Review

Cytokine regulation of epithelial permeability and ion transport

Introduction

The intestinal epithelium serves as a dynamic barrier which, in the course of its normal function, maintains regulated uptake of nutrients and water at the same time as excluding potential pathogens. Enteropathies, including inflammatory bowel disease (IBD) result in, or perhaps even from, perturbed epithelial function. Despite the idio-pathic nature of IBD, it is apparent that much of the pathophysiology, tissue damage and symptomatology of these disorders are due to inappropriate or exaggerated immune reactions. Cytokines are, inter alia, regulators of both innate and acquired immunity; so therefore it is not surprising that exaggerated effects of cytokines in acute or chronic inflammatory states may be due to unregulated production of pro-inflammatory cytokines or inadequate synthesis of anti-inflammatory cytokines. Indeed, increases in cytokine concentrations in models of enteric inflammation or IBD are well documented. Here, we provide an overview of cytokine regulation of epithelial ion transport and permeability, highlighting, where appropriate, the potential of the epithelium to modulate mucosal immune reactions and its own function.

Relatively few studies have demonstrated cytokine effects on epithelial function in vivo or by examining intestinal tissue responses ex vivo. Several features contribute to difficulties with interpretation of such studies. Firstly, cytokines act in a coordinated interplay of often redundant, pleiotropic and occasionally opposing actions. Secondly, given the complex intercellular signalling that occurs within the gut, cytokine effects may involve other, non-cytokine mediators. Finally, virtually every putative mediator of inflammation has been implicated in diseases of the intestine, representing at one time the academic pharmacologist’s dream and the therapist’s nightmare.

The advent of epithelial cell lines, principally the tumour derived T84, Caco-2 and HT-29 cell lines, has offered opportunities to study influences of cytokines on epithelial function by taking a reductionist approach. These cell lines, when grown on permeable filters, form polarised, high electrical resistance monolayers (rodent derived epithelial cells have been extensively used to examine epithelial restitution, cytokine production and other parameters. However, these cells do not generally form monolayers suitable for studies in Ussing chambers). Although there are several disadvantages inherent with using cell lines11 such investigations have made a significant contribution to the awareness of cytokine regulation of epithelial physiology in general, and ion transport and permeability in particular. Moreover, cell lines are now a basic tool in cell biology, allowing both generation and testing of novel hypotheses which may unveil potential therapeutic targets which require subsequent validation by whole tissue or animal studies in advance of experimental clinical manipulation(s).

The enteric epithelial cell layer regulates normal homoeostasis, innate immunity and regulation of acquired immunity at both sensory and effector levels.12-14 As such, the epithelium may play a role in the immunological outcome of luminal exposure to antigens resulting in tolerance, protective immunity or hypersensitivity reactions. There exists a substantial literature on the capacity of epithelial cells to produce bioactive cytokines and other immunoregulatory molecules either constitutively or in response to stimuli including other cytokines or microorganisms.14-15 The epithelium is now identified as both target and source of cytokines (fig 1). Epithelial derived cytokines have the potential to play an autocrine role or to influence adjacent non-epithelial cells. For example, epithelial cells can synthesise interleukin (IL) 6, a cytokine that can inhibit lipopolysaccharide (LPS) induced IL-1 and tumour necrosis factor (TNF) α in vivo and in vitro. IL-6 is also a major inducer of tissue inhibitor of matrix metalloproteinase (TIMP) and here it is noteworthy that much of the tissue damage associated with T cells is due to the release of metalloproteinases.16

Epithelial permeability

Intestinal permeability is a rather loose but clinically useful term used to denote integrity of the epithelial barrier in vivo which may be compromised in malnutrition,21 infection22 23 and in pathological conditions including IBD and coeliac disease.24 In vitro, permeability to macromolecules is readily determined quantitatively25 and can be used to study modulation of epithelial permeability by extracellular macromolecules including cytokines. Trans-epithelial electrical resistance (TER; and its inverse — conductance) is determined by the Ohmic relation and has been used as an index of barrier permeability. However, there is not always a direct correlation between electrical resistance and permeability to marker molecules.26

Luminal material crosses the epithelial sheet via the transcellular or paracellular pathways. Tight junctions form a circumferential seal at the luminal pole of adjacent epithelial cells and are the rate limiting factor regulating paracellular permeability.27 A growing body of evidence has recently accumulated showing that such junctions express a high degree of plasticity and may be greatly attenuated in disease states.28 Most findings to date implicate pro-inflammatory cytokines as agents which reduce barrier integrity.

EXOGENOUS CYTOKINES

Direct actions of cytokines may readily be examined pharmacologically in model epithelia. Addition of interferon (IFN) γ, but not IFN-α, to the basolateral surface of T84 monolayers causes a significant increase in paracellular

Abbreviations used in this paper: IBD, inflammatory bowel disease; IL, interleukin; LPS, lipopolysaccharide; TNF, tumour necrosis factor; TIMP, tissue inhibitor of matrix metalloproteinase; TER, transepithelial electrical resistance; IFN, interferon; TGF, transforming growth factor; PMBC, peripheral blood mononuclear cells; SEB, Staphylococcus enterotoxin B; PGE, fibroblast growth factor; EGF, epidermal growth factor; IGF, insulin-like growth factor; IEL, human intraepithelial lymphocytes; PG, prostaglandin; CFTR, cystic fibrosis transmembrane regulator; MLCK, myosin light chain kinase; NF-κB, nuclear factor κB; STAT, signal transduction and activators of transcription; MALT, mucosa associated lymphoid tissue.
IFN-γ and IL-13, which have broadly similar biological functions, both cause increased epithelial permeability. However, unlike IL-4, IL-13 does not affect ion transport in T84 cells. These divergent effects of IL-4 and IL-13 could be due to interaction with different receptors. Another possibility is that both cytokines may bind to the same receptor (IL-4 and IL-13 receptors have a structural homologous subunit), with IL-13 triggering only part of the signal transduction cascade that is inducible by IL-4.

TNF-α concentrations are increased in a variety of inflammatory disorders and the use of TNF-α neutralising antibody has been found to be an effective therapeutic modality in some patients with Crohn’s disease. However, anti-TNF-α immunotherapy clinical trials have not examined changes in intestinal permeability before and after treatment. TNF-α increases endothelial permeability and studies with a variety of epithelia, including the renal cell line LLC-PK1 (in which protein kinase C was implicated in the TNF-α evoked increase in permeability) suggest that epithelial barrier function is reduced by TNF-α. However, observations from four independent laboratories have shown that low dose TNF-α (<10 ng/ml) has no effect on T84 permeability, whereas another report indicates that TNF-α can potentiates the drop in T84 transepithelial resistance induced by IFN-γ. Additionally, chronic TNF-α treatment of T84 cells has been found to increase permeability (100 ng/ml/day for four days) induced by the immune cell are, at least in principle, better understood than the TNF-α induced increases in T84 epithelial responses. For example, supernatants from antigen stimulated PBMC from children allergic to cow’s milk decrease the TER of HT-29 monolayers. Given the rapidity of this response, which was maximal within 24 hours, it seems unlikely that it is due to the singular activity of any of the cytokines considered earlier which require longer exposure to generate effects. Indeed, it is rather unlikely that epithelia in situ would be exposed to a single cytokine. Exposure of model epithelia to co-cultured immunocytes or immune cell derived conditioned media which contain mixtures of cytokines in the relative proportions in which they are produced by the immune cell are, at least in principle, better understood than the TNF-α induced increases in T84 epithelial responses.

CELL DERIVED CYTOKINES

In reconstructing more complex systems, co-culture studies where epithelial monolayers and immune cells or immunocyte derived supernatants are juxtaposed results in disruption of epithelial barrier function. T84 cell monolayers co-cultured with mucosal-like lymphocytes, anti-CD3 or SEB activated PBMC, endotoxin activated monocytes or human intraepithelial lymphocytes (IEL) or an IEL cell line invariably display a decrease in transepithelial resistance within 24 hours of co-culture. Although cell–cell contact may be a requirement for full expression of such intercellular cross talk, there is evidence to suggest that paracrine signalling occurs as quantitatively similar increases in epithelial permeability are elicited by conditioned media from the immune cell cultures. For example, supernatants from antigen stimulated PBMC from children allergic to cow’s milk decrease the TER of HT-29 monolayers. Given the rapidity of this response, which was maximal within 24 hours, it seems unlikely that it is due to the singular activity of any of the cytokines considered earlier which require longer exposure to generate effects. Indeed, it is rather unlikely that epithelia in situ would be exposed to a single cytokine. Exposure of model epithelia to co-cultured immunocytes or immune cell derived conditioned media which contain mixtures of cytokines in the relative proportions in which they are produced by the immune cell are, at least in principle, better understood than the TNF-α induced increases in T84 epithelial responses.
mirrors of native mucosa than using single cytokines to elicit biological responses. We suggest that immune mediated alterations in epithelial permeability are due to the combinatorial effects of multiple mediators, including some which may yet have to be identified, where the stoichiometry and sequence of mediator release may be crucial. Several lines of evidence support these arguments. For instance, in HT-29 monolayers, the increased paracellular permeability and altered tight junction structure induced by TNF-α was enhanced by co-treatment with low dose IFN-γ. Also pharmacological blockade of the effects of single cytokines failed to prevent significantly the actions of IEL supernatant. Finally, IELs produce at least two molecules that differentially modulate epithelial permeability, a factor(s) <30 kDa that causes a rapid (within four hours) 30% drop in resistance and a larger molecule(s) (>30 kDa), the effect of which was slower in onset and caused a dramatic loss of epithelial barrier function (reduced by ~90%).

Molecules other than cytokines may contribute to signalling cascades which result in altered epithelial permeability. For instance, tissue and cell culture studies have shown that nitric oxide can regulate epithelial permeability and ion transport. Evidence has been presented implicating nitric oxide as the final mediator in IFN-γ evoked disruption of epithelial barrier function. This raises the issue of active involvement of the epithelium, and indeed the effects of IFN-γ are only observed after more than 24 hours of culture and are protein synthesis dependent. Also, 5-ASA significantly reduced the IFN-γ induced increased transepithelial flux of mannitol (a marker of paracellular permeability) across HT-29-19A monolayers. The authors suggest that this effect was due to the 5-aminosalicylic acid competing for the IFN-γ receptor, but the disruption of arachidonic acid metabolism may also contribute. TNF-α and bacterial infection can result in increased epithelial arachidonic acid metabolism and prostaglandin synthesis, begging the question whether epithelial derived lipid metabolites exert autocrine effects on epithelial permeability and ion secretion? In fact, bacterial infection of HT-29 cells evoked epithelial prostaglandin (PG) E2 and PGF2α production, and transfer of conditioned media from the infected cells to naive epithelial monolayers evoked a transient increase in electrogenic electrolyte transport similar to that caused by PGE2.

Transcytosis and antigen presentation
Cytokine modulation of transcytosis by intestinal epithelial cells is a likely event, consistent with a role for cytokines and epithelial cells in the sensory arm of adaptive immunity. Soluble factors and immune cell–epithelial cell contact are required for the formation of M cells that overlay the Peyer’s patches, creating a portal of entry for bacteria and particulate antigens. M cell density is increased in vivo in response to exposure to bacteria, perhaps as a consequence of local generation of cytokines induced by bacterial infection which, although they interact with epithelial cells in different ways, seem to generate a rather conserved set of responses including cytokine generation. Further support for cytokine participation in antigen uptake, presentation and processing comes from observations that IFN-γ, and to a lesser extent TNF-α, enhances HLA-DR expression in T84 cells, increasing the potential of such cells, in vivo, to play a role in antigen processing and that inactivation of genes in the TNF-lymphotoxin ligand and receptor system prevents normal organisation of intestinal Peyer’s patches.

Intact antigen may also cross enterocytes, which are more abundant than M cells. Regulation of transepithelial permeability by absorption enhancers which have been investigated for increasing oral bioavailability of drugs may result in delivery of antigenic material (e.g. gut microflora or indeed the drug itself) to the antigen processing compartments of the lamina propria. Inappropriate entry into the mucosal compartment of otherwise contained luminal entry could, at least in principle, lead to other pathology as a consequence of altered permeability and/or immune status in IBD. Reports of associated conditions range from arthropathies to autism. Intestinal epithelial cells seem to satisfy most of the criteria by which they may be considered as antigen presenting cells.

Epithelial ion transport
Transepithelial vectorial electrolyte movement creates driving forces for directed water movement and results from the coordinated activity of ion channels, transporters and pumps, which have polar distributions in the cell membrane. Extremes of electrogenic ion transport can result in malabsorptive or secretory diarrhoea or constipation, often in response to extracellular molecules which influence epithelial cell activity.

Electrophysiological studies using single (epithelial) cell types tend to show few rapid or direct effects of cytokines on electrogenic ion transport. IFN-γ and IL-4 have no direct effect on baseline ion transport in the T84 model of CI secretion, but both dramatically reduce the ability of the cells to secrete Cl– vectorially in response to a range of directly acting secretagogues. Early studies correlated changes in ion transport characteristics evoked by IFN-γ with changes in membrane proteins and more recently a reduction in the cystic fibrosis transmembrane regulator has been reported. Co-culture of confluent T84 monolayers with mucosal-like lymphocytes or anti-CD3 or SEB activated immune cells results in reduced secretory responsiveness. T84 monolayers exposed to conditioned media from activated immune cells also display diminished responses to secretory stimuli which are apparent six hours after exposure and are maximal by 18–24 hours after treatment. The low levels of IFN-γ in the conditioned media and the rapidity of the response implies that the dysregulated epithelial ion transport is not due solely to IFN-γ and is likely due to the effects of other mediators acting alone or in concert with IFN-γ.

Several pro-inflammatory cytokines are pro-secretory in intact, native mucosal preparations in vitro. For example, IL-1, IL-3, and TNF-α have been found to stimulate epithelial ion secretion. In each case, the effects of the cytokines were found to be indirect with the change in ion transport being mediated by prostaglandins from subepithelial cells. Similarly, co-culture studies using T84 epithelial cells revealed that ion transport events evoked by TNF-α and known secretagogues were modulated by the presence of myofibroblasts. Data have been reported which illustrate stimulation of T84 ion transport following 24–48 hours of co-culture with human monocytes activated with endotoxin and the bacterial tri-peptide, formyl-met-leu-phe. The response was accounted for by electrogenic chloride secretion and could be inhibited by neutralisation of TNF-α. The influence of TNF-α seems to be an autocrine action on the monocytes. Contrary effects have been found when endotoxin activated alveolar macrophages were co-cultured with lung epithelium; this resulted in reduced ion transport due to diminished Na+ movement. Patients receiving IL-2 immunotherapy can develop diarrhoea although it is not clear whether this is a direct action on epithelial cells as IL-2 does not directly
affect T84 barrier function. However, IL-2 receptors have been demonstrated on isolated epithelial cells and the rat IEC-6 cell line, upon which IL-2 has biphasic effects on proliferation rates.

Interleukin 10 exerts an antisecretory influence on gut epithelium. Thus, there is a drop in rat jejunal and ileal ion transport in response to IL-10, and the tissues exposed to IL-10 display diminished responses to carbachol (cholinergic agonist) and forskolin which evoke electrogenic Cl− secretion via Ca2+ and cAMP, respectively.

NF-κB can abrogate murine experimental colitis. Furthermore, the nature and magnitude of an immune response will be governed by the local cytokine environment.

**Signal transduction**

There is a general paucity of information relating specifically to signal transduction events and cytokine regulation of epithelial ion transport and permeability. Enteropathogenic *Escherichia coli* infection of T84 cell monolayers results in an increase in permeability and a concomitant increase in phosphorylation of the myosin light chain. This change in epithelial permeability was prevented by inhibition of myosin light chain kinase (MLCK) activity by ML-9 and increased permeability was elicited by retroviral transfection of the catalytic domain of MLCK into kidney epithelial cells. A role for MLCK has not been determined in immune modulation of epithelial permeability in the way it has been characterised in response to bacterial infection.

As therapeutic usefulness of cytokines or blocking antibodies may be limited by their pharmacokinetics, a more effective approach may be to direct therapy towards common signal transduction pathways used by or in the generation of cytokines. Gut epithelial cells infected with *Helicobacter pylori* or treated with IL-1β or TNF-α mobilise nuclear factor κB (NF-κB), which has been linked to IL-8 production. Increasing evidence implicates NF-κB as a critical transcription factor in chronic inflammatory disease and local administration of antisense oligonucleotides to NF-κB can abrogate murine experimental colitis.

Signal transduction and activators of transcription (STAT) proteins dimerise and are tyrosine phosphorylated by Janus kinases prior to dissociation from the receptor–ligand complex and translocation to the nucleus. Few studies have explored STAT protein expression in gut epithelial cells; a single report examined STAT3 activity in gut epithelial cells in response to IL-15. The hypothesis that NF-κB and/or STAT proteins govern changes in epithelial ion secretion and permeability remains largely untested.

**Regulation of mucosal inflammation and protection**

Cytokine induced epithelial expression of immune accessory molecules imparts upon the epithelium the capacity to influence the progress of inflammation. IFN-γ, TNF-α and other stimuli can upregulate cytokine and chemokine secretion, thus providing signals for recruitment and activation of neutrophils, monocytes and lymphocytes. Cytokine regulation of epithelial permeability may play a role in the pathogenesis of mucosal damage in IBD as abnormal and uncontrolled immune responses to microbial products and luminal antigens have been implicated in the induction of IL-10 display diminished responses to carbachol (cholinergic agonist) and forskolin which evoke electrogenic Cl− secretion via Ca2+ and cAMP, respectively.

Thus, in addition to an inhibitory effect on the cystic fibrosis transmembrane regulator (CFTR), it is tempting to speculate that IFN-γ effects on epithelial ion transport may be due, at least in part, to modulation of cytoskeletal elements that affect ion transport across the basolateral membrane or to an influence on intracellular signalling cascades.

Other studies of note include the demonstration of increased glucose and NaCl absorption in rabbit intestine in response to EGF, and EGF inhibition of Ca2+ mediated (in response to carbachol or thapsigargin) Cl− secretion in T84 cells via phosphatidylinositol-3-kinase. In contrast, rat alveolar epithelium treated with EGF displayed increased baseline Isc due to active Na+ transport that was sensitive to inhibition of tyrosine kinase activity.

**Figure 2** Impaired epithelial permeability, whether in response to cytokines or other stimuli, reduces barrier function. Increased exposure to luminal contents will activate lamina propria cells with consequent generation of pro-inflammatory cytokines. These may exacerbate the “leakiness” of the epithelium which can both promote and sustain inflammation.

**Figure 3** Cytokines and mucosal protection. The epithelium is involved in deployment of a number of effector mechanisms of innate and adaptive immunity. In terms of sensory function, antigen delivery requires trafficking (and perhaps processing) of antigens across the epithelial barrier. Locally active cytokines may influence antigen sampling in a number of ways. For example, by altering epithelial permeability or alternatively by enhancing M-cell differentiation. Furthermore, the nature and magnitude of an immune response will be governed by the local cytokine environment.
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Concluding comments

In this brief synopsis we have illustrated that cytokines directly and indirectly influence epithelial ion transport and permeability. What is perhaps most evident is the fact that with at least 30 known cytokines/growth factors, we have only begun to recognize the impact of cytokines on epithelial function. Indeed one of the major tasks facing enteric pharmacologists/physiologists is the dissection of pathways which transfer information and coordinate enteric responses to an extraordinary range of stimuli. Studies at subcellular, cellular and tissues levels are now possible to investigate how cytokines interact with each other and with non-cytokine neuroimmune mediators to regulate homeostasis in the intestine. Such knowledge will be essential for the rational development of new treatment regimens.

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