distance of at least 10 cm was maintained from any inflamed localisations in the colon (n=6). The specimens were carefully washed and stored at -80°C.

For subsequent fatty acid analysis in the whole lipid fraction, samples were weighed (range: 10–15 mg), homogenised separately with liquid nitrogen in a mortar, and kept in 4 ml 2:1 chloroform/methanol (vol/vol) at 4°C for 24 hours. After volume reduction in a nitrogen stream and addition of heptadecanoic acid methyl ester (C17:0) as internal standard, the samples were saponified overnight (4°C) with 2.5 ml 90% methanol potassium hydroxide solution. Fatty acid methyl esters were prepared by boiling on reflux with 7.5 ml 10% boron trifluoride-methanol-complex at 80°C for 20 minutes, fourfold extraction with hexane, drying on molecular sieves, and careful evaporation in the vacuum and with a nitrogen stream.

<table>
<thead>
<tr>
<th>Complications:</th>
<th>Control (n=16)</th>
<th>Crohn’s disease (n=15)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Perianal fistula</td>
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<td></td>
</tr>
<tr>
<td>Dietary habits</td>
<td>Normal Western diet</td>
<td>Normal Western diet</td>
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<td>Sample localisations:</td>
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<td></td>
</tr>
<tr>
<td>Ileum only</td>
<td>6</td>
<td>4</td>
</tr>
<tr>
<td>Ileum + colon</td>
<td>4</td>
<td>5</td>
</tr>
<tr>
<td>Colon only</td>
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<td>6</td>
</tr>
<tr>
<td>Colon (macroscopically normal)</td>
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<td></td>
</tr>
</tbody>
</table>

#### Figure 1: Fatty acid distribution in inflamed ileal and colonic mucosa in patients with Crohn’s disease. Values are given as percentages of total fatty acids. Fatty acids are listed in the order of their metabolic pathways; ▲ indicates a skipped step. *indicates differences between the ileum and the colon in the control group; △ indicates differences between controls and Crohn’s disease in either the ileum or the colon; p<0.05, Wilcoxon-Mann-Whitney test, mean (SD).
TABLE II Fatty acids in inflamed ileal and colonic mucosa in patients with Crohn’s disease

<table>
<thead>
<tr>
<th></th>
<th>Ileum</th>
<th></th>
<th>Colon</th>
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</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>Crohn’s disease</td>
<td>Control</td>
<td>Crohn’s disease</td>
</tr>
<tr>
<td>Estimated data</td>
<td></td>
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<td></td>
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<tr>
<td>SFPA (%)</td>
<td>35.9 (4.0)*</td>
<td>36.1 (2.7)</td>
<td>31.1 (1.5)</td>
<td>33.3 (2.7)</td>
</tr>
<tr>
<td>SFPA/ΣUA</td>
<td>0.6 (0.1)*</td>
<td>0.6 (0.1)</td>
<td>0.4 (0.03)</td>
<td>0.5 (0.1)</td>
</tr>
<tr>
<td>UNID</td>
<td>126 (6)†</td>
<td>142 (11)</td>
<td>116 (8)†</td>
<td>135 (5)†</td>
</tr>
</tbody>
</table>

Values of fatty acids (FA) are calculated as percentage of total FA. *Indicates differences between ileum and colon in the control group; † indicates differences between controls and Crohn’s disease in either the ileum or the colon; p<0.05, Wilcoxon-Mann-Whitney test, mean (SD); SFPA: sum of saturated FA; ΣUA: sum of unsaturated FA; UNID: Σ(FA percentage×number of double bonds).

The residue was dissolved in 200 μl heptan and submitted to fatty acid measurements with a 5890 Hewlett Packard gas chromatograph using nitrogen as carrier gas (pressure: 16 psi). It was equipped with a flame ionisation detector and a 50 m×0.22 mm internal diameter capillary column (CP SIL88, Chrompack, Frankfurt, Germany). Initial oven temperature was 60°C for one minute followed by heating periods of 20°C/min up to 180°C and 5°C/min to reach the final temperature of 200°C; the injector and detector temperatures were set at 250°C and 270°C, respectively.

Fatty acids (C14:0–22:6 n3) were identified using standards (Sigma, Deisenhofen, Germany) and their portion in the samples was determined as a percentage of total fatty acid (wt/wt).

STATISTICS

Data were presented as mean (SD), whereby data from similar sites in each patient group were added. They were checked for significant differences using the Wilcoxon-Mann-Whitney test with a probability value of p<0.05.

Results

The mucosal fatty acid profile of the inflamed terminal ileum in patients with Crohn’s disease compared with controls was basically characterised by significant changes in the distribution of polysaturated fatty acids (Fig 1). In the series of n6 fatty acids, a decrease of the essential linoleic acid (18:2 n6) was accompanied by an increase of highly polyunsaturated desaturation and elongation products, arachidonic acid (20:4 n6) and docosatetraenoic acid (22:4 n6). A similar pattern was obvious in the series of the n3 fatty acids showing a decrease of α linolenic acid (18:3 n3) and an increase of docosahexaenoic acid (22:6 n3) percentages. The proportions of dihomo-γ-linolenic acid (20:3 n6), eicosapentaenoic acid (20:5 n3) of the individual series, and the proportion of the n9 fatty acid eicosadienoic acid (20:2 n9) varied only slightly between the control group and the patients with Crohn’s disease. The ratio of saturated and unsaturated fatty acids in the mucosa was unaffected by the current inflammation (Table II). In contrast, the unsaturation index, taking into account the number of double bonds of the particular fatty acid, increased compared with controls.

So far the results in the inflamed colon confirmed the findings in the terminal ileum. The colonic fatty acid profile in Crohn’s disease was, however, additionally characterised by an increase of saturated fatty acids (stearic acid (18:0)) and a decrease of monounsaturated fatty acids (palmitoleic acid (16:1 n7), oleic acid (18:1 n9)) compared with controls (Fig 1). Therefore the sum of saturated fatty acids as well as the ratio of saturated and unsaturated fatty acids tended to increase (Table II). A similar change of C18 saturated and monounsaturated fatty acids was already obvious in macroscopically normal colonic sites of patients with Crohn’s disease (Fig 2). The distribution of polysaturated fatty acids in these sites, however, did not differ significantly from that in controls.

Differences between the fatty acid profiles of small and large intestine occurred only in the control group, being characterised by a lower percentage of 18:0 and a higher percentage of 18:1 n9 in the colon (Fig 1). Accordingly, the sum of saturated fatty acids and the quotient of saturated and unsaturated fatty acids were lower in colonic than in ileal mucosa (Table II).

A small percentage of structurally altered transisomeric fatty acids was found in all mucosal samples. The percentage of elaidinic acid, the most common 18:1 n9 transisomer, was on average 0.8% of total fatty acids, showing no differences between controls and Crohn’s disease or local differences.

The current drug treatment in Crohn’s disease had no significant effects on the proportions of eстерified fatty acid in the ileum or the colon. An example is given for ileal 20:4 n6: untreated patients (n=3) 12.3 (1-8)% treated patients (n=6) 13.3 (1-9)%, and colonic 20:4 n6: untreated patients (n=5) 12.7 (2-2)% treated patients (n=6) 10.4 (1-8)%.

Discussion

In active Crohn’s disease the fatty acid profiles of both inflamed ileal and colonic mucosa are

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*Figure 2: Macroscopically normal (NAD: no abnormality discovered) colonic areas of patients with Crohn’s disease were characterized by a higher percentage of 18:0 and a lower percentage of 18:1 compared with controls, respectively (p<0.05, Wilcoxon-Mann-Whitney test). The distribution of other fatty acids was unaltered, mean (SD).*
basically characterised by a change in distribution of n3 and n6 polyunsaturated fatty acids compared with controls. Changes in the proportions of saturated and monounsaturated fatty acids are an additional feature but limited to colonic mucosa. It is most likely that these differences derive from the current inflammatory disease. Firstly, the nutritional recordings did not provide evidence of any significantly different dietary fat intake in the two patient groups. This is important because dietary fats are known modulators of intestinal fatty acid distribution. Secondly, patients with Crohn’s disease could suffer from essential fatty acid deficiency. The typical fatty acid pattern, however, indicating this status (decrease of n6 and increase of n9 fatty acids) was not obvious in this group. Thirdly, the current drug treatment in Crohn’s disease patients did not significantly influence the proportion and distribution of esterified fatty acids in ileal and colonic mucosa. This is in agreement with findings in the rectum. It is known that these drugs do influence the fatty acid metabolism, but this may concentrate on free, but not on esterified fatty acids, as shown for arachidonic acid.

Profound changes of polyunsaturated fatty acids similarly characterised the large and small intestine in Crohn’s disease and might be regarded as a general feature of the chronically inflamed areas. Regarding the pattern of changes it is striking that an increase of long chain polyunsaturated n3 and n6 fatty acids is combined with a decrease of the individual essential precursors – that is, 18:2 n6 and 18:3 n3. This would suggest an enhanced local biosynthesis of long chain polyunsaturated fatty acids or a substitutive incorporation of fatty acids. This second mechanism is known in the regulation of structural-functional homoeostasis of membranes. Evidence of both mechanisms is provided by studies in rats. To clarify this finding, however, it is necessary to define the mucosal compartments or cell types, which are the predominant site of changes in individual fatty acids and to elucidate their metabolic interactions. The change in the fatty acid profile led to an increase of the unsaturation index in the inflamed mucosa. This might contribute to the enhanced intestinal permeability in patients with Crohn’s disease.

In detail, this study showed a considerable increase of esterified 20:4 n6 in the inflamed areas. This confirms findings in the rectum of patients with Crohn’s disease. As repeatedly supposed, this may be directly related to the increase of saturated and monounsaturated fatty acids in the mucosa of patients with Crohn’s disease. The proportions of 20:3 n6 and 20:5 n3, which are known precursors of other eicosanoids, did not differ between controls and Crohn’s disease. They may therefore play a minor part in the pathomechanism of the disease. The increase of 22:6 n3 is interesting, as high concentrations are also reported in the plasma of IBD patients. There is still little knowledge about the biological activity and role of 22:6 n3 in the intestine. Recently an in vitro study reported an inhibition of mammalian prostaglandin synthetase by 22:6 n3. It could be cautiously speculated that the high proportion might therefore have a regulatory implication. This point needs detailed in vivo analysis, however, especially to be certain of the validity of a further increase of 22:6 n3 by supplementation with fish oils in the treatment of IBD.

Changes in the distribution of saturated and monounsaturated fatty acids also occurred in the patients with Crohn’s disease, but this was only a feature of the colon not of the terminal ileum. It was basically an increase of 18:0 and decrease of 18:1 n9 proportions compared with controls. The changes led to an alignment of ileal and colonic fatty acid profiles in patients with Crohn’s disease, which contrasts with the findings in the control group. It would be interesting to discover if this is followed by a functional alignment. These findings suggest site variations between the small and large intestine in specific fatty acid incorporation or intestinal Δ 9 desaturase activity in response to pathophysiological signals. Studies in rats provide evidence of such mechanisms, showing intestinal site variations in these parameters during fasting or different dietary fat composition. However, information in humans is rare. A most striking finding was that these individual colonic changes were not limited to the inflamed areas, but were already seen in macroscopically normal colon in patients with active Crohn’s disease. The distribution of polyunsaturated fatty acids remained unchanged. This suggests that colonic fatty acid metabolism in Crohn’s disease is altered by degrees, showing changes in saturated and monounsaturated fatty acids as a primary step. It supports the concept that the ‘quiescent’ mucosa in Crohn’s disease is not at all quiescent, as already reported for morphological parameters. This corresponds with recent findings in Crohn’s disease indicating that quiescent mucosal cells of the colon are probably primed for a rapid change of metabolism.

The changes in colonic fatty acid profile do not seem to be specific for Crohn’s disease as our data show clear similarities to those ascertained for the inflamed colon in ulcerative colitis. Variations (concerning the proportions of 18:2 n6 and 18:3 n3) might result from the fact that in ulcerative colitis the analysis focused only on the phospholipid fraction of mucosal fat.

Additionally, we checked the mucosal fatty acid profiles for the presence of structurally altered fatty acids. In Crohn’s disease, it has been suggested that these fatty acids play a part in the pathogenesis of Crohn’s disease. Their proportion in the adipose tissue of patients with Crohn’s disease was found to be unusually high. According to our results, this is not the case in the intestinal mucosa. Therefore it has to be assumed that in our patient group the consumption of dietary fats containing these fatty acids was not actually increased.

When comparing these results with recently
published data on plasma fatty acid profile in inactive 10 and active Crohn’s disease, 11 it is seen that apart from one fatty acid (22:6 n3) the changes of the fatty acid distribution in the plasma phospholipids were concordant with the changes in the intestinal mucosa. This clearly shows that plasma fatty acid composition in IBD did not reflect tissue composition, as was recently stated. 11 The two compartments obviously react differently to disease dependent signals in the organism. It is difficult to draw conclusions, however, on the metabolic flux based on the fatty acid patterns alone. In this context recent studies in rats stress the importance of enzymatic assays of the fatty acid pathway. Their findings strongly suggest, for example, that liver and intestinal desaturase enzymes respond differently to changes in physiological conditions such as fasting or dietary n3 fatty acid intake. 22 32 It would be an advance to elucidate such mechanisms for the pathophysiological condition of IBD especially in terms of dietary fat treatment in these patients.

This study showed profound changes of ideal and colonic fatty acid profiles in active Crohn’s disease. Thereby changes in polyunsaturated fatty acids seem to be a general feature of inflamed areas while changes in saturated and monounsaturated fatty acids are limited to the colon but already involve the so called ‘quiescent’ areas.

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