

Major virulence factors, VacA and CagA, are commonly positive in *Helicobacter pylori* isolates in Japan

S Maeda, K Ogura, H Yoshida, F Kanai, T Ikenoue, N Kato, Y Shiratori, M Omata

Abstract

Background—VacA and CagA proteins have been reported to be major virulence factors of *Helicobacter pylori*. However, antibodies against these proteins are frequently found in the sera of Japanese patients regardless of their gastro-duodenal status.

Aim—To evaluate the expression of VacA and CagA proteins by *H pylori* strains isolated in Japan.

Methods—By using specific antibodies raised against recombinant VacA and CagA proteins, the expression of VacA and CagA was evaluated in 68 *H pylori* strains isolated from Japanese patients; a vacuolating assay and genotyping of the *vacA* gene were also used in the evaluation. The results were analysed in relation to the gastro-duodenal diseases of the hosts.

Results—VacA and CagA proteins were expressed in 59/68 (87%) and in 61/68 (90%) isolates respectively. The vacuolating assay was positive in 57/68 (84%) isolates, indicating that most immunologically VacA positive strains produced active cytotoxin. The prevalence of infection with strains expressing CagA and positive for vacuolating activity (Type I) was very high, 54/68 (79%), irrespective of the gastro-duodenal status of the host.

Conclusion—Most *H pylori* isolates in Japan are positive for vacuolating cytotoxin and CagA, and thus these virulence factors cannot be used as markers to discern the risk of developing serious gastro-duodenal pathologies in the hosts. However, the high prevalence of infection with strains positive for vacuolating cytotoxin and CagA may contribute to the characteristics of *H pylori* infection in Japan.

(Gut 1998;42:338-343)

Keywords: VacA; *Helicobacter pylori*; CagA; ulcer; non-ulcer dyspepsia

Helicobacter pylori is a Gram negative microaerophilic bacterium that can infect human gastric mucosa.¹ The organism is closely associated with not only active gastritis but also peptic ulcers and gastric cancer.²⁻⁵ The prevalence of *H pylori* infection is high world wide, and about 80% of the general population are infected at age 40-50 in Japan.⁶ However, the infection leads to peptic ulcers or gastric cancer in only a small subset of infected patients. Although the

pathogenesis of *H pylori* related gastro-duodenal diseases is not well understood, there are several putative virulence factors of *H pylori* that may contribute to the exacerbation of mucosal damage.

Vacuolating cytotoxin, which induces vacuolation in cultured eukaryotic cells, is secreted by about 50% of *H pylori* isolates in Western countries. Infection with vacuolating cytotoxin positive strains is reported to be associated with particular gastro-duodenal diseases.⁷⁻¹¹ For instance, Figura *et al*⁷ showed that strains with vacuolating cytotoxin activity were found in 16 of 24 (67%) patients with peptic ulcers but in only 16 of 53 (30%) patients without. The cytotoxin is encoded by the *vacA* gene,¹²⁻¹⁵ and thus is also called VacA protein.

CagA protein is another putative virulence factor, encoded by the *cagA* gene (cytotoxin associated gene A).^{16,17} It has been reported that this gene is present in about 60% of *H pylori* isolates, and nearly all *cagA* gene positive strains express the protein.¹⁸ Although the function of CagA protein is not known, the protein has been reported to be associated with particular gastro-duodenal diseases.¹⁹⁻²² Ching *et al*²² found serum anti-CagA antibody in 165 of 197 (84%) patients with duodenal ulcer, but in only 25 of 45 (56%) patients with non-ulcer dyspepsia. Recently, CagA positive *H pylori* strains have been shown to be associated with interleukin-8 (IL-8) induction in gastric epithelium. Neutrophilic infiltration into the gastric epithelium, which is characteristic of *H pylori* infection, may be due to increased production of IL-8. CagA protein may thus be related to gastric inflammation and gastro-duodenal diseases.²³

Expression of CagA protein is closely associated with that of vacuolating cytotoxin,¹⁸ although the underlying mechanism is not understood. Thus Xiang *et al*¹⁹ classified *H pylori* strains into two groups, Type I and Type II. They and others reported that the Type I strains, which are positive for both vacuolating cytotoxin and CagA, were strongly associated with peptic ulcer diseases in the host.^{19,24}

However, there have been few reports on the expression of VacA and CagA proteins in *H pylori* strains isolated in regions other than Western countries. In previous studies, we made recombinant CagA and VacA proteins and investigated serum anti-CagA and anti-VacA antibodies in Japanese patients with *H pylori* infection. These studies showed unexpectedly high seropositivity for both

Second Department of Internal Medicine, Faculty of Medicine, University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113, Japan

S Maeda
K Ogura
H Yoshida
F Kanai
T Ikenoue
N Kato
Y Shiratori
M Omata

Correspondence to:
Dr S Maeda.

Accepted for publication
23 September 1997

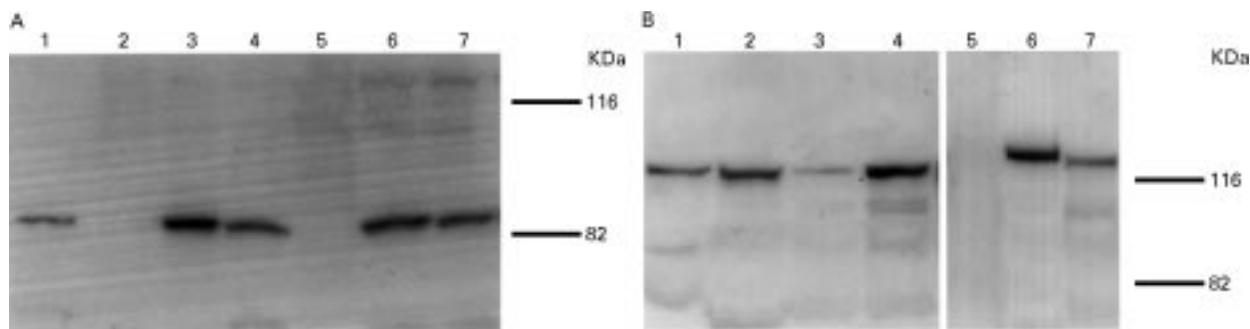


Figure 1 (A) Immunoblot analysis of VacA. Surface proteins obtained from seven *H pylori* strains were probed with antiserum against the recombinant VacA protein. VacA protein was expressed in T-1 (lane 1), T-5, T-6 (lanes 3 and 4), T-24 and T-39 (lanes 6 and 7), but not in T-48 (lane 2) or T-25 (lane 5). (B) Immunoblot analysis of CagA. Total lysates obtained from seven *H pylori* strains were probed with antiserum against the recombinant CagA protein. CagA protein was expressed in T-1, T-48, T-5, T-6 (lanes 1–4), T-24 and T-39 (lanes 6 and 7), but not in T-25 (lane 5). Molecular mass is shown in kDa.

anti-VacA²⁵ and anti-CagA²⁶ antibodies irrespective of the gastroduodenal disease.

Since the presence of antibodies does not necessarily imply on-going infection, our previous studies suggested but did not prove high prevalence of infection with Type I strains in Japan. Therefore we conducted the current study to examine, by immunoblot analysis using specific antibodies raised against recombinant proteins, the expression of VacA and CagA proteins by *H pylori* isolates obtained from Japanese patients.

All *H pylori* isolates reported so far were found to possess the *vacA* gene, and strains without cytotoxin activity are thought to produce inactive cytotoxin and/or have defective secretion mechanisms.²⁷ Thus we also examined the vacuolating activity and the genotypes of the *vacA* gene reported to be associated with the ability to secrete vacuolating cytotoxin.

Materials and methods

BACTERIAL STRAINS AND GROWTH

H pylori strains were isolated from gastric biopsy samples of 68 patients (40 men and 28 women; age range 22–85 years; mean age 52.5) who underwent endoscopy in Tokyo University Hospital. The endoscopic findings were gastric ulcer in 16, duodenal ulcer in 17, non-ulcer dyspepsia in 25, and gastric cancer in 10 patients. Biopsy specimens were cultured on Columbia agar with 5% horse blood and Dent antibiotic supplement (Oxoid, Basingstoke, Hants, UK) at 37 °C for five days under micro-aerobic conditions. Isolates were identified as *H pylori* by Gram stain morphology and by positive urease, catalase, and oxidase activities. They were kept at –80 °C in brucella broth with 5% fetal bovine serum (FBS) containing 16% glycerol. ATCC43526 cells were used as VacA and CagA positive controls, and strain Tx30a was used as CagA negative control.^{25 26}

RECOMBINANT VACA AND CAGA PROTEINS AND ANTISERA TO RECOMBINANT VACA AND CAGA

Recombinant VacA²⁵ and CagA²⁶ proteins and the rabbit antiserum to each recombinant protein were prepared as described previously. In brief, the entire *vacA* gene (GenBank accession number AF001358) and *cagA* gene (GenBank accession number AF001357) of the

ATCC43526 *H pylori* strain were cloned, and partial fragments of *vacA* (encoding amino acids 1, which is the ATG start codon, to 744) and *cagA* (encoding amino acids 107–531) genes were subcloned in frame with the sequence coding for the histidine tag of pQE vectors (Qiagen, Chatsworth, CA, USA). The histidine-tagged VacA and CagA proteins expressed in *Escherichia coli* were purified on Ni/nitrilotriacetic acid resin (Qiagen). Antibodies against the recombinant proteins were raised in New Zealand White rabbits.

DETECTION OF VACA AND CAGA PROTEINS IN H PYLORI ISOLATES

Immunoblot analysis was performed to detect VacA and CagA proteins produced by *H pylori* strains. VacA protein was examined using *H pylori* surface proteins prepared by the method of Xiang *et al.*¹⁹ In brief, the cultured bacterial cells were washed twice with distilled water, and, to separate the surface protein from bacterial cells, vortex mixed with distilled water for one hour at room temperature. After centrifugation at 10 000 *g* for ten minutes, the supernatants were stored at –30 °C until use.

CagA protein was examined using whole cell lysates. The whole cell lysates were washed twice in normal saline and centrifuged, and boiled at 100 °C for 5 min in Laemmli sample buffer before use.²⁸ The surface proteins and the whole cell lysates were separated by electrophoresis in sodium dodecyl sulphate/10% polyacrylamide gel and transferred to a nitrocellulose membrane (Schleicher & Schuell, Dassel, Germany). The filter was blocked with buffer containing 5% skimmed milk in 25 mM Tris/HCl, pH 7.4, containing 150 mM NaCl and 5 mM KCl. The membrane was incubated with a 1:100 dilution of rabbit antiserum against the recombinant VacA or CagA. After being washed, the membrane was incubated with ¹²⁵I-labelled goat anti-rabbit immunoglobulin G, and then exposed to *x* ray film.

VACUOLATING ASSAY

H pylori strains were cultured at 37 °C in brucella broth containing 5% FBS under micro-aerobic conditions for 48 hours. The culture medium was centrifuged at 10 000 *g* for 20 min, and cell-free supernatant was sterilised by passage through a 0.2 µm pore size filter and

Table 1 *Helicobacter pylori* strains used in this study

Strain* no.	Protein secretion		Vacuolating assay	Type†
	VacA	CagA		
Gastric ulcer				
1 (T-1)	+	+	+	I
2 (T-6)	+	+	+	I
3 (T-24)	+	+	+	I
4 (T-39)	+	+	+	I
5 (T-72)	+	+	+	I
6 (T-84)	+	+	+	I
7 (T-97)	+	+	+	I
8 (T-23)	+	+	+	I
9 (T-59)	+	+	+	I
10 (T-78)	+	+	+	I
11 (T-99)	+	+	+	I
12 (T-48)	-	+	+	I
13 (T-87)	-	+	+	I
14 (T-79)	+	+	-	Intermediate
15 (T-98)	+	+	-	Intermediate
16 (T-57)	+	-	-	Intermediate
Duodenal ulcer				
17 (T-10)	+	+	+	I
18 (T-29)	+	+	+	I
19 (T-54)	+	+	+	I
20 (T-56)	+	+	+	I
21 (T-60)	+	+	+	I
22 (T-71)	+	+	+	I
23 (T-74)	+	+	+	I
24 (T-92)	+	+	+	I
25 (T-101)	+	+	+	I
26 (T-9)	+	+	+	I
27 (T-13)	+	+	+	I
28 (T-35)	+	+	+	I
29 (T-63)	+	+	+	I
30 (T-82)	+	+	+	I
31 (T-12)	-	-	+	Intermediate
32 (T-64)	+	-	-	Intermediate
33 (T-15)	-	+	-	Intermediate
Non-ulcer dyspepsia				
34 (T-5)	+	+	+	I
35 (T-11)	+	+	+	I
36 (T-17)	+	+	+	I
37 (T-61)	+	+	+	I
38 (T-75)	+	+	+	I
39 (T-76)	+	+	+	I
40 (T-77)	+	+	+	I
41 (T-80)	+	+	+	I
42 (T-89)	+	+	+	I
43 (T-90)	+	+	+	I
44 (T-111)	+	+	+	I
45 (T-16)	+	+	+	I
46 (T-73)	+	+	+	I
47 (T-88)	+	+	+	I
48 (T-93)	+	+	+	I
49 (T-100)	+	+	+	I
50 (T-102)	+	+	+	I
51 (T-103)	+	+	+	I
52 (T-94)	+	-	+	Intermediate
53 (T-53)	+	-	+	Intermediate
54 (T-47)	+	+	-	Intermediate
55 (T-3)	-	+	-	Intermediate
56 (T-25)	-	-	-	II
57 (T-85)	-	-	-	II
58 (T-68)	-	-	-	II
Gastric cancer				
59 (T-7)	+	+	+	I
60 (T-115)	+	+	+	I
61 (T-116)	+	+	+	I
62 (T-22)	+	+	+	I
63 (T-26)	+	+	+	I
64 (T-81)	+	+	+	I
65 (T-112)	+	+	+	I
66 (T-113)	+	+	+	I
67 (T-114)	+	+	+	I
68 (T-108)	-	+	-	Intermediate
Controls				
69 (43526)	+	+	+	
70 (Tx30a)	+	-	-	

*The strains whose designation starts with a T were isolated at Tokyo University Hospital. 43526 was ATCC 43526 from American Type Culture Collection and Tx30a was from Texas.

†Type I strains were positive for vacuolating activity and CagA protein. Type II strains were negative for both, and intermediate strains belonged to neither Type I nor Type II.

concentrated 20-fold by using a 30 kDa ultra-filtration membrane (Amicon, Lexington, MA, USA), then stored at -80°C until use for the vacuolating assay. HeLa cells were cultured in plastic flasks in Dulbeccos' modified Eagle's

medium containing 10% FBS at 37°C in 5% CO_2 atmosphere. Confluent cell cultures were treated with trypsin and adjusted to a concentration of 1×10^4 cells per ml. At 24 hours before the addition of culture supernatant, 100 μl of cells were seeded into each well of a 96-well microtitre plate. A 10 μl sample of concentrated supernatant obtained from the culture medium was incubated with the HeLa cells for 24 hours. The result of the vacuolating assay was defined as positive when vacuolation was visible in more than 50% of HeLa cells by light microscopy.

TYPING OF *VAC*A GENE BY POLYMERASE CHAIN REACTION (PCR)

The *vacA* gene encodes a precursor protein made up of amino-terminal signal sequence, the cytotoxin proper, and a membrane protein which is removed during transmembrane transport. Genotyping of *H. pylori* was proposed on the basis of the nucleotide sequence in the signal sequence (types s1a, s1b, and s2) and that in the cytotoxin region (m1, m2). We amplified the *vacA* gene by PCR using previously described genotype-specific primers.²⁷ Thermal cycling was at 94°C for 30 seconds, 50°C for 15 seconds, and 72°C for one minute, for a total of 35 cycles. PCR products were analysed by 1% agarose gel electrophoresis with ethidium bromide staining. ATCC43526 was used as a s1a/m1 control, and Tx30a was used as a s2/m2 control.

STATISTICAL ANALYSIS

Fisher's exact probability test and χ^2 test were used to test the independence on contingency tables. $p < 0.05$ was considered to be statistically significant.

Results

DETECTION OF VACA PROTEIN BY IMMUNOBLOT

VacA protein was detected among the cell surface proteins obtained from 59 of 68 (87%) *H. pylori* isolates (fig 1A, tables 1 and 2). The positive rate was 14/16 (88%) in the isolates from patients with gastric ulcer, 15/17 (88%) from those with duodenal ulcer, 21/25 (84%) from those with non-ulcer dyspepsia, and 9/10 (90%) from gastric cancer patients. Statistically, the positive rate of VacA protein was not dependent on the gastroduodenal disease of the host (table 2).

The immunoblot analysis using whole cell lysates detected VacA protein even in the isolates that were negative by immunoblot analysis using cell surface proteins, suggesting that VacA protein was produced by all isolates but its secretion was disturbed in some of them.

VACUOLATING ASSAY

The vacuolating assay was positive in 54 of 59 (92%) VacA positive isolates and three of nine (33%) VacA negative ones (tables 1 and 2). Although discordant results were found in 8 of 68 (12%) isolates, the correlation between the two assays was strong enough ($p < 0.001$) to assume that most isolates positive for VacA

Table 2 Vacuolating activity and expression of VacA and CagA protein in clinical isolates

Diagnosis	Vacuolating activity		
	(%)	VacA (%)	CagA (%)
Gastric ulcer (n = 16)	13 (81)	14 (88)	15 (94)
Duodenal ulcer (n = 17)	15 (88)	15 (88)	15 (88)
Non-ulcer dyspepsia (n = 25)	20 (80)	21 (84)	21 (84)
Gastric cancer (n = 10)	9 (90)	9 (90)	10 (100)
Total (n = 68)	57 (84)	59 (87)	61 (90)

Table 3 Typing of vacA gene

	m1	m2	Neither	Total
s1a	5	2	38	45
s1b, s2	0	0	0	0
Total	5	2	38	45

protein by the immunoblot analysis did secrete active cytotoxin.

Five strains were positive for VacA protein by immunoblot but negative for the vacuolating assay. These strains may be secretors of inactive cytotoxin similar to the Tx30a strain. Another three isolates were positive for the vacuolating assay but negative by the immunoblot analysis. The reason for this discrepancy is not known, but it may be that substances other than VacA induced vacuolation.

GENOTYPE OF *VAC A* GENE

The genotype of the *vacA* signal sequence was determined in 45 isolates, including six that were negative for VacA by both immunoblot and vacuolating assay. Interestingly, the signal sequence in all isolates was of type s1a, which is supposed to represent high vacuolating activity.²⁷ The genotype of the mid region was also investigated in 45 isolates, as well as type m1 in five (11%) and type m2 in two (4%). PCR failed to amplify the mid region in the remaining 38 (84%) isolates (table 3).

DETECTION OF *CAG A* PROTEIN BY IMMUNOBLOT

The immunoblot analysis detected CagA protein in 61 of 68 (90%) *H pylori* isolates (fig 1B, tables 1 and 2). The positive rate was 15/16 (94%) in the isolates from patients with gastric ulcer, 15/17 (88%) from those with duodenal ulcer, 21/25 (84%) from those with non-ulcer dyspepsia, and 10/10 (100%) from gastric cancer patients. As in the case of the VacA protein described above, the positive rate of expression of CagA protein was high irrespective of the gastroduodenal disease of the host (table 2).

The estimated molecular mass of the CagA protein visualised by immunoblot varied from 120 to 140 kDa. This may indicate its heterogeneity among *H pylori* isolates, although we could find no correlation between its molecular mass and the gastroduodenal status of the host.

CORRELATION BETWEEN *VAC A* AND *CAG A*

The expression of CagA protein correlated strongly with that of VacA, as determined by immunoblot ($p = 0.005$), and with vacuolating cytotoxin activity in vitro ($p = 0.002$). Overall, 54 of 68 (79%) isolates were positive for cytotoxin activity and CagA protein. These isolates are defined as Type I strains in the following

text. Type II strains were negative for these assays, and intermediate strains were neither Type I nor Type II.

The prevalence of Type I strains was 14/17 (82%) in the isolates from patients with duodenal ulcer, 13/16 (81%) from those with gastric ulcer, 18/25 (72%) from those with non-ulcer dyspepsia, and 9/10 (90%) from gastric cancer patients. The prevalence did not differ significantly between the groups.

Three isolates were negative for VacA protein, vacuolating activity, and CagA protein (Type II). The prevalence of these all-negative isolates was 3/25 (12%) in patients with non-ulcer dyspepsia and 0/43 (0%) in patients with peptic ulcer or gastric cancer. This difference was statistically significant ($p = 0.046$).

Discussion

Our current study shows that most *H pylori* strains isolated in Japan were positive for vacuolating cytotoxin and CagA protein. Indeed, 54/68 (79%) isolates were positive for the vacuolating assay and CagA protein and were therefore Type I strains. These results agree with our previous findings that the seropositivity of anti-VacA and anti-CagA antibodies is very high in Japanese patients with *H pylori* infection. We can conclude that most *H pylori* strains in Japan produce both VacA and CagA proteins not only in vitro but also in vivo.

Weel *et al*²⁴ reported that infection with Type I strains was found in 43/76 (57%) patients with peptic ulcers, but in only 28/76 (37%) patients with non-ulcer dyspepsia in the Netherlands. In contrast, the prevalence of infection with Type I strains was 25/33 (76%) in patients with peptic ulcers and 18/25 (72%) in those with non-ulcer dyspepsia in Japan. Compared with previous reports from Western countries, Type I infection is very common in Japan irrespective of gastroduodenal status of the host. Since there was no significant relation between a particular gastroduodenal disease in the host and the expression of VacA or CagA protein by the isolate, the virulence factors VacA and CagA cannot be used as a discriminator of gastroduodenal diseases in the host.

Recent studies on the *cagA* gene in Japan, Korea, and China, have shown that its prevalence is very high and not related to particular gastroduodenal diseases.²⁹⁻³¹ For example, Pan *et al*²⁹ reported that 47 of 48 (98%) strains and 35 of 35 (100%) isolates were positive for the *cagA* gene in Chinese patients with peptic ulcers and gastritis respectively.

However, the current study does not rule out an association between the expression of VacA or CagA protein and the virulence of *H pylori*. The incidence of gastric cancer is high in Japan, with 38.2 deaths in 100 000 people in 1993.³² Epidemiological studies have shown that the prevalence of *H pylori* infection is significantly higher in patients with gastric cancer than in the general population in Japan as well as in other countries.^{4 5 33} It is possible that the high prevalence of infection with Type I strains shown in this study is associated with the high incidence of gastric cancer in Japan. In the current study, no patients with gastric cancer were

infected with VacA and CagA negative strains, suggesting a possibility that the incidence of gastric cancer may be low in the regions where infection with Type I strains is rare. It would require worldwide surveillance to elucidate whether Type I infection is actually related to gastric carcinogenesis. Recombinant proteins may be used in such studies for the examination of serum specific antibodies.

The vacuolating assay and analysis of VacA protein expression produced slightly different results. The difference between vacuolating activity and expression of VacA protein suggests that some strains secreted either inactive cytotoxin or only a small amount of active cytotoxin, so that the cytotoxin was detected by immunoblot analysis but its activity was not detected by the vacuolating assay. Indeed, some strains such as Tx30a secrete inactive cytotoxin,²⁷ so that no vacuolation was observed by the vacuolating assay while VacA protein was detected by immunoblot analysis. Three isolates (T-12, T-47, T-87) were positive in the vacuolating assay but negative by immunoblot analysis. It is possible that substances other than VacA induced the vacuolation, or that VacA protein was decomposed. All isolates that were negative by immunoblot analysis of surface proteins showed VacA protein by immunoblot using cell lysates. These results suggest that VacA protein was produced, at least in bacterial cells.

The genotyping of the *vacA* gene showed genetic diversities between *H. pylori* isolates from Japan and those from Western countries. The signal sequence of the *vacA* gene was of the s1 type (s1a subtypes) in all isolates examined in this study, and the mid region of the gene was of neither the m1 or m2 type in 38/45 (84%) isolates. A recent study reported similar results on the signal sequence in Japanese isolates.³⁴ These results were substantially different from the results obtained in Western countries. For example, Atherton *et al.*²⁷ reported that 21/23 (91%) isolates from patients with peptic ulcer but only 16/33 (48%) isolates from patients without ulcer were of the s1 type. They also reported that the mid region was either of the m1 or m2 subtype in 56 isolates, and no other subtypes were found. The distinct genotypes of the *vacA* gene of isolates in Japan may be associated with functional differences of the VacA protein, and may be a reason for the high incidence of gastric cancer in Japan.

CagA protein was reported to be produced in most cytotoxin positive strains of *H. pylori*.^{16, 17} We investigated the association between CagA protein expression and vacuolating activity in isolates in Japan. CagA protein expression and vacuolating activity were both present in 54 of 68 strains, and both absent in five of 68 strains. This result indicates that CagA protein is strongly associated with vacuolating cytotoxin activity in Japan as well as in other countries.^{16, 17}

The function of CagA protein is not understood at all. CagA has been thought to be associated with IL-8 induction in the gastric epithelium, and bacterial eradication reduces IL-8 expression and neutrophil infiltration.²³

However, recent studies have shown that the IL-8 induction may not be directly due to CagA protein, but to the proteins encoded by the genes in the pathogenicity island upstream of the *cagA* gene. These proteins may be co-expressed with CagA and therefore CagA may be used as a marker of enhanced virulence.^{35, 36}

In conclusion, we examined the expression of VacA and CagA proteins by *H. pylori* strains clinically isolated in Japan. The prevalence of infection with strains that express vacuolating cytotoxin and CagA was very high compared with previous reports from Western countries. No significant correlation was found between the expression of these proteins by the isolates and gastroduodenal status of the host. Thus these virulence factors cannot be used as markers to discern the risk of developing serious gastroduodenal disease in the host in Japan. However, it is possible that the high prevalence of infection with these Type I strains contributes to the characteristics of *H. pylori* infection in Japan.

This work was supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science, and Culture of Japan. F.K is a Research Fellow of the Japan Society for the Promotion of Science.

- Warren JR, Marshall BJ. Unidentified curved bacilli on gastric epithelium in active chronic gastritis. *Lancet* 1983;1:1273-5.
- NIH consensus conference. Helicobacter pylori in peptic ulcer disease. NIH Consensus Development Panel on Helicobacter pylori in Peptic Ulcer Disease. *JAMA* 1994;272:65-9.
- Graham DY, Lew GM, Klein PD, *et al.* Effect of treatment of Helicobacter pylori infection on the long-term recurrence of gastric or duodenal ulcer: a randomized, controlled study. *Ann Intern Med* 1992;116:705-8.
- Parsonnet J, Friedman GD, Vandersteen DP, *et al.* Helicobacter pylori infection and the risk of gastric carcinoma. *N Engl J Med* 1991;325:1127-31.
- Nomura A, Stemmermann GN, Chyou PH, *et al.* Helicobacter pylori infection and gastric carcinoma among Japanese American in Hawaii. *N Engl J Med* 1991;325:1132-6.
- Asaka M, Kimura T, Kudo M, *et al.* Relationship of Helicobacter pylori to serum pepsinogens in an asymptomatic Japanese population. *Gastroenterology* 1992;102:760-6.
- Figura N, Guglielmetti P, Rossolini A, *et al.* Cytotoxin production by Campylobacter pylori strains isolated from patients with peptic ulcers and from patients with chronic gastritis only. *J Clin Microbiol* 1989;27:225-6.
- Fox JG, Correa P, Taylor NS, *et al.* High prevalence and persistence of cytotoxin-positive Helicobacter pylori strains in a population with high prevalence of atrophic gastritis. *Am J Gastroenterol* 1992;87:1554-60.
- Leunk RD. Production of a cytotoxin by Helicobacter pylori. *Rev Infect Dis* 1991;13:S686-9.
- Cover TL, Dooley CP, Blaser MJ. Characterization of and human serologic response to proteins in Helicobacter pylori broth culture supernatants with vacuolating cytotoxin activity. *Infect Immun* 1990;58:603-10.
- Tee W, Lambert JR, Dwyer B. Cytotoxin production by Helicobacter pylori from patients with upper gastrointestinal tract diseases. *J Clin Microbiol* 1995;33:1203-5.
- Cover TL, Blaser MJ. Purification and characterization of the vacuolating toxin from Helicobacter pylori. *J Biol Chem* 1992;267:10570-5.
- Phadnis SH, Ilver D, Weelzon L, *et al.* Pathological significance and molecular characterization of the vacuolating toxin gene of Helicobacter pylori. *Infect Immun* 1994;62:1557-65.
- Schmitt W, Haas R. Genetic analysis of the Helicobacter pylori vacuolating cytotoxin: structural similarities with the IgA protease type of exported protein. *Mol Microbiol* 1994;12:307-19.
- Telford JL, Ghiara P, Dell'Orco M, *et al.* Gene structure of the Helicobacter pylori cytotoxin and evidence of its key role in gastric disease. *J Exp Med* 1994;179:1653-8.
- Covacci A, Censini S, Bugnoli M, *et al.* Molecular characterization of the 128-kDa immunodominant antigen of Helicobacter pylori associated with cytotoxicity and duodenal ulcer. *Proc Natl Acad Sci USA* 1993;90:5791-5.
- Tummuru MKR, Cover TL, Blaser MJ. Cloning and expression of a high-molecular-mass major antigen of Helicobacter pylori: evidence of linkage to cytotoxin production. *Infect Immun* 1993;61:1799-809.

- 18 Crabtree JE, Figura N, Taylor JD, *et al.* Expression of 120 kilodalton protein and cytotoxicity in *Helicobacter pylori*. *J Clin Pathol* 1992;45:733-4.
- 19 Xiang Z, Censini S, Bayeli PF, *et al.* Analysis of expression of CagA and VacA virulence factors in 43 strains of *Helicobacter pylori* reveals that clinical isolates can be divided into two major types and that CagA is not necessary for expression of the vacuolating cytotoxin. *Infect Immun* 1995;63:94-8.
- 20 Blaser MJ, Perez-Perez GI, Cover TL, *et al.* Infection with *Helicobacter pylori* strains possessing CagA is associated with an increased risk of developing adenocarcinoma of stomach. *Cancer Res* 1995;55:2111-15.
- 21 Cover TL, Glupczynski Y, Lage AP, *et al.* Serologic detection of infection with *cagA+* *Helicobacter pylori* strains. *J Clin Microbiol* 1995;33:1496-500.
- 22 Ching CK, Wong BCY, Kwok E, *et al.* Prevalence of CagA-bearing *Helicobacter pylori* strains detected by the anti-CagA assay in patients with peptic ulcer disease and in controls. *Am J Gastroenterol* 1996;91:949-53.
- 23 Crabtree JE, Farmery SM, Lindley IKD, *et al.* CagA/cytotoxic strains of *Helicobacter pylori* and interleukin-8 in gastric epithelial cell lines. *J Clin Pathol* 1994;47:945-50.
- 24 Weel JF, van der Hulst RW, Gerrits Y, *et al.* The interrelationship between cytotoxin-associated gene A, vacuolating cytotoxin, and *Helicobacter pylori*-related diseases. *J Infect Dis* 1996;173:1171-5.
- 25 Ogura K, Kanai F, Maeda S, *et al.* High prevalence of cytotoxin positive *Helicobacter pylori* in patients irrelevant to the presence of peptic ulcers in Japan. *Gut* 1997;41:463-8.
- 26 Maeda S, Kanai F, Ogura K, *et al.* High seropositivity of anti-CagA antibody in *Helicobacter pylori* infected patients irrelevant to the presence of peptic ulcers and gastric cancer in Japan. *Dig Dis Sci* 1997;42:1841-7.
- 27 Atherton JC, Cao P, Peek RJ, *et al.* Mosaicism in vacuolating cytotoxin alleles of *Helicobacter pylori*. Association of specific *vacA* types with cytotoxin production and peptic ulceration. *J Biol Chem* 1995;270:17771-7.
- 28 Laemmli UK. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 1970;227:680.
- 29 Pan ZJ, van der Hulst RWM, Feller M, *et al.* Equally high prevalences of infection with *cagA*-positive *Helicobacter pylori* in Chinese patients with peptic ulcer disease and those with chronic gastritis-associated dyspepsia. *J Clin Microbiol* 1997;35:1344-7.
- 30 Miehke S, Kibler K, Kim JG, *et al.* Allelic variation in the *cagA* gene of *Helicobacter pylori* obtained from Korea compared to the United States. *Am J Gastroenterol* 1996;91:1322-5.
- 31 Shimoyama T, Fukuda S, Tanaka M, *et al.* High prevalence of the CagA-positive *Helicobacter pylori* strains in Japanese asymptomatic patients and gastric cancer patients. *Scand J Gastroenterol* 1997;32:465-8.
- 32 Health and Welfare Statistics Office. Table on mortality. *Journal of Health and Welfare Statistics* 1995;42:404.
- 33 Asaka M, Kimura T, Kato M, *et al.* Possible role of *Helicobacter pylori* infection in early gastric cancer development. *Cancer* 1994;73:2691-4.
- 34 Ito Y, Azuma T, Ito S, *et al.* Analysis and typing of the *vacA* gene from *cagA*-positive strains of *Helicobacter pylori* isolated in Japan. *J Clin Microbiol* 1997;35:1710-14.
- 35 Tummuru MKR, Sharma SA, Blaser MJ. *Helicobacter pylori* picB, a homologue of the *Bordetella pertussis* toxin secretion protein, is required for induction of IL-8 in gastric epithelial cells. *Mol Microbiol* 1995;18:867-76.
- 36 Censini S, Lange C, Xiang Z, *et al.* *cag*, a pathogenicity island of *Helicobacter pylori*, encodes type I-specific and disease-associated virulence factors. *Proc Natl Acad Sci USA* 1996;93:14648-53.