Duodenal expression of iron transport molecules in untreated haemochromatosis subjects

K A Stuart, G J Anderson, D M Frazer, L W Powell, M McCullen, L M Fletcher, D H G Crawford

Background and aims: In HFE associated hereditary haemochromatosis, the duodenal enterocyte behaves as if iron deficient and previous reports have shown increased duodenal expression of divalent metal transporter 1 (DMT1) and iron regulated gene 1 (Ireg1) in affected subjects. In those studies, many patients had undergone venesection, which is a potent stimulus of iron absorption. Our study investigated duodenal expression of DMT1 (IRE) and Ireg1 expression were inappropriately increased relative to serum ferritin concentration. Hephaestin and Dcytb levels were not upregulated in haemochromatosis. DMT1 (IRE) and Ireg1 levels showed significant inverse correlations with serum ferritin concentration in each group of patients.

Conclusions: These findings are consistent with DMT1 (IRE) and Ireg1 playing primary roles in the adaptive response to iron deficiency. Untreated haemochromatosis patients showed inappropriate increases in DMT1 (IRE) and Ireg1 expression for a given level of serum ferritin concentration, although the actual level of expression of these iron transport genes was not significantly different from that of normal subjects.

Hereditary haemochromatosis (HHC) is a common autosomal recessive disorder of iron metabolism characterised by progressive accumulation of iron, primarily in the liver, pancreas, and heart, which may ultimately result in organ failure. The majority (80–90%) of patients with HHC are homozygous for a missense mutation that results in substitution of tyrosine for cysteine at position 282 (C282Y) in the haemochromatosis protein, HFE. A major pathophysiological feature of HHC is a rate of intestinal iron absorption that is inappropriately elevated for a given level of body iron stores. The precise mechanism by which HFE regulates intestinal iron absorption is unknown but the available evidence indicates that in HHC the duodenal enterocyte behaves as if it is iron deficient. This conclusion is supported by studies showing that in patients with HHC, the duodenal mucosal content of ferritin mRNA and protein is relatively low while the corresponding levels of transferrin receptor gene and protein expression are relatively high, and the binding affinity of iron regulatory protein is increased.

Iron absorption involves the uptake of iron from the lumen of the small intestine into the mature duodenal enterocyte, followed by its basolateral transfer into the portal circulation. Kinetic studies suggest that the transfer of iron across the basolateral membrane is the main regulatory step in normal subjects and HHC patients. Each iron transport step involves a transmembrane transporter that is likely to be coupled to an enzyme that changes the oxidation state of iron. Our understanding of intestinal iron absorption has improved in recent years following identification of several key proteins involved in the trafficking of iron into and out of cells. These include divalent metal transporter 1 (DMT1), iron regulated gene 1 (Ireg1 also known as SCL11A3, ferroportin1, or MTP1), hephaestin, and duodenal cytochrome-b (Dcytb).

DMT1 is a brush border transmembrane transporter for several divalent metals, including iron, copper, and zinc. It pumps iron into the enterocyte via an active proton coupled transport mechanism. At least two alternatively spliced transcripts of DMT1 exist, encoding alternative carboxy termini and 3′ untranslated regions (UTRs). The 3′ UTR of one variant contains a consensus iron responsive element (IRE) sequence that binds to iron regulatory proteins. DMT1 transports ferrous iron, indicating that dietary iron (which is mainly in the ferric form) must first be reduced before it can be utilised. A strong candidate for this ferrireductase activity is Dcytb, an enzyme that was recently identified as upregulated in the duodenal mucosa of the hpx mouse. Dcytb localises to the brush border membranes of mature enterocytes of the upper villous region, and both protein and mRNA levels are regulated by hypoxia, iron deficiency, and anaemia. Ireg1 is predicted to function as an iron transporter on the basolateral membrane of the mature villous enterocyte and elevated duodenal Ireg1 mRNA levels are observed in situations associated with increased intestinal iron absorption. Ireg1 is postulated to associate with hephaestin, a membrane bound ceruloplasmin homologue with ferroxidase activity.
Recently, Zoller et al reported 100-fold increase in duodenal DMT1 expression in 20 C282Y homozygous HHC subjects who were undergoing, or who had recently completed, therapeutic phlebotomy, compared with 10 subjects with normal body iron stores. Subsequently, the same authors (using a different technique) reported much lower increases, 8–10-fold and 2–5-fold in duodenal DMT1 and Ireg1 expression, respectively, in eight untreated C282Y homozygous HHC subjects compared with normal subjects, and reported that expression of these genes was similar to that observed in 19 subjects with iron deficiency. Rolfs et al have reported similar findings of increased DMT1 and Ireg1 expression in nine C282Y homozygous HHC subjects, all of whom had been treated to a state of iron deficiency or were undergoing venesection at the time of duodenal biopsy. Prior studies indicate that venesection per se causes a significant increase in iron absorption in HHC patients, and inclusion of phlebotomised subjects may have had a confounding effect on the results of these studies. Thus despite recent advances, the relationship between body iron stores and expression of these iron transporters has not been clearly defined in untreated HHC patients. To address this, duodenal expression of DMT1, Ireg1, hephaestin, and Dcytb was investigated in untreated C282Y homozygous HHC patients, subjects with iron deficiency, and subjects with normal biochemical iron indices. Expression of these iron transport genes was related to serum iron indices and haematological parameters.

**METHODS**

**Study population**

Twelve C282Y homozygous HHC patients, 11 subjects with iron deficiency, and 13 iron replete subjects (that is, normal serum iron indices) were studied. The 12 HHC subjects were diagnosed in a tertiary hospital phenotypic screening study, whereby subjects with a corrected total iron binding capacity (TIBC) >40% were genotyped for the common mutations in the HFE gene. All subjects homozygous for the C282Y mutation were contacted and reviewed in a dedicated hepatology clinic. Percutaneous liver biopsy was performed at the time of diagnosis in six HHC subjects, all of whom had (i) a serum ferritin concentration (SF) ≥1000 µg/l, (ii) elevated serum alanine or aspartate transaminase levels, (iii) hepatomegaly (assessed either clinically or ultrasonographically) and/or (iv) coexistent cause for liver disease. Hepatic iron stores were assessed and graded following Perls’ Prussian blue stain by a single hepatopathologist according to the method described by Searle and colleagues. Quantitative hepatic iron measurements were performed on fresh liver tissue using a modification of the colorimetric method of Torrance and Bothwell.

All study subjects underwent gastroscopy for investigation of a range of clinical indications, including dyspepsia, iron deficiency, anaemia, heartburn, or dysphagia. For the purposes of this study, iron deficiency was defined as a transferrin saturation (TS) of <15% and an SF of <30 µg/l, or documented absence of iron stores on bone marrow biopsy. Subjects in the iron replete group had serum iron indices within the absence of iron stores on bone marrow biopsy. Subjects in the iron replete group had serum iron indices within the laboratory reference ranges (that is, TS 15–45%; SF 30–300 µg/l). HFE genotyping was determined in non-HHC subjects, as described elsewhere.

Fasting serum iron indices and haematological parameters were measured in all 36 subjects within two weeks of gastroscopy. Patient records were reviewed for a history of red blood cell or iron transfusions, or upper gastrointestinal bleeding during the weeks preceding gastroscopy. None of the subjects was a regular blood donor or taking oral iron during the study. Written consent was obtained from all subjects and the study was approved by the Princess Alexandra Hospital Research Ethics Committee.

**Tissue collection**

All duodenal biopsy specimens were obtained before phlebotomy was initiated in HHC subjects with significant iron overload who required treatment. Biopsies were collected from the second part of the duodenum, snap frozen in liquid nitrogen, and stored at −70°C for total RNA extraction at a later date.

**RNA preparation and ribonuclease protection assays**

Total RNA was extracted from two pooled duodenal biopsies from each individual using TRIzol reagent (Invitrogen, Melbourne, Australia) according to the manufacturer’s instructions. Ribonuclease protection assays (RPAs) were performed on total RNA using riboprobes corresponding to the cDNA sequences (table 1). Riboprobe synthesis was performed using the appropriate linearised vector and the Riboprobe Combination System (SP6/T7 polymerase kit; Promega, Madison, Wisconsin, USA) with minor modifications. Each transcription was carried out in a 20 µl reaction containing 40 mM Tris HCl (pH 7.9), 10 mM NaCl, 6 mM MgCl2, 2 mM spermidine, 10 mM diethiothreitol, 0.5 mM each of rATP, rGTP, and rCTP, 17.5 µM cold rUTP, 1 µg vector DNA, 50µCi [α-32P]UTP (3000 Ci/mmol; NEN, Adelaide, Australia), 20 U RNasin, and 15–20 U T7 or SP6 RNA polymerase, as appropriate. The transcription reaction was incubated at 37°C for one hour before digestion with 1 U RQI RNase-Free DNasel (Promega) for a further 15 minutes at 37°C. The riboprobes were electrophoresed on a 6% polyacrylamide, 50% formamide gel run at 100 V for 4 hours and then transferred to Hybond XAR films according to the instructions. Riboprobes visualised using autoradiography were excised and eluted for three hours at 37°C in 400 µl elution buffer (2 M ammonium acetate, 1% sodium dodecyl sulphate, 25 µg/ml yeast RNA) before ethanol precipitation and resuspension in 50 µl of hybridisation buffer (40 mM PIPES, pH 6.4, 400 mM NaCl, 1 mM EDTA, 80% formamide).

Human duodenal RNA samples (2 µg) were hybridised overnight at 45°C with 10 µg from all riboprobes. Following hybridisation, unpurified RNA was digested by addition of RNase digestion buffer (10 mM Tris HCl, pH 7.5, 300 mM NaCl, 5 mM EDTA) containing 80 µg/ml of RNase A and 80 µU/ml of RNase T1 (Roche Molecular Biochemicals, Castle Hill, Australia), and incubated at 37°C for 60 minutes. The reaction was extracted with 400 µl of phenol/chloroform/isoamyl alcohol (25:24:1) and the RNA-riboprobe hybrid precipitated with ethanol. Hybridised riboprobe samples were resolved on a 6% polyacrylamide, 50% urea gel for five hours at 200 V and detected after exposure for up to 96 hours at −70°C. The intensity of specific bands was determined by densitometry using ImageQuant software (Molecular Dynamics, Sunnyvale, California, USA). All RPAs were examined at different exposures and only those bands that lay within the linear range of the film were analysed. Expression of each of the iron transport genes was reported as a ratio of glyceraldehyde-3-phosphate dehydrogenase (GAPDH) to correct for differences in total mRNA loading.

<table>
<thead>
<tr>
<th>Table 1</th>
<th>cDNA sequences of riboprobes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Size (nt)</td>
<td>cDNA sequence</td>
</tr>
<tr>
<td>DMT1 (non-IRE)</td>
<td>273</td>
</tr>
<tr>
<td>DMT1 (IRE)</td>
<td>224</td>
</tr>
<tr>
<td>Ireg1</td>
<td>173</td>
</tr>
<tr>
<td>Hephaestin</td>
<td>190</td>
</tr>
<tr>
<td>Dcytb</td>
<td>242</td>
</tr>
<tr>
<td>GAPDH</td>
<td>155</td>
</tr>
</tbody>
</table>

*This riboprobe is based on the non-IRE splice variant but can also detect the iron responsive element (IRE) form as it crosses the splice site that distinguishes the two transcripts.
**Table 2 Clinical and laboratory characteristics of the study population**

<table>
<thead>
<tr>
<th></th>
<th>Haemochromatosis (n=12)</th>
<th>Iron deficient (n=11)</th>
<th>Iron replete (n=13)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex (M:F)</td>
<td>9:3</td>
<td>4.7</td>
<td>7.6</td>
<td>0.174</td>
</tr>
<tr>
<td>Age (y)</td>
<td>44.8 (15.9)‡ [17-69]</td>
<td>60.1 [19.6] [19-80]</td>
<td>57.4 [11.6] [39-77]</td>
<td>0.05</td>
</tr>
<tr>
<td>Hb (g/l)</td>
<td>148.8 (14.3)‡</td>
<td>126.2 [17.2]†††</td>
<td>142.1 [11.4]†</td>
<td>0.001</td>
</tr>
<tr>
<td>Hct (%)</td>
<td>43.9 (3.9)‡</td>
<td>33.6 (4.4)†††</td>
<td>42.9 (2.9)</td>
<td>0.001</td>
</tr>
<tr>
<td>TS (%)†††</td>
<td>63.0 (16.0)‡</td>
<td>6.7 (3.1)</td>
<td>25.3 (8.6)</td>
<td>0.001</td>
</tr>
<tr>
<td>SF (µg/l)†††</td>
<td>900 (39-1806)‡</td>
<td>13 (2-65)</td>
<td>137 (31-244)</td>
<td>0.001</td>
</tr>
</tbody>
</table>

Hb, haemoglobin; Hct, haematocrit; TS, transferrin saturation; SF, serum ferritin concentration.

Laboratory reference ranges for TS, SF, Hb, and Hct are 15-45%, 30–300 µg/l, 135–180 g/l, and 39–52%, respectively.

Data expressed as mean (SD) [range] or median (range).

Pearson’s χ² analysis was used to determine significant differences between groups for sex distribution. For all other variables, ANOVA was used to compare data between the three groups (right hand column) and the Student’s t test to compare data between two specific groups.

*p<0.05 for subjects with hereditary haemochromatosis (HHC) compared with iron deficient or iron replete subjects; †††p<0.001 for iron deficient subjects compared with HHC patients or iron replete subjects; ††p<0.001 for HHC subjects compared with both other groups and iron deficient subjects compared with iron replete subjects.

**RESULTS**

Demographic and laboratory characteristics of the study population

Age and sex distributions, and laboratory characteristics of the 36 patients included in the study are shown in table 2. Sixteen of 36 patients were female. While the majority of iron deficient subjects were female, most HHC patients were male. Mean age of the HHC patients was significantly lower than subjects in the iron deficient and iron replete groups (p=0.05). There were significant differences in mean haemoglobin concentration, haematocrit, TS, and median SF between the three groups (p<0.001, p<0.001, p<0.001, and p<0.001, respectively).

All HHC patients had TS levels >40% but three subjects, all female, had normal SF values (39, 59, 65 µg/l). Two of these women were of child bearing age and still menstruating, and the third was aged 56 years with a history of eight pregnancies. The remaining nine subjects were male and all had SF values >504 µg/l. Of the six HHC patients who underwent liver biopsy, five subjects had minimal or no fibrosis and one subject had evolving cirrhosis. Three HHC subjects, all male, had a history of heavy alcohol consumption (>60 g/day) but only one had mild steatosis on liver biopsy. Another patient with a body mass index of greater than 30 kg/m² but no history of alcohol consumption had severe steatosis. All six patients had grade 3 or 4 stainable liver iron and a hepatic iron concentration between 47 and 160 µmol/kg dry weight.

**Expression of DMT, Ireg1, hephaestin, and Dcytb**

mRNA levels of the iron transport molecules were examined by RPA and assays of three representative patients from each group are shown in fig 1A. Figure 1B shows mean levels for DMT1 (IRE), Ireg1, hephaestin, and Dcytb expressed as a percentage of the relative expression of GAPDH. There were significant differences in mean levels of DMT1 (IRE). Ireg1, and hephaestin between the three groups (p<0.001, p=0.001, and p=0.015, respectively) but there was no significant difference between the groups in Dcytb expression (p=0.12). Duodenal expression of DMT (IRE) and Ireg1 was similar in HHC patients and iron replete subjects (0.54 (0.07) v 0.59 (0.11), p=0.95;
all patients and did not contribute significantly to the level of DMT1

Expression of DMT1 (IRE) and Ireg1 was increased 4–5-fold and 3–4-fold, respectively, in patients with iron deficiency compared with iron replete subjects (2.96 (0.38) vs 5.9 (0.11), p<0.0001; 2.26 (0.18) vs 6.1 (0.07), p<0.0001) (fig 1B). In contrast, there were no significant differences in hephaestin or Dcytb expression in subjects with iron deficiency compared with iron replete subjects (0.96 (0.14) vs 0.9 (0.09), p=0.076; 1.32 (0.23) vs 1.12 (0.19), p=0.44, respectively). Expression of DMT1 (non-IRE) was extremely low in duodenal biopsies from all patients and did not contribute significantly to the level of total DMT1 present (data not shown).

Expression of DMT1 (IRE), Ireg1, hephaestin, and Dcytb in relation to serum iron indices and haematological parameters

In this series of untreated HHC patients, a strong inverse relationship between log SF and DMT1 (IRE) expression (r=-0.789, p=0.002) was found and a similar association was observed for Ireg1 (r=-0.621, p=0.03) but there was no significant correlation between TS, haemoglobin concentration, or haematocrit and duodenal iron transport gene expression (table 3). In non-HHC subjects, strong inverse correlations were found between DMT1 (IRE) expression and haemoglobin concentration (r=-0.680, p=0.0002), haematocrit (r=-0.677, p=0.003), log SF (r=-0.569, p=0.004), and TS (r=-0.677, p=0.003) (table 3). Similar results were observed for Ireg1 (table 3). There were no significant correlations between expression of hephaestin or Dcytb and serum iron indices or haematological parameters in any of the three groups.

In order to determine if DMT1 (IRE) and Ireg1 expression were inappropriately increased in relation to body iron stores in HHC patients, the relationship between gene expression of these iron transporters and log SF was examined based on correlations observed in non-HHC patients (fig 2). Nine of 12 HHC patients and lay outside and above the predicted linear extension of the regression lines for DMT1 (IRE) and Ireg1 based on non-HHC patients, suggesting that for a given SF, DMT1 (IRE) and Ireg1 levels were higher than predicted in HHC subjects with elevated SF. The three female HHC patients had appropriate levels of DMT1 (IRE) and Ireg1 expression for their SF.

When all 36 subjects were analysed together, a significant positive correlation was found between expression of DMT1 (IRE) and Ireg1 (r=0.826, p<0.0001). This relationship was still present when HHC patients (r=0.754, p=0.005) and non-HHC patients were analysed separately (r=0.841, p<0.0001).

DISCUSSION

Recent identification of several molecules that play critical roles in iron transport across the duodenal enterocyte has greatly advanced our understanding of intestinal iron absorption. The potential roles of DMT1, Ireg1, hephaestin, and Dcytb in this pathway have been examined in animal models of disturbed iron metabolism but in relatively few human studies.

In this study, duodenal expression of DMT1 (IRE) and Ireg1 was similar in untreated HHC patients and iron replete subjects but in most of the HHC patients there appeared to an increase in DMT1 (IRE) and Ireg1 expression relative to SF. Furthermore, hephaestin and Dcytb were not upregulated in HHC and appear unlikely to play primary roles in the relative increase in iron absorption observed in HHC. As predicted, DMT1 (IRE) and Ireg1 expression was increased in iron deficient patients compared with iron replete subjects, thus acting as internal controls and confirming that the laboratory methods used in this study were appropriate. Significant positive correlations were present between DMT1 (IRE) and Ireg1

| Table 3 Correlation between biochemical iron indices, haematological parameters, and expression of DMT1 (IRE) and Ireg1 in non-HHC subjects and patients with HFE related haemochromatosis. |
|-----------------|-----------------|-----------------|-----------------|-----------------|
|                 | Hb              | Hct             | Log SF           | TS              |
| Non-HHC subjects (n=23) |                 |                 |                 |                 |
| Log DMT1 (IRE) | r: -0.680       | p: 0.002        | 0.003           | 0.004           | 0.003           |
|                  | p: 0.002        | 0.003           | 0.004           | 0.003           |
| Log Ireg1       | r: -0.589       | p: 0.002        | 0.003           | 0.004           |
|                  | p: 0.002        | 0.003           | 0.004           |
| HHC subjects (n=12) |                 |                 |                 |                 |
| Log DMT1 (IRE) | r: 0.069        | p: 0.820        | 0.450           | 0.002           | 0.881           |
|                  | p: 0.390        | 0.171           | 0.030           | 0.750           |

Hb, haemoglobin concentration; Hct, haematocrit; TS, transferrin saturation; SF, serum ferritin concentration; HHC, hereditary haemochromatosis DMT1, divalent metal transporter 1; Ireg1, iron regulated gene 1.
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expression in all patient groups. There were significant inverse correlations between DMT1 (IRE) and Ireg1 and SF in HHC patients, suggesting that body iron stores still play a role in the regulation of duodenal expression of these iron transport genes. Expression of these iron transporters was assessed by RQ-PCR only as recent studies indicate that protein levels parallel changes observed in gene expression of these molecules.

Total body iron absorption may be normal or elevated only 2–3-fold in untreated HHC subjects compared with normal controls and this appears to be predominantly due to an increase in mucosal transfer and not to an increase in mucosal uptake of iron. Despite increased body iron stores, phlebotomy in HHC patients results in a further marked increase in total body iron absorption, indicating the importance of increased erythropoiesis and plasma iron turnover in the control of iron uptake and absorption. Iron absorption frequently remains elevated for a prolonged period after completion of venesection in HHC. In the original study of intestinal gene expression performed by Zoller et al, the 20 HHC patients were heterogenous. Six of these patients had completed treatment or were undergoing venesection at the time of the study and DMT1 expression was reported to be increased 100-fold. Similarly, in a recent study by Rolfs et al, five of nine subjects with HHC were fully treated and the remaining four subjects were being venesected. Venesence is likely to have contributed to the increase in DMT1 expression by Zoller et al and for such an effect was provided in a subsequent study by Zoller et al who reported only a 2–10-fold increase in DMT1 and Ireg1 expression in eight untreated HHC patients with biochemical iron overload compared with normal controls. This marked variation in the reported increase in DMT1 expression in HHC patients and conflicting data on iron transporter expression in animal models of iron overload for the need for iron overload for studies of iron transporters in untreated human HHC patients. In contrast with previous reports, an increase in the absolute level of DMT1 (IRE) or Ireg1 expression in untreated HHC patients was not demonstrated in this study. However, whether the level of expression of these iron transport genes was inappropriate high for the degree of iron loading was warranted further consideration. SF, a laboratory indicator of body iron stores, was significantly associated with expression of both DMT1 (IRE) and Ireg1 in non-HHC subjects. Figure 2A and 2B show that all but three of the HHC patients lay beyond and above the regression lines relating SF and DMT1 (IRE) and Ireg1 levels in non-HHC subjects, suggesting that for a given SF, DMT1 (IRE) and Ireg1 levels were higher than predicted in most HHC subjects. For this comparison, we have ascertained the relationship between SF and iron overload for these iron transport genes over all values of SF. These results support early HHC studies showing that although the absolute value of intestinal iron absorption is frequently not increased compared with normal controls, it is inappropriately increased in relation to their SF. Similarly, these findings are relevant in the context of previous studies investigating markers of intracellular iron status in duodenal tissue from patients with HHC. These studies showed that although the duodenal mucosal content of ferritin mRNA and protein is relatively low and the corresponding levels of transferrin receptor gene and protein expression are relatively high in patients with HHC, absolute levels were similar to those observed in normal control subjects.

Furthermore, in HHC patients there were significant linear relationships between SF and DMT1 (IRE) and Ireg1 expression. In addition, DMT1 (IRE) expression showed a positive correlation with Ireg1 expression in HHC and non-HHC patients. Iron absorption studies have shown a similar inverse relationship between SF and whole body iron absorption in subjects with HHC. Collectively, these results suggest that in HHC patients, changes in duodenal transport gene expression and consequently intestinal iron absorption are still subject to some regulation by body iron stores. In the current study, patient numbers were too small to examine the relationship between hepatic iron stores and duodenal expression of the iron transport genes.

Interestingly, three HHC patients had appropriate levels of DMT1 and Ireg1 for their SF. All three subjects were female with normal SF. It is interesting to speculate whether phenotypic non-expression in these three patients was a result of environmental factors or whether they have constitutively lower levels of expression of iron transporters. Further studies of intestinal iron transporters in non-expressing C282Y homozygous HHC patients would be of particular interest in determining the contributors to their attenuated phenotype.

Dietary factors may account in part for the difference in results between this study and that of Zoller and colleagues. Iron absorption is markedly influenced by recent dietary iron intake and studies conducted in rodents support a rapid regulatory effect of increased dietary non-haem iron content resulting in decreased duodenal expression of iron transport genes.

The majority of intestinal gene expression performed by Zoller et al is reported not to possess an IRE. Expression of DMT1 in Hfe−− mice of different genetic backgrounds is reported not to possess an IRE. Expression of DMT1 in Hfe−− mice of different genetic backgrounds.

Recently, Dupic et al showed that differences in the severity of hepatic iron loading between Hfe−− mice of two different strains were probably accounted for by variations in expression of DMT1, Ireg1, and Dcytb.

There are no prior data on Dcytb protein or gene levels in human subjects with iron deficiency or HHC. The tissue Dcytb levels were similar in iron deficient patients and iron replete subjects. However, in hpx−− mice, duodenal Dcytb mRNA and protein levels increased in response to iron deficiency and anaemia. Studies from our group showed regulation of Dcytb mRNA levels by changes in dietary iron content and body iron status in rodents. However, the mechanism by which iron regulates Dcytb expression is still unclear as Dcytb is reported not to possess an IRE. Expression of Dcytb was not increased in HHC subjects, suggesting that it does not play a primary role in the increased iron absorption characteristic of HHC. In vitro studies examining mucosal surface ferric iron reducing activity in duodenal fragments taken from untreated and treated patients with HHC have shown a significant increase in duodenal ferric iron reduction and...
increased iron uptake compared with normal controls. However, in \textit{Hfe} mice, Griffith et al showed no difference in duodenal ferrireductase activity between \textit{Hfe} and wild-type animals. More recently, Dupic et al showed that \textit{Dcyt} follows a similar expression pattern to that of DMT1 and Ireg1, with marked differences observed between \textit{Hfe} knockout mice of different genetic backgrounds. The role of Dcyt in the adaptive response to iron deficiency in humans remains to be clarified.

Expression of \textit{hephaestin} was decreased in subjects with HHC compared with iron replete and iron deficient subjects but levels of \textit{hephaestin} were similar in iron deficient and iron replete subjects. It is unlikely that the decrease in \textit{hephaestin} levels is of physiological significance as the difference was less than twofold in magnitude. These results, together with those of Rolfs and colleagues,\textsuperscript{11} suggest that changes in \textit{hephaestin} expression cannot account for the increased iron absorption characteristic of HHC. Duodenal expression of \textit{DMT1} (non-IRE) was negligible and there were no observable differences in expression between the three groups. From these results and those of others,\textsuperscript{12} it appears unlikely that \textit{DMT1} (non-IRE) plays a major role in the changes observed in duodenal DMT1 gene levels.

In summary, this study supports the primary roles of \textit{DMT1} and \textit{Ireg1} in mediating increased iron absorption in human subjects with iron deficiency. Duodenal expression of these iron transport genes was inversely regulated by body iron stores in iron deficient and iron replete subjects. Although we did not find an absolute increase in expression of \textit{DMT1} (IRE) or \textit{Ireg1} in a group of untreated C282Y homozygous HHC patients, there appears to be a relative increase in the levels of these iron transport genes for any given serum ferritin concentration in most HHC patients. In these patients, \textit{DMT1} (IRE) and \textit{Ireg1} levels were inversely proportional to serum ferritin concentration, suggesting that body iron stores still have some regulatory effect on expression of these iron transport genes in HHC.

**ACKNOWLEDGEMENTS**

RAS is a recipient of a National Health and Medical Research Council Postgraduate Medical Scholarship. LMF and DHC are affiliated with the University of Queensland. This study was supported by grants from the National Health and Medical Research Council, the Queensland Liver Transplant Service, and the Princess Alexandra Hospital Research and Development Foundation. The authors wish to thank Dr Ross Campbell and Dr Andrew Evered for their advice and assistance.

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Clinical Evidence also needs to recruit a number of new peer reviewers specifically with an interest in the clinical areas stated above, and also others related to general practice. Peer reviewers are health care professionals or epidemiologists with experience in evidence based medicine. As a peer reviewer you would be asked for your views on the clinical relevance, validity and accessibility of specific topics within the journal, and their usefulness to the intended audience (international generalists and health care professionals, possibly with limited statistical knowledge). Topics are usually 2000–3000 words in length and we would ask you to review between 2–5 topics per year. The peer review process takes place throughout the year, and our turnaround time for each review is ideally 10–14 days.

If you are interested in becoming a peer reviewer for Clinical Evidence, please complete the peer review questionnaire at www.clinicaledge.com or contact Claire Folkes (cfolkes@bmjgroup.com).