Motility and visceral sensation

Pan-colonic decrease in interstitial cells of Cajal in patients with slow transit constipation

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Background: Interstitial cells of Cajal (ICC) are required for normal intestinal motility. ICC are found throughout the human colon and are decreased in the sigmoid colon of patients with slow transit constipation.

Aims: The aims of this study were to determine the normal distribution of ICC within the human colon and to determine if ICC are decreased throughout the colon in slow transit constipation.

Patients: The caecum, ascending, transverse, and sigmoid colons from six patients with slow transit constipation and colonic tissue from patients with resected colon cancer were used for this study.

Methods: ICC cells were identified with a polyclonal antibody to c-Kit, serial 0.5 µm sections were obtained by confocal microscopy, and three dimensional software was employed to reconstruct the entire thickness of the colon. ICC volume was measured in the slow transit constipation (STC) patients across all colonic regions.

Results: ICC were decreased in all regions of the colon. ICC volume was significantly lower in the slow transit constipation patients compared with controls.

Conclusions: The data suggest that ICC distribution is relatively uniform throughout the human colon and that decreased ICC volume is pan-colonic in idiopathic slow transit constipation.

MOTILITY AND VISCERAL SENSATION

Interstitial cells of Cajal (ICC) are required for normal intestinal motility. Their role as intestinal pacemakers has been established in a number of model systems. ICC are involved in the regulation of smooth muscle activity. The aetiology of STC remains unclear. Abnormalities in the neural networks of the colon have been demonstrated in patients with constipation but the specific neuronal defect remains unclear. Abnormalities in nitric oxide synthase (NOS) and vasoactive intestinal peptide (VIP) containing neurones have been demonstrated. Our group previously reported an alteration in ICC within the sigmoid colon of constipated patients. Specifically, the total ICC volume is decreased in the sigmoid colon of STC patients compared with controls and the defect appears throughout the thickness of the sigmoid colon.

In the present study, we used immunohistochemistry and three dimensional computer reconstruction to determine the distribution of ICC in all regions of the normal human colon and to determine if, in STC, ICC volume and distribution are altered in all regions of the human colon.

MATERIALS AND METHODS

Procurement of human tissue was approved by the Institutional Review Board of the Mayo Clinic. All tissue was immediately snap frozen after procurement and stored at −80°C until use.

Immunocytochemistry

Colon samples were fixed either with 4% paraformaldehyde in 0.1 M sodium phosphate buffer (PBS), pH 7.4, or Zamboni’s fixative (Newcomer Supply, Middleton, Wisconsin, USA) for four hours or held overnight at 4°C. Next, samples were

Abbreviations: ICC, interstitial cells of Cajal; STC, slow transit constipation; SMB, submucosal border; CM, circular muscle; MPR, myenteric plexus region; LM, longitudinal muscle; NOS, nitric oxide synthase; VIP, vasoactive intestinal peptide; PBS, phosphate buffered saline.
washed three times with 0.1 M PBS, transferred to 30% sucrose in 0.1 M PBS, and refrigerated for up to 24 hours until sectioned. Serial longitudinal, transverse, and horizontal sections (40–150 µm thick) were cut with a Cryostat (Miles Inc., Elkhart, Indiana, USA), placed in spot plates, and flooded with 0.1 M PBS with 0.3% Triton X-100 (Sigma, St Louis, Missouri, USA) and 5% normal donkey serum for two hours to permeabilise the tissue and reduce background staining. Immunohistochemical staining was performed with a c-Kit rabbit polyclonal antibody (1:400 dilution; MBL, Nagoya, Japan) and a c-Kit goat polyclonal antibody (1:200 dilution; Santa Cruz Biotech, California, USA). No significant difference was noted between the two c-Kit antibodies used. Immunoreactivity was demonstrated by incubating the tissues with primary antibodies (overnight, 4°C). Bound antibodies were visualised by incubating tissues for two hours with fluorescent labelled secondary antibodies to rabbit or mouse IgG (Jackson ImmunoResearch, West Grove, Pennsylvania, USA). After washing with PBS, sections were mounted on slides, air dried overnight at room temperature, and coverslipped. Non-specific labelling for c-Kit was assessed by omitting the primary antibody and by preabsorbing with the purified antigen in separate experiments. No staining was seen with either control. Additionally, non-immune serum produced no staining.

Volume determination of ICC and neural networks
Immunostained tissues were examined with a laser scanning confocal microscope (Zeiss 310, Oberkochen, Germany). Cy3 fluorescence was visualised using an Argon/Krypton ion laser at 568 nm for excitation and emission taken over 590 nm. Each specimen examined was divided into four anatomical regions: longitudinal muscle (LM), myenteric plexus region (MPR), circular muscle (CM), and submucosal border (SMB). Images were collected using an oil immersion x40 objective (NA 1.3) and an oil immersion x100 objective (NA 1.3). The 100× objective was used to ensure that weakly stained ICC were included in the quantitative analyses. In every region, four areas in the X-Y plane were examined. Under the 40× oil objective, the maximum area scanned in the horizontal X-Y plane was 320 µm×320 µm; under the 100× oil objective, the area scanned was 128 µm×128 µm. Optical sections (512×512 pixels) were recorded at 0.5 µm depth (optical Z axis) increments through each of the four adjacent areas. The volume (µm³) of total tissue (including interstitial space) was calculated by: area of the X-Y plane (µm²)×(number of optical
The volume of the ICC network was obtained directly from the volume data set after mathematically correcting for focal anomaly.

Mast cells were identified by their typical round shape and size and excluded from the analysis.

The investigator performing the data analysis was blinded to the clinical diagnosis—that is, control versus STC. The sigmoid colon specimens used in this study (both control and STC specimens) were different than those previously reported.21

**Statistics**

Values in the text are mean (SEM). Statistical differences among different regions were tested using an analysis of variance (ANOVA); p values <0.05 were considered statistically significant.

**RESULTS**

**Demographics**

All STC patients were female and between the ages of 18 and 55 years. Constipation was present in all patients for at least 10 years. Evaluations included barium enema or colonoscopy to rule out structural lesions. Colon motility, as assessed by marker studies, was significantly prolonged (>100 hours) in all patients. Significant pelvic floor dysfunction was excluded with anorectal manometry and balloon expulsion. One patient demonstrated a mildly abnormal balloon expulsion test and a markedly prolonged colonic transit test. Colonic resection was performed after the patient underwent pelvic floor retraining.

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**Figure 3** Three dimensional reconstruction of c-kit immunoreactivity in the transverse colon for control tissue (left panel) and slow transit constipation (right panel) specimens. Each panel is of a representative sample. Panels represent separate reconstructions in the submucosal border (SMB), circular smooth muscle layer (CM), myenteric plexus region (MPR), and longitudinal muscle layer (LM). Scale bar is 50 µm.

**Figure 4** Three dimensional reconstruction of c-kit immunoreactivity in the sigmoid colon for control tissue (left panel) and slow transit constipation (right panel) specimens. Each panel is of a representative sample. Panels represent separate reconstructions in the submucosal border (SMB), circular smooth muscle layer (CM), myenteric plexus region (MPR), and longitudinal muscle layer (LM). Scale bar is 50 µm.
with no improvement in symptoms. All STC patients failed a medical regimen of stimulant and osmotic laxatives as well as bulking agents prior to colectomy.

All control patients underwent colectomy for colon cancer. Because patients underwent segmental colectomy, samples were obtained from more than six patients so that a total of six samples were analysed for each colon region. Control ages ranged from 23 to 82 years. Four patients were male and eight were female.

**ICC distribution in normal human colon**

Mean (SEM) ICC volume in the SMB, CM, MPR, and LM in the caecum was 9.9 (1.4), 4.3 (0.5), 16.5 (1.5), and 3.6 (0.3)% respectively. A representative set of images is shown in fig 1. Ascending colon ICC volumes were 15.3 (2.0), 5.3 (0.5), 18.4 (2.6), and 4.6 (0.6)% (representative images in fig 2). Transverse colon ICC volumes were 16.2 (3.1), 4.3 (0.6), 22.6 (2.6), and 6.1 (0.9)% (representative images in fig 3). Sigmoid colon ICC volumes were 6.3 (1.0), 4.4 (0.4), 20.3 (1.8), and 4.7 (0.4)% (representative images in fig 4). These data are shown graphically in fig 5. The distribution of ICC volume in each region did not differ significantly from one anatomical segment to another. In contrast, ICC volume was significantly higher in the MPR and SMB regions than in the CM and LM regions in all anatomical segments (p<0.05).

**ICC in slow transit colon**

In STC patients, the caecum ICC volumes were 0 (0) in the SMB, 1.6 (0.4) in the CM, 2.8 (0.5) in the MPR, and 1 (0.3)% in the LM (representative images shown in fig 1). Ascending colon ICC volumes were 1.4 (0.5), 1.9 (0.3), 7.4 (1.2), and 1.3 (0.2)% (representative images shown in fig 2). Transverse colon ICC volumes were 2.5 (0.9), 2.4 (0.3), 9.5 (1.0), and 2.2 (0.3)% (representative images shown in fig 3). Sigmoid colon ICC volumes were 2.0 (0.7), 2.6 (0.4), 8.0 (1.4), and 0.8 (0.3)% (representative images shown in fig 4). As with control colon layers, there was no significant difference when each region was compared across colon segments, and the SMB and MPR values were higher compared with the LM and CM regions in all anatomical segments (p<0.05) (fig 5). ANOVA revealed a significantly lower ICC volume in slow transit patients compared with controls (p<0.05) and subsequent pairwise analysis revealed a significant difference at each region analysed (p<0.05).

**DISCUSSION**

In the normal human colon, we observed c-kit staining in each of the regions studied—that is, in the SMB, MPR, CM, and LM layers, and in all colonic segments analysed. A higher per cent volume of ICC was present in the SMB and MPR compared with the CM and LM layers. These data are similar to those reported for other species. No significant variation in the distribution of ICC volume among the caecum, ascending, transverse, and sigmoid colon was found. These results contrast with previous reports wherein the density of ICC has been reported to be higher in the myenteric plexus of the transverse colon. Methodological differences may explain the different results. Our tissues were snap frozen immediately after procurement and stored at −70°C until processing. We did not employ paraffin embedding which decreases c-Kit staining (unpublished results). Our scoring of ICC utilised computer reconstruction which allows for quantification of ICC volume and may be sensitive not only to changes in cell number but also cell and process size. Mast cells are the only other cell type known to express c-Kit in the intestine. Previous reports suggest these cells are less than 8% of c-Kit positive cells in the intestine, and hence were specifically excluded from the analysis.

We have focused our studies on ICC because mounting evidence suggests a crucial role for this cell population in regulating intestinal function. Recent studies suggest that ICC mediate neurotransmission from enteric motor neurons. Moreover, ICC act as intestinal pacemaker cells to generate slow waves. Two complimentary lines of evidence support this notion. Firstly, mutations of the c-kit ligand, steel factor, also disrupt intestinal ICC cell development and electrical rhythmicity. A second line of evidence demonstrates that ICC cells are sufficient to generate an electrical slow wave. ICC in primary culture systems exhibit a slow wave activity—that is, have the proper characteristics to be an intestinal pacemaker. Taken together, these studies suggest a central role for ICC in the regulation of intestinal motility. Important anatomical differences exist in ICC slow wave generation. For example, the ICC of the MPR generate the slow wave within the small intestine. In the colon however the ICC of the SMB region appear to produce the slow wave. The present demonstration of a distinct SMB ICC network throughout the human colon is consistent with this functional.
observation. Previous reports have suggested that the transverse colon is the primary colonic pacemaker site and that the myenteric plexus ICC is denser in the transverse colon. In this study, we were unable to demonstrate an anatomical correlation with this functional observation as both the MPR and SMB ICC volume appeared to be relatively uniform throughout the colon, including the caecum.

In addition to describing ICC distribution in the normal colon, we found reduced ICC volume in idiopathic STC. This reduction was present across all colon regions, including the caecum, ascending, transverse, and sigmoid colon. Our present report replicates our previous findings of decreased ICC volume in the sigmoid colon of patients with STC and expands our findings to demonstrate that this decrease is pervasive in all of the patients in our study who had been treated preoperatively with bulking agents as well as stimulant and osmotic laxatives, including anthraquinone preparations. Some concern exists over the possibility that laxatives may cause histological changes but controlled prospective trials in animals and humans demonstrate no neuronal changes with anthraquinone usage.

The effects of age on human ICC number or volume has not been studied in any age group. Our patient sample was lower than that of the control sample. This work was supported by National Institutes of Health grants DK17238, DK52766, and DK57061. The authors thank Kristy Zodrow and Michael Thompson for technical assistance. This study, we were unable to demonstrate an anatomical correlation with this functional observation as both the MPR and SMB ICC volume appeared to be relatively uniform throughout the colon, including the caecum.

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