Microsatellite instability and the clinicopathological features of sporadic colorectal cancer

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Abstract

Background and aims—In this study, we prospectively examined the clinical significance of the microsatellite instability (MSI) phenotype in sporadic colorectal cancer, and investigated methods for effective identification of these tumours in routine pathology practice.

Methods—DNA was extracted from 310 tumours collected from 302 consecutive individuals undergoing curative surgery for sporadic colorectal cancer. Microsatellite status was determined by polymerase chain reaction amplification using standard markers, while immunostaining was used to examine expression of MLH1, MSH2, and p53.

Results—Eleven per cent of tumours showed high level instability (MSI-H), 6.8% had low level instability (MSI-L), and the remainder were stable. MSI-H tumours were significantly more likely to be of high histopathological grade, have a mucinous phenotype, and to harbour increased numbers of intraepithelial lymphocytes. They were also more likely to be right sided, occur in women, and be associated with improved overall survival. In total, 25 (8%) tumours showed loss of staining for MLH1 and a further three tumours showed absence of staining for MSH2. The positive and negative predictive value of immunohistochemistry in the detection of MSI-H tumours was greater than 95%.

Conclusions—We conclude that the MSI-H phenotype constitutes a pathologically and clinically distinct subtype of sporadic colorectal cancer. Immunohistochemical staining for MLH1 and MSH2 represents an inexpensive and accurate means of identifying such tumours.

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Keywords: colorectal carcinoma; microsatellite instability; survival; MLH1; MSH2; immunohistochemistry

The phenomenon of DNA microsatellite instability (MSI) results from failure of repair of DNA mismatches immediately after DNA replication,¹ and is characterised experimentally by contraction or expansion of mono-, di-, tri-, or tetranucleotide repeat sequences within DNA.² Studies of the rare syndrome of hereditary non-polyposis colorectal cancer (HNPCC) led to recognition of MSI as the phenotypic expression of a germline mutation in one or more of the genes associated with

DNA mismatch repair.³⁻⁵ At least five of these genes have now been identified including MSH2, MLH1, hPMS1, hPMS2, and hMSH6/GTBP.⁶⁻⁹ However, mutations in either MLH1 or MSH2 account for the majority of known germline mutations in HNPCC families.

Colorectal carcinomas arising in the setting of HNPCC, as well as showing high level MSI (MSI-H), have a number of distinctive clinicopathological features that separate them from the sporadic forms of the disease. Importantly, individuals with HNPCC associated tumours appear to have a significantly better outcome than those with sporadic disease of similar stage and grade.¹⁰⁻¹² They are also at greater risk of metachronous tumour development.

However, MSI is not confined to the setting of hereditary disease and may be seen in up to 15% of colorectal carcinomas in those with no family history of disease.¹³⁻¹⁵ In the case of these so-called sporadic colorectal tumours, mutation of MMR genes is relatively infrequent whereas biallelic hypermethylation of the promoter of MLH1 appears to be the most important mechanism for inactivation of mismatch repair genes.¹⁶⁻¹⁸

While it is well established that a proportion of sporadic colorectal cancers demonstrate an MSI-H phenotype, the clinical significance of such a finding has not yet been determined. Some studies have shown that individuals with MSI-H tumours have improved survival relative to those with MSS tumours of similar stage.^{4 19-23} Others have not demonstrated such a survival advantage,²⁴⁻²⁷ and one recent study suggested that the prognostic significance varies with site in the colon or rectum.²⁸ Recently, a number of small retrospective studies have also suggested that individuals with MSI-H sporadic tumours are at greater risk of metachronous cancers.^{22 23 29 30} Finally, there are indications that MSI-H tumours, either hereditary or sporadic, may be resistant to the actions of certain commonly used chemotherapeutic agents,³¹⁻³³ raising the important possibility that neoadjuvant or adjuvant chemotherapy may have different outcomes in this group of patients.

Much of the current confusion in the literature regarding the clinical significance of these tumours has arisen because studies to date have been retrospective, and in general have been limited to relatively small populations.

Abbreviations used in this paper: MSI, microsatellite instability; MSI-H, high level MSI; MSI-L, low level MSI; MSS, microsatellite stable; HNPCC, hereditary non-polyposis colorectal cancer; PCR, polymerase chain reaction; HR, hazard ratio.

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Furthermore, they have often been conducted in cancer dense families, or in selected subgroups of sporadic disease such as individuals with an early onset disease or with multiple primary tumours.^{21–23}

Adding to this confusion, it has recently been proposed that sporadic colorectal carcinoma can be categorised into three distinct groups, with either high (MSI-H), low (MSI-L), or absent (microsatellite stable, MSS) levels of microsatellite instability.¹⁵ In the case of MSI-L sporadic tumours, little is known of the prognostic implications or risk of metachronous tumour development.²³ Nevertheless, it has been proposed that the MSI-L phenotype reflects a distinctive pathway of tumour development that may be reflected in altered clinical behaviour.³⁴

In this study, we prospectively examined the clinical significance of these genetic subtypes of sporadic colorectal cancer and evaluated the optimal methods for identification of these tumours.

Materials and methods

PATIENTS AND SPECIMENS

After obtaining informed consent, 302 consecutive individuals undergoing surgical resection of adenocarcinoma of the colon or rectum at St Vincent's Hospital, Sydney, were enrolled in this prospective study from 1993 to 1998. Individuals were excluded where preoperative radiotherapy or chemotherapy had been administered. Patients with inflammatory bowel disease or with a known history of familial adenomatous polyposis or HNPCC were also excluded from this study, as were those individuals in whom the primary tumour was incompletely resected (R1 or R2).³⁵

Fresh representative tissue samples (500 µg) from all tumours and paired normal colonic mucosa were immediately frozen at -70°C . In total, 310 fresh tumour specimens were assayed from 172 males and 130 females (aged 29–95 years; mean 68.4 (12.4)). Fifty one of these tumours were TNM stage I while 113 were stage II, 97 were stage III, and 49 were stage IV.

Family histories of colorectal carcinoma and other malignancies were obtained by interviewing patients or their next of kin. Attempts were made to verify all suspected diagnoses of cancer and uncertain causes of death, either by obtaining death certificates and medical records or by contacting the treating physician. Family history was used to identify those families that met either the Amsterdam or modified Amsterdam criteria for HNPCC.³⁶

HISTOPATHOLOGICAL ANALYSIS OF TUMOURS

For all tumours, histopathological type, stage, and size were determined independently by a histopathologist within the Department of Anatomical Pathology, St Vincent's Hospital. Tumour grade, extent of mucin production, tumour growth pattern, as well as the presence of a Crohn's-like inflammatory infiltrate, intraepithelial lymphocytes, or peritumoural lymphocytes were determined prospectively

without knowledge of the mismatch repair status. Tumours in which less than 10% of cells formed glands were classified as high grade (poorly differentiated) while those containing more than 50% extracellular mucin were classified as mucinous.³⁷ The tumour growth pattern was interpreted as either infiltrative or expansile, according to previously published criteria.³⁸ The extent of peritumoural and Crohn's-like lymphoid reactions was classified according to the method of Jass and colleagues.³⁸ Intraepithelial lymphocytes were identified by light microscopy on haematoxylin and eosin sections as cells with the morphology of lymphocytes, seen wholly within tumour epithelium. They were classified as conspicuous when more than 30 were present per 10 high power fields.

Tumour volume in this study was estimated from reported tumour dimensions, using the formula $V = \pi ((L+T)/4)^2 \times D$, where V =volume (ml); L =longitudinal dimension (cm); T =circumferential dimension (cm); and D =depth of tumour (cm).

DNA EXTRACTION

For preparation of DNA from fresh tissues, frozen tissue was macerated in 500 µl of ice cold lysis buffer (10 mM Tris HCl, 1 mM EDTA, 100 mM NaCl, 1% sodium dodecyl sulphate, 500 µg/ml proteinase K) using a sterile Eppendorf homogeniser. Following incubation overnight with shaking at 50°C , DNA was extracted with phenol/chloroform and precipitated with ethanol.

For extraction of DNA from paraffin embedded tissues, 4×5 µm sections from the tissue block were incubated in 250 µl of paraffin lysis buffer (50 mM KCl, 2.5 mM MgCl_2 , 0.45% Triton-X, 10 mM TRISMA base, 500 µg/ml proteinase K) for two hours at 65°C and then overnight at 55°C . The tubes were then boiled for five minutes, centrifuged at 14 000 g for two minutes, and finally chilled prior to removal of the DNA solution from beneath the solidified paraffin. For each tumour block an adjacent section was stained with haematoxylin and eosin, and examined by light microscopy to ensure that the section contained more than 60% tumour tissue.

ANALYSIS OF MICROSATELLITE INSTABILITY

In all reactions, DNA was amplified in a 10 µl volume containing 100 ng of DNA, 200 µM dNTPs, 1.5 mM MgCl_2 , 0.27 µM of each primer, 0.25 U Tth polymerase in a buffer of 16.6 mM $(\text{NH}_4)_2\text{SO}_4$, 0.45% Triton X-100, 0.2 mg/ml gelatin, and 67 mM Tris HCl, pH 8.8 (Biotech International, Ltd, Western Australia). The reactions were incubated at 95°C for five minutes followed by 35 cycles of 95°C , 57°C , and 72°C for one minute each. The primers used in this study were Bat25, Bat26, D5S346, D2S123 and D17S250.³⁹ Products were run on an ABI 377 sequencer (Foster City, California, USA) and analysed using Genescan and Genotyper software.

MSI was defined as either marked alteration in repeat length (often appearing as a "ladder-like" expansion) or as a new discrete band

above or below the expected allele. Small increases in amplitude or numbers of stutters in the tumour samples were not considered to represent instability at the marker. A single observer (RW) performed analysis of MSI and equivocal samples were re-evaluated by a second observer (IT). Results were reported without knowledge of pathological or immunohistochemical status. A tumour sample was considered to be MSI-H if two or more of the markers demonstrated instability and to be MSI-L when only one marker was unstable.³⁹ In some cases MSI analysis was repeated using DNA extracted from paraffin embedded tissue. This was performed where the individual had synchronous cancers and the origin of fresh samples was uncertain, where polymerase chain reaction (PCR) amplification of DNA from fresh tissue was not satisfactory, or where there were indeterminate results from one or more markers. Overall, analysis of paraffin extracted DNA was necessary to obtain reliable data in 29 tumours.

IMMUNOHISTOCHEMICAL STAINING FOR MSH2 AND MLH1

Following dewaxing and rehydration of 4 µm tissue sections on silane coated slides, endogenous peroxidase was blocked using 3% hydrogen peroxide in methanol for 20 minutes. Antigen retrieval was performed by pressure cooking for five minutes in 0.1 M citrate buffer (pH 6.0). Slides were then blocked with 20% normal horse serum diluted in 2% bovine serum albumin/Tris buffered saline prior to overnight incubation with monoclonal antibodies against MSH2 (1:400) and MLH1 (1:200) (Pharmingen, San Diego, California, USA). Bound antibody was detected with biotinylated horse antimouse immunoglobulin (1:100; Dako, Dakopatts, Denmark) and horseradish peroxidase labelled streptavidin (SA-HRP, 1:200; Dako). Staining was considered assessable where nuclear staining was seen in either stromal or germinal follicle lymphocytes, or in normal epithelial cells in the base of crypts. In assessable slides, complete absence of staining of tumour cells for one of the mismatch repair proteins in the presence of staining for the other protein was considered indicative of a mismatch repair defect.

ANALYSIS OF SOMATIC CHANGES IN p53 AND K-ras
Mutations at the first and second bases of codon 12 of the K-ras gene were detected using restriction endonuclease mediated selective PCR, as previously described.⁴⁰ For identification of accumulation of p53 within tumour cells, paraffin sections of tumour tissue were subjected to immunohistochemical analysis of p53, as previously described,⁴¹ using the mouse antihuman p53 antibody DO7 (Dako). Tumour was considered to show accumulation of p53 protein when more than 20% of tumour cells showed nuclear staining of moderate to high intensity, in the absence of staining in the stromal cells and normal epithelium.

STATISTICAL ANALYSIS

Survival was measured from the date of resection of colorectal cancer until death or until the censor date of 1 March 2000. Time to recurrence was the period from resection to medical documentation of tumour recurrence. Survival and time to recurrence were censored at time of death for patients dying of causes other than colorectal cancer. Survival curves were prepared according to the method of Kaplan and Meier, and statistical significance between curves was tested using the log rank test. Univariate survival analysis was performed by Cox proportional hazards model, and the hazard ratio and its 95% confidence interval (CI) were assessed for each factor. A multivariate analysis by stepwise backward conditional selection was used to determine which factors were independent prognostic factors for survival. For the 210 individuals alive at the time of the last follow up, the median follow up period was 32 months (range 1–74).

Categorical variables were compared using the χ^2 test or Fisher's exact test as appropriate. For comparison of MSI status, the Student *t* test was used for analysis of patient age, Wilcoxon rank sum test for analysis of tumour volume, and Spearman's test for tumour stage. A probability value of less than 0.05 was considered significant. All data were analysed using the SPSS statistical software v9.0 (SPSS Inc., Chicago, Illinois, USA).

Figure 1 Microsatellite analysis of paired normal (N) and tumour (T) DNA using Bat 26 (A, B) and D5S346 (C, D). The tumours in (A), (C), and (D) are unstable. The tumour in (B) (stable) has a germline polymorphism that is only identifiable as such by the presence of the same pattern in normal tissue. (C) (laddering) and (D) (new allele) are representative of the patterns of changes observed at the dinucleotide markers. Size in base pairs is indicated, and the scale on the right shows signal amplitude.

Table 1 Comparison of differences in a range of clinicopathological characteristics between MSI-H and MSS colorectal carcinomas. MSS tumours include the MSI-L group. Except where indicated (*Spearman's), significance was determined by Fisher's exact test (two tailed)

	MSI		p Value	PPV	NPV
	+	-			
Right sided tumour					
+	26	89	<0.001	0.23	0.96
-	7	188			
Intraepithelial lymphocytes					
+	22	41	<0.001	0.35	0.96
-	11	234			
Poor differentiation					
+	11	31	0.002	0.26	0.92
-	22	246			
Crohn's-like reaction					
+	11	32	0.002	0.26	0.91
-	22	235			
Mucinous tumour					
+	10	40	0.022	0.20	0.91
-	22	235			
Peritumoural lymphocytes					
+	16	52	<0.001	0.24	0.93
-	17	225			
Circumscribed margin					
+	29	198	0.059	0.13	0.95
-	4	79			
Tumour stage					
I	5	46	0.27*		
II	17	96			
III	7	90			
IV	4	45			
Sex					
F	21	107	0.004		
M	10	162			
Amsterdam criteria					
No	30	266	0.35	0.25	0.90
Yes	1	3			

MSI, microsatellite instability; MSI-H, high level of microsatellite instability; MSI-L, low level of microsatellite instability; MSS, microsatellite stable; PPV, positive predictive value, NPV, negative predictive value.

Results

FREQUENCY OF MICROSATELLITE INSTABILITY

Of the 310 tumours in which MSI analysis was possible, 33 (10.6%) were MSI-H, 21 (6.8%) were MSI-L, and 256 (82.6%) were MSS. DNA derived from fresh and paraffin embedded tissue of two tumours was inadequate for PCR and thus MSI status could not be determined in these cases.

The Bat26 germline allele (approximately 120 bp) was present in all evaluable samples while an additional shortened allele was present at 105–118 bp in 30 of the 33 MSI-H tumours (fig 1A). Two of the MSI-H tumours were stable at both Bat26 and Bat25 while a further tumour was unstable at Bat25 but polymorphic at Bat26. As a marker of instability, Bat25 proved to be marginally less sensitive than Bat26 as only 29 of the 33 tumours were unstable at this locus. While three individuals were polymorphic for Bat25 and/or Bat26 (overall frequency 1.0 %), concomitant analysis of normal tissue allowed correct classification of these tumours (fig 1B).

The most frequently unstable dinucleotide marker was D5S346, followed by D2S123 and D17S250 (fig 1C, D). Although the pattern of change was marker dependent, the appearance of a new allele was the least frequent pattern seen in unstable tumours (30%). In contrast, all MSI-L tumours were stable at Bat26 and Bat25, and the predominant change in the dinucleotide markers was the appearance of a new allele (50–60% of cases). In MSI-L tumours, the unstable marker was found to be

D5S346 in 52%, D2S123 in 30%, and D17S250 in 20% of cases.

PATHOLOGICAL, GENETIC, AND CLINICAL

CORRELATES OF MICROSATELLITE INSTABILITY

Statistical analysis showed that there were no significant differences between MSI-L and MSS tumours in any of the clinicopathological characteristics examined in this study. In contrast, MSI-L tumours differed significantly from MSI-H tumours in terms of right sidedness ($p<0.001$) and presence of intraepithelial lymphocytes ($p=0.01$). On this basis, MSI-L tumours were included in the MSS category for all subsequent analysis of microsatellite status and its relationship to clinicopathological and genetic variables.

MSI-H tumours were significantly more likely to be of high histopathological grade, have a mucinous phenotype, and to harbour increased numbers of both intraepithelial and peritumoural lymphocytes. They were also more likely to be right sided and to occur in women (table 1). Although the MSI-H phenotype appeared to be over represented in stage II tumours, this association did not reach statistical significance. There was no association between MSI-H status and patient age or family history, as determined by the Amsterdam criteria (table 1) or modified Amsterdam criteria (data not shown).

Positive and negative predictive values were calculated for each pathological parameter (table 1) as well as for all possible simple combinations of four of these values (sidedness, intraepithelial lymphocytes, grade, and margin type). On this basis, it was apparent that no individual pathological parameter had a clinically useful positive predictive value. In simple combination, the presence of both right sidedness and intraepithelial lymphocytes was the most accurate predictor of MSI-H phenotype, with a positive predictive value of 57% and a negative predictive value of 95%.

In general, MSI-H tumours had a greater tumour volume, although this was not statistically significant (MSS median 13 ml, range 0–567; MSI-H median 25 ml, range 1–187; $p=0.06$). When considered by stage, this effect was not present in stage I tumours but was present in stage II tumours (MSS median 15 ml, range 1–448; MSI-H median 24 ml, range 6–110; $p=0.20$) and most marked in stage III tumours (MSS median 12 ml, range 1–232; MSI-H median 31 ml, range 1–70; $p=0.12$).

Sixteen individuals in the study had either synchronous (10 individuals) or metachronous (seven individuals) colorectal carcinomas (one individual had both synchronous and metachronous tumours). One individual had two synchronous MSI-H tumours while another had simultaneous MSI-H and MSS carcinomas. A further individual with an MSI-H tumour had two metachronous colorectal carcinomas. The remaining 13 individuals were MSS and there was no significant increase in the frequency of metachronous or synchronous tumours in those with MSI-H tumours ($p=0.39$).

Figure 2 Kaplan-Meier survival curves for individuals with colorectal carcinoma. (A, B) Survival according to microsatellite instability status in stage II and stage III tumours, respectively. (C) Overall survival as a function of sex. MSI-H, high level of microsatellite instability; MSS, microsatellite stable.

K-ras mutations were detected in 82 of 300 tumours analysed (27%) while immunohistochemical evidence of p53 accumulation was observed in 157 of 309 tumours (50.8%). While the incidence of K-ras mutations was less frequent in tumours with MSI (16%) than in either MSI-L (29%) or stable tumours (29%), this difference was not significant ($p=0.20$). On the other hand, accumulation of p53 was much less common in MSI-H cancers (18%) than in non-MSI cancers (55%) ($p<0.001$).

DISEASE OUTCOMES AND MICROSATELLITE STATUS

Analysis of overall survival and time to recurrence was performed on all individuals who had undergone potentially curative tumour resection (stages I–III, R0).

Cox's univariate analysis showed that female sex (hazard ratio (HR) 0.4 (CI 0.17–0.87);

$p=0.02$), lower tumour stage (HR 4.62 (CI 2.3–9.5); $p<0.0001$), peritumoural lymphocytes (HR 0.24 (CI 0.07–0.77); $p=0.02$), circumscribed tumour margins (HR 3.46 (CI 1.75–6.8); $p<0.003$), and increased intraepithelial lymphocytes (HR 0.29 (CI 0.09–0.95); $p=0.04$) were each associated with improvement in overall survival. No survival differences were accorded by any other pathological factor, MSI status, or by the presence of p53 accumulation or K-ras mutation within the tumour. In the multivariate analysis, tumour stage (HR 4.34 (CI 2.1–8.77); $p<0.001$) and female sex (HR 0.41 (CI 0.18–0.91); $p=0.03$) proved to be the only independent predictors of improved survival.

Survival curves for patients with stage II and III cancers according to microsatellite status, as well as overall survival by sex, are shown in fig 2. Although the differences between microsatellite groups did not reach statistical significance it is apparent that individuals with MSI-H tumours showed a trend towards both a prolonged recurrence free interval and improved overall survival. It is notable that of the 17 patients with stage II cancer, none had developed recurrent disease or died from cancer as of the censor date.

In relation to survival, it is also important to note that there were an increased number of stage II tumours in the MSI-H group, and a concomitant decrease in relative terms in stages III and IV (table 1). However, these differences in stage distribution did not reach statistical significance and were not reflected in improved overall survival.

DETECTION OF MSI USING IMMUNOHISTOCHEMISTRY

The typical appearance of tumours stained with antibodies against MLH1 and MSH2 is shown in fig 3. Immunohistochemistry could not be interpreted in two tumours, both from the MSS group, because of failure of staining of normal cells. Overall, 25 (8%) tumours showed loss of staining for MLH1 and a further three tumours (1%) showed absence of staining for MSH2. No tumour showed loss of staining with both antibodies.

There was a highly significant correlation between MSI-H status and loss of expression of MSH2 or MLH1 ($p<0.001$) (table 2). In fact, immunohistochemical analysis with MLH1 and MSH2 offered a specificity of 99.6% for detection of MSI as only one of 275 MSS tumours was reported to show loss of expression of a mismatch repair enzyme (IHC false positive). The sensitivity of the immunohistochemical analysis was 81% as it was unable to detect six of 33 MSI-H tumours (IHC false negatives). The positive predictive value of immunohistochemistry in the detection of MSI-H tumours was calculated to be 96% and the negative predictive value was 98%.

The one IHC false positive tumour was MSI-L by genetic testing. However, this was determined using fresh tissue and it is possible that there may have been insufficient tumour in

Figure 3 Typical appearance of tumours showing loss of mismatch repair enzymes, as demonstrated by immunohistochemistry for MLH1. (A) Poorly differentiated carcinoma cells (upper) show no nuclear staining for MLH1 while normal crypt epithelium (lower left) and stromal lymphocytes show nuclear positivity (brown stain); bar=500 µm. (B) Details of negative tumour cell staining in the presence of dark staining positive lymphocytes, both within tumour epithelium and in surrounding stroma. Normal epithelium is seen in the lower left; bar=100 µm; immunoperoxidase with haematoxylin counterstain.

Table 2 Comparison of microsatellite analysis with MSH2 and MLH1 immunohistochemical findings. Mismatch repair defective tumours are those carcinomas which showed loss of either MLH1 or MSH2 expression by immunohistochemistry

	MSI		Total
	MSS	MSI-H	
MLH1 and MSH2 present	274	6	280
MLH1 loss	1	24	28
MSH2 loss	0	3	3
Total	275	33	308

MSI, microsatellite instability; MSI-H, high level of microsatellite instability; MSS, microsatellite stable.

the sample to ensure accurate analysis of all markers.

The clinicopathological features of the six IHC false negative tumours are shown in table 3. While most cases have the typical morphology of MSI-H tumours it is interesting that two

cases in the transverse colon did not show increased levels of intraepithelial lymphocytes or mucinous phenotype, were moderately differentiated, and had mutations of p53. In terms of microsatellite analysis, one of these two cases (FN3) was unstable only at the BAT25 and BAT26 while the other case (FN4) was unstable only at D5S346 and D2S123. The other four cases showed instability at three or more microsatellite loci, including BAT markers. Review of immunohistochemical data confirmed the fact that they all expressed both MSH2 and MLH1 protein.

Based on clinical data, in combination with immunohistochemical and genetic analysis of tumours, it was possible that up to six individuals in this study with apparently sporadic MSI-H tumours may actually harbour germline mutations of a mismatch repair gene.

Table 3 Characteristics of individuals and tumours with MSI-H tumours but with no mismatch repair defect, as determined by immunostaining (false negative IHC)

ID	Age	Sex	AC	Stage	Site	IEL	Mucin	Grade	p53	K-ras
FN1	62	F	No	I	AsC	+	No	Mod	Wild	Mut
FN2	62	M	No*	II	AsC	+	Yes	Poor	Wild	Wild
FN3	67	F	No	II	TC	–	No	Mod	Mut	Wild
FN4	77	F	No	II	TC	–	No	Mod	Mut	Wild
FN5	83	F	No	IV	Caecum	+	Yes	Poor	Wild	Wild
FN6	84	F	No	IV	SC	–	No	Poor	Wild	Wild

AC, Amsterdam criteria, IEL, intraepithelial lymphocytes, Mut, mutant, TC, transverse colon, AsC, ascending colon, SC, sigmoid colon; MSI-H, high level of microsatellite instability.

*This patient had three first degree relatives with either breast or gastric carcinoma.

These included three individuals with tumours that were negative for MSH2 by immunohistochemistry (aged 46, 72, and 80 years) as somatic mutations of this gene are reported to be infrequent in sporadic MSI-H cancers.^{15 17} Interestingly, none of these three individuals fulfilled the Amsterdam criteria. Alternatively, reduced expression of MSH2 in these tumours may have been secondary to another genetic defect.²⁰ A further two individuals with MLH1 negative tumours were considered likely to represent individuals with germline disease as one was aged 32 years at the time of surgery while the other (aged 67) fulfilled the Amsterdam criteria. Finally, a sixth individual had a personal history of three colorectal carcinomas from age 38 as well as an MLH1 negative tumour. She was shown by subsequent genetic testing to have a germline mutation at exon 9 of MLH1 even though she did not fulfill the Amsterdam criteria.

When individuals likely to have a germline defect were excluded from the analysis, the remaining 21 individuals with MSI-H tumours that lacked MLH1 expression differed from other individuals in the study in that they were older (mean age 76 (9) years; $p=0.004$) and were more likely to be female (66% female; $p=0.04$).

Discussion

It is now well accepted that a significant proportion of sporadic colorectal carcinomas arise through a pathway that is characterised by defects in DNA repair enzymes and that is defined phenotypically by MSI. The extent and significance of this phenomenon in sporadic disease remain less well delineated.

Accrual of accurate data on the frequency and significance of MSI in sporadic colorectal cancer must be predicated on the unbiased selection of cases. It is now established that the rate of MSI-H tumours is relatively high in certain selected populations, including younger individuals (17–58%^{42–45}) and those with mucinous tumours (36%²⁴). In the case of the younger population, this is almost certainly related to a significantly higher frequency of germline defects.⁴⁵ In this study, we have examined the frequency and significance of MSI in a prospective fashion from a population of individuals with apparently sporadic colorectal carcinoma. It is inevitable that some individuals enrolled in this study with apparent sporadic disease were in fact harbouring germline mutations and we estimate that up to six individuals in this study may fall into this category. Nevertheless, for the purposes of this study, all individuals were considered to represent sporadic disease, in the sense that they presented for surgery and pathological assessment of their tumour without any expectation by the relevant medical staff that their condition represented a hereditary disease. In practical terms, we believe that this is the “sporadic” population that confronts the surgeon, pathologist, or oncologist outside centres with an interest in hereditary disease, and the population from which data on the value and significance of MSI is most urgently needed.

PATHOLOGICAL AND GENETIC FEATURES OF MSI IN SPORADIC COLORECTAL CARCINOMA

Many of the observations made in this study regarding the frequency, phenotype, and genotype of sporadic MSI-H tumours are in concordance with previously published studies.^{13 23} We found that MSI-H tumours were more frequent in the right colon, that they often had a mucinous and poorly differentiated phenotype, and that intraepithelial and peritumoural lymphocytic infiltration were often prominent. In common with others, we found that MSI-H tumours associated with apparent somatic loss of MLH1 were more common in the elderly and in women, and that there was a strong correlation between MSI-H and absence of p53 mutations, as determined by immunostaining.^{13 46}

However, MSI-H tumours were seen in only 10.6% of this unselected population, a rate that is lower than any previously reported study.^{22 23 27} For the reasons indicated above, we consider that this number may represent a more accurate estimate than has previously been reported in the literature on sporadic MSI-H colorectal carcinoma. Similar to many other reports, we found that mononucleotide repeat markers provided the most sensitive and specific single marker of instability.^{15 47}

As a result of this work, we are also able to comment on the significance of the MSI-L phenotype in sporadic colorectal carcinoma. It has previously been proposed that the MSI-L phenotype reflects a distinctive pathway of tumour development,³⁴ involving sequences other than mononucleotide repeats.¹⁵ While data from this study support the association of the MSI-H phenotype with a range of characteristic pathological features, the same cannot be said of the MSI-L group of cancers. In fact, we were unable to find any clinicopathological features which distinguished this group of cancers from the MSS group. On the other hand, MSI-L tumours showed clear pathological differences from MSI-H tumours in a number of areas.

CLINICAL SIGNIFICANCE OF MSI IN THE SPORADIC SETTING

Detection of MSI in the setting of hereditary disease appears to identify individuals at increased risk of second tumours, as well as those with an improved survival compared with MSS tumours. To date, information on these clinical parameters has been less clear in the setting of sporadic disease.

Several small and retrospective studies have suggested that individuals with MSI-H sporadic tumours are at greater risk of metachronous cancers.^{22 23 29 30} More recently, Gryfe and colleagues found a higher incidence of both metachronous and synchronous tumours in younger patients with MSI-H cancers, a study which is likely to have included more patients with germline mismatch repair defects.⁴⁵ However, we were unable to show an increase in the risk of such tumours in our study group, which consisted largely of older individuals.

The relationship between MSI status and disease outcomes in sporadic colorectal carcinoma remains uncertain. In this study, patients with MSI-H cancers showed a trend towards improved clinical outcome, and it is possible that as the period of follow up matures, these differences will reach statistical significance. Results from other studies on unselected populations, in combination with our own, suggest that MSI-H tumours have a survival advantage.⁴⁸

Interestingly, the most compelling evidence to date for increased survival in sporadic MSI-H tumours has come from studies of young individuals.⁴⁵ It is possible that the discrepancies in reported outcomes seen in recent studies may reflect a biological distinction between old and young individuals with microsatellite unstable tumours. In this regard, it has been proposed that MSI may influence survival through induction of an antitumour response, as evidenced by the marked lymphocytic infiltrate in the tumour. It is tempting to speculate that the apparent improved survival seen in younger people with MSI-H tumours may be related to a more intense host immune response in these individuals.

A further consideration in assessing survival is the tendency for MSI-H tumours to fail to progress from stage II to III. This is manifest in this study by the increased percentage of stage II tumours, decreased percentage of stage III tumours, and also by the observation that MSI tumours tend to be larger than stable tumours in all stages except for stage I. The biological basis of this phenomenon is not clear from this study but may be related to the low frequency of p53 mutations in MSI tumours, perhaps resulting in inhibition of tumour progression. Importantly, if delayed progression from local (stage II) to metastatic (stage III) disease is a feature of MSI tumours, it may result in an overall survival advantage, even in the absence of stage specific improvements in survival.

ALTERNATE METHODS FOR DETECTION OF MSI TUMOURS

While PCR amplification of microsatellite repeats remains the gold standard for recognition of the MSI phenotype, this approach is clearly not feasible in routine pathology laboratories. Nevertheless, the effectiveness of morphological and immunohistochemical analysis in the prospective identification of sporadic tumours remains uncertain. An important aim of this study was to identify the most appropriate methods for detection of MSI-H tumours in the setting of routine pathology practice.

It is recognised that sporadic MSI-H colorectal cancers share a number of the pathologically important features of hereditary disease, and it has been suggested that careful observation of tumour morphology can assist in the identification of these unstable tumours.⁴⁹ It is clear from this study that while some morphological parameters are useful in identifying those tumours that are unlikely to be MSI-H, the same is not true for the identification of tumours likely to be MSI positive. Based on

data from this study, a simple scoring that considered the presence of both right sidedness and intraepithelial lymphocytes had a positive predictive value of 57% and a negative predictive value of 95% (table 1). However, such a strategy would miss 13 of the 33 MSI high tumours identified in this study.

In contradistinction to morphology, we found that immunohistochemical detection of loss of expression of mismatch repair enzymes provided a highly specific approach to the identification of microsatellite unstable tumours (positive and negative predictive values 96% and 97.8%, respectively). Unlike PCR analysis for MSI, it also provides a rapid and technically simple approach to this problem.

Recently, Cawkwell and colleagues provided evidence for the clinical utility of this approach in a retrospective analysis of a highly selected phenotypic subgroup of tumours. On the basis of this work, they concluded that all MSI-H cancers could be detected by immunostaining.²² However, the current study clearly identified six tumours that were MSI-H yet had apparently normal expression of MLH1 and MSH2 by immunostaining. A similar false negative rate for immunohistochemistry has previously been reported⁵⁰⁻⁵¹ and is probably multifactorial in origin.

One possible explanation is that missense mutation of a DNA repair enzyme gene leads to expression of an immunologically reactive yet poorly functional protein. A second possibility is that these false negative tumours harbour mutations in other mismatch repair genes, or in as yet unidentified genes that affect MMR function. For example, hMSH6 mutations have been reported in HNPCC and sporadic colorectal carcinomas with MSI-H.⁵² Despite these findings, the role of hMSH6 remains to be determined as these MSI-H tumours also had mutations in other mismatch repair genes, and a recent study using colorectal cancer cell lines identified two cell lines with mutations in hMSH6 but with no MSI.⁵³ Regardless of the genetic aetiology, we feel that such MSI-H tumours with normal immunostaining represent a small but important group of tumours which warrant further study.

In this study, we have examined MSI in a large group of colorectal cancer patients with apparently sporadic disease. On the basis of this work, it appears feasible to incorporate immunostaining as part of the standard assessment of colorectal tumours. This knowledge is of considerable relevance to the future management strategies for all individuals with microsatellite unstable cancers.

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