

Interleukin 1 β induces gastric epithelial cell matrix metalloproteinase secretion and activation during *Helicobacter pylori* infection

M Gööz, M Shaker, P Gööz, A J Smolka

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See end of article for authors' affiliations

Correspondence to:
Dr A J Smolka, Department of Medicine CSB 921E, Medical University of South Carolina, 96 Jonathan Lucas St, Charleston, SC 29425, USA; smolkaaj@musc.edu

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Background and aims: Matrix metalloproteinases (MMPs) are endopeptidases with roles in extracellular matrix remodelling, cell proliferation, and inflammatory processes. We showed previously that *Helicobacter pylori* infection of human gastric adenocarcinoma (AGS) cells increased epithelial secretion of epithelial MMP-1 and MMP-3 and bacterial secretion of MMP-3-like activity. In the present study, we sought to characterise the role of interleukin (IL)-1 β in *H pylori* induced secretion of epithelial MMPs.

Methods and results: AGS cells were treated with *H pylori* and/or IL-1 β . Comparable IL-8 secretory responses (~1700 ng/ml) measured by ELISA were induced by 2.0 ng/ml IL-1 β and by *H pylori* at a multiplicity of infection (MOI) of 50. The same IL-1 β and *H pylori* concentrations induced comparable increases in AGS cell caseinolytic activity at 60 kDa. MMP-3 monoclonal antibody immunoblots of AGS cell conditioned media detected immunoreactive bands at 71 kDa and 56 kDa. *H pylori* (MOI=50-100) induced dose dependent increases in both bands whereas IL-1 β (0.2-2 ng/ml) induced dose dependent increases only in the 71 kDa band, which was identified as a MMP-3/TIMP-3 (tissue inhibitor of metalloproteinases 3) heterodimer. AGS/*H pylori* conditioned media expressed 24 times more MMP-3 activity than AGS/IL-1 β conditioned media. There was a strong interaction between IL-1 β and *H pylori* on MMP-3 secretion.

Conclusions: We conclude that IL-1 β induces gastric epithelial cell MMP-3 secretion, contributing to epithelial tissue destruction during *H pylori* infection. However, other bacterial/host factors are needed to mediate the full gastric epithelial cell MMP-3 secretory response induced by *H pylori* infection.

Helicobacter pylori is a causative agent of superficial gastritis, gastric and duodenal ulcers, atrophic gastritis, and gastric cancer.¹ Gastric colonisation by *H pylori* provokes host inflammatory responses associated with elevated levels of mucosal cytokines (for review see Crabtree²). The inflammatory interleukins IL-1 β , IL-6, and IL-8, and tumour necrosis factor α (TNF- α), have been detected in patients infected by cytotoxin associated protein (*cagA*⁺) strains of *H pylori*,³ and in vitro cultures of antral gastric biopsies from *H pylori* infected patients.⁴ In vitro studies with human gastric cell lines have confirmed in vivo observations that *H pylori* stimulates IL-8 secretion by gastric epithelial cells.⁵

Gastric colonisation by *H pylori* also induces a transient period of achlorhydria.² Factors contributing to altered gastric acid secretion during *H pylori* infection include urease activity,⁶ a soluble protein of 46 kDa,⁷ and certain fatty acid components of *H pylori* lipopolysaccharide.⁸ In addition, *H pylori* induced achlorhydria may also be mediated by IL-1 β which has been shown to inhibit gastric acid secretion in rats.⁹ We have shown that acute *H pylori* infection of gastric epithelial cells transiently transfected with human H,K-ATPase α subunit promoter downregulates promoter activity,¹⁰ a finding which may correlate mechanistically with decreased acid output in acute *H pylori* infection in vivo.

Bacterial factors also mediate gastric epithelial damage induced by *H pylori* infection.¹¹ Strain specific bacterial factors include *cagA*, the protein product of the *cagA* gene, which is injected into host cells during *H pylori* infection and stimulates dephosphorylation of host-cell proteins.¹² Other bacterial factors are the vacuolating toxin (VacA)¹³ and the protein product of the *iceA* gene¹⁴ which are associated with more severe clinical outcome.

As *H pylori* strains with similar known virulence markers can induce several distinct patterns of inflammation,¹⁵ the roles of other bacterial and host factors in mediating *H pylori* induced gastroduodenal epithelial damage are of considerable interest. Matrix metalloproteinases (MMPs) are extracellular matrix degrading endopeptidases with roles in inflammatory processes, cell migration, and tumour growth and metastasis.¹⁶ In normal human and rabbit gastric mucosa, MMP-1, MMP-2, MMP-9, and tissue inhibitor of metalloproteinases 2 (TIMP-2) have been detected using immunocytochemistry.¹⁷ Enhanced expression of matrilysin, collagenase, and stromelysin-1 (MMP-3) in gastrointestinal ulcers has been reported.¹⁸ We have shown that *H pylori* infection of AGS cells increases secretion of epithelial cell MMP-1 and MMP-3 and enhances secretion of TIMP-3.¹⁹ Furthermore, our data indicated that bacterial MMP-3-like activity and TIMP-3-like immunoreactivity contribute to the host-cell response.¹⁹

As noted above, IL-1 β has been implicated in the development of a gastric inflammatory response and transient achlorhydria,² and plays roles in several pathways leading to gastric injury and tissue destruction. Secreted MMP-1 and MMP-3 levels are upregulated by IL-1 β in human intestinal mucosa²⁰ and in fibroblasts.²¹ Furthermore, polymorphisms of the human IL-1 β gene promoter are associated with an

Abbreviations: IL, interleukin; TNF- α , tumour necrosis factor α ; MOI, multiplicity of infection; MMPs, matrix metalloproteinases; TIMP, tissue inhibitor of metalloproteinases; vacA, vacuolating cytotoxin; *cagA*, cytotoxin associated protein; PBS, phosphate buffered saline; ELISA, enzyme linked immunosorbent assay; SDS-PAGE, sodium dodecyl sulphate-polyacrylamide gel electrophoresis.

increased risk of gastric cancer,²² and involvement of IL-1 β in mucosal apoptosis induced by *H pylori* has also been proposed.¹⁵ As both MMPs and the cytokine cascade are implicated in tissue destruction and are activated during *H pylori* infection, in the present study we tested the hypothesis that activation of gastric epithelial MMPs by *H pylori* is mediated by the proinflammatory cytokine IL-1 β .

MATERIALS AND METHODS

Cell culture media and additives were acquired from Cellgro Mediatech (Herndon, Virginia, USA), and fetal bovine serum was obtained from Atlanta Biologicals (Norcross, Georgia, USA). TIMP-3 (AB802) polyclonal antibody and MMP-3 (MAB 1339) monoclonal antibody were purchased from Chemicon Inc. (Temecula, California, USA). Secondary antibodies were obtained from Rockland Immunochemicals Inc. (Gilbertsville, Pennsylvania, USA) and Jackson ImmunoResearch Laboratories Inc. (West Grove, Pennsylvania, USA). Control MMPs and TIMPs prepared from human skin fibroblast conditioned media were obtained from Sigma (St Louis, Missouri, USA). Fluorogenic substrates for measurement of MMP enzyme activity were purchased from Bachem Inc. (Torrance, California, USA). All other reagents were of molecular biology grade or the highest grade of purity available.

Cells and bacteria

Human gastric adenocarcinoma cells (AGS cells, ATCC CRL 1739) were maintained in AGS medium (Ham's F-12, 10% fetal bovine serum, 100 units/ml penicillin G, 0.25 μ g/ml amphotericin B, 100 μ g/ml streptomycin) at 37°C in a 5% CO₂/95% air incubator and used between passages 42 and 56. A strain of *H pylori* positive for vacuolating cytotoxin (*vacA*⁺), cytotoxin associated protein (*cagA*⁺), and urease (ATCC number 49603) was cultured on 5% horse blood agar plates (Remel; Lenexa, Kansas, USA) incubated at 37°C in sealed microaerophilic pouches (BBL Campy Pouch; Becton Dickinson, Cockeysville, Maryland, USA). Cultures were routinely screened for urease activity.

Cell treatments

For *H pylori* infection of AGS cells, *H pylori* were harvested between 48 and 72 hours after inoculation of agar plates, resuspended in sterile phosphate buffered saline (PBS), and counted by absorbance at 600 nm (1 OD_{600nm} = 2.4 \times 10⁸ colony forming units/ml). AGS cells (2.5 \times 10⁶) were seeded into T-75 flasks and infected at 90% confluence with *H pylori* at multiplicities of infection (MOI) of 50–100 or incubated with IL-1 β (0.2–20 ng/ml) in serum free Ham's F12 culture medium for 24 hours. Controls included uninfected/untreated AGS cells and *H pylori* alone (in the same amounts as used for infection) which were cultured in serum free medium for the same length of time. Conditioned media were centrifuged for 10 minutes at 4000 *g* to remove cells and bacteria, and serum free media were concentrated 100–130 \times by ammonium sulphate precipitation. Unconditioned serum free medium was concentrated and used as a negative control. Media aliquots were stored at –70°C until used.

Measurement of IL-8 secretion

IL-8 content of culture media conditioned by untreated, IL-1 β treated, or *H pylori* infected AGS cells was measured using an IL-8 enzyme linked immunosorbent assay (ELISA) kit according to the manufacturer's instructions (R&D Systems Minneapolis, Minnesota, USA).

Zymography

Concentrated AGS cell culture media were mixed with sodium dodecyl sulphate- polyacrylamide gel electrophoresis (SDS-PAGE) sample buffer (10% glycerol, 2% SDS, 63 mM Tris, pH 7.0) without reducing agent and applied to non-reducing 10%

acrylamide gels containing 0.1% gelatin, or to non-reducing 4–16% acrylamide gels containing 0.1% casein (NuPAGE; Novex, Encinitas, California, USA). Electrophoresis was carried out for 90 minutes at 125 V at room temperature, and resolved proteins were renatured in situ by immersing the gels in 2.7% (w/v) Triton-X 100 for 30 minutes at room temperature. The gels were then rinsed in zymogram developing buffer (Novex) for 30 minutes and incubated overnight at 37°C in the same buffer. Gelatinolytic or caseinolytic activity in the gels was visualised as negative staining with Coomassie Brilliant Blue.

Enzyme activity assay

MMP activity of unconcentrated and 100–130 \times concentrated serum free media conditioned by untreated, IL-1 β treated, and/or *H pylori* infected AGS cells, or by *H pylori* culture, was measured using the fluorogenic MMP-3 specific synthetic peptide substrate Mca- Arg-Pro-Lys-Pro-Val-Glu-Nva-Trp-Arg-Lys(Dnp)-NH₂. Substrate fluorescence evoked by metalloproteinase activity was measured for 200 seconds in an Aminco-SLM DW2 spectrofluorimeter (excitation 328 nm, emission 399 nm) with a slit width of 1 nm and sensitivity of 800 V.

Immunoprecipitation

MMP-3 antibody was covalently coupled to cyanogen bromide activated agarose beads (Sigma, St Louis, Missouri, USA) in binding buffer (0.1 M NaHCO₃/0.5 M NaCl, pH 7.6) for two hours at room temperature. Reactive sites were blocked with 0.1 M Tris buffer (pH 8.0) for one hour at room temperature, and the beads were washed with 50 mM diethylamine and then with PBS (pH 7.4). Coupling efficiency was assessed by comparison of antibody banding densities on SDS-PAGE gel before and after the coupling reaction. Concentrated conditioned media from IL-1 β treated and *H pylori* infected AGS cell cultures were desalted by dialysis against non-denaturing buffer (50 mM Tris HCl, pH 7.4, 150 mM NaCl, 1% Triton X-100, 0.5% Na-deoxycholate, 1 mM EDTA, 0.1% SDS, 10 μ l/ml protease inhibitor cocktail III (Calbiochem, San Diego, California, USA), and 1 mM PMSF). Samples were incubated with the antibody coupled beads in non-denaturing buffer at 4°C overnight with constant rotation. After removal of supernatants, the beads were washed with binding buffer, eluted with 50 mM diethylamine, and the eluents analysed by TIMP-3 immunoblotting.

Immunoblotting

Aliquots of concentrated media conditioned by untreated, IL-1 β treated, or *H pylori* infected AGS cells, or by *H pylori* culture, and material eluted from MMP-3 antibody coupled beads were mixed with SDS-PAGE sample buffer without reducing agent and heated at 95°C for five minutes. Electrophoresis was carried out on 4–12% or on 10% acrylamide Bis-Tris gels (NuPAGE; Novex) for 35 minutes at 200 V. Resolved proteins were transferred to 0.2 μ m nitrocellulose membranes (Osmonics, Westborough, Massachusetts, USA) and blocked with 5% non-fat dry milk for two hours. Protein replicas were washed 3 \times in TTBS (20 mM Tris, 0.5 M NaCl, pH 7.5, 0.05% Tween-20), and incubated overnight at room temperature in the appropriate dilutions of MMP or TIMP antibodies. The replicas were then washed 3 \times in TTBS and incubated for one hour at room temperature in appropriate dilutions of secondary antibodies. Immunoreactive protein bands were visualised using enhanced chemiluminescence and recorded on ECL Hyperfilm (ECL kit, Amersham Pharmacia, Piscataway, New Jersey, USA). Immunoblots shown in this study are representative replicates selected from at least three experiments.

Protein measurement

Culture medium protein concentrations were measured using Bio-Rad protein assay reagent (Bio-Rad, Hercules, California,

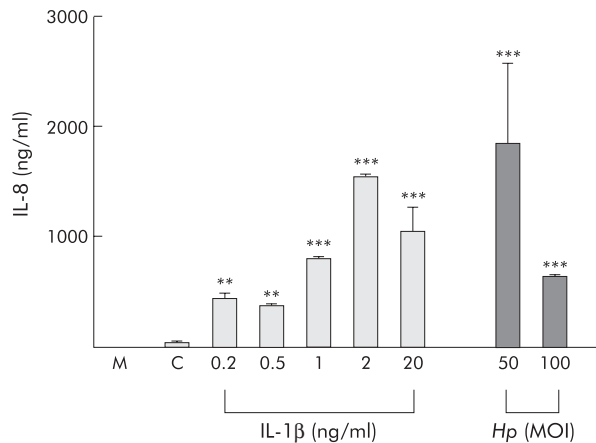


Figure 1 Interleukin (IL)-8 secretory response in AGS cells induced by IL-1 β (0.2–20 ng/ml) and *Helicobacter pylori* (Hp) (multiplicity of infection (MOI)=50 and 100), as measured by enzyme linked immunosorbent assay. M, cell culture medium alone; C, cell culture medium from untreated AGS cells. n=3, **p<0.01, ***p<0.001 versus untreated cells.

USA). The protein concentration of serum free unconditioned medium was 0.38 (0.08) mg/ml, that of AGS conditioned medium was 0.75 (0.17) mg/ml, that of infected AGS conditioned medium was 1.96 (0.29) mg/ml, and that of *H pylori* conditioned medium was 0.82 (0.21) mg/ml. Data are expressed as mean (SEM), n=6. Culture medium aliquots (5 μ l) were used for enzyme activity assays. For zymographic and immunoblot assays, 18 μ l of culture medium aliquots were applied to each lane.

Data analysis

Densitometric analysis of zymograms and immunoblots was carried out using image analysis software (Scion Corporation). Data are expressed as per cent of control activity (the densitometric intensity of uninfected AGS cell conditioned medium was set to 100%) and are shown below the gels. Enzyme activity data were analysed by linear regression and were expressed as initial reaction velocity (fluorescence unit/s). Data comparisons were made using the Student's *t* test and ANOVA, with p<0.05 regarded as significant. All experiments were carried out at least three times.

RESULTS

IL-8 secretory response induced by IL-1 β and *H pylori*

H pylori and IL-1 β have previously been shown to induce gastric epithelial cell IL-8 secretion.⁵ As a first step in clarifying the role of IL-1 β in MMP mobilisation by AGS cells, we determined the concentrations of IL-1 β and *H pylori* that would induce comparable levels of AGS cell IL-8 secretion. AGS cells were treated with either IL-1 β (0.2–20 ng/ml) or infected with *H pylori* (MOIs of 50 and 100) in serum free Ham's F-12 medium. IL-8 contents of unconcentrated conditioned supernatants were measured by ELISA. As shown in fig 1, IL-1 β dose dependently increased the IL-8 secretory response of AGS cells. The IL-8 content of medium conditioned by untreated AGS cells was 39.2 (2.1) ng/ml (fig 1C). Comparable IL-8 secretory responses (1541 (26) ng/ml and 1840 (729) ng/ml) were induced by 2.0 ng/ml IL-1 β and by *H pylori* at MOI=50, respectively. In all subsequent experiments, AGS cells were treated with these IL-1 β and *H pylori* concentrations.

Casein zymography of AGS cell conditioned media

MMP activity secreted by AGS cells during IL-1 β treatment or *H pylori* infection was assessed by electrophoresis of 130 \times concentrated serum free cell culture media through casein containing zymogram gels. To study the possible interaction

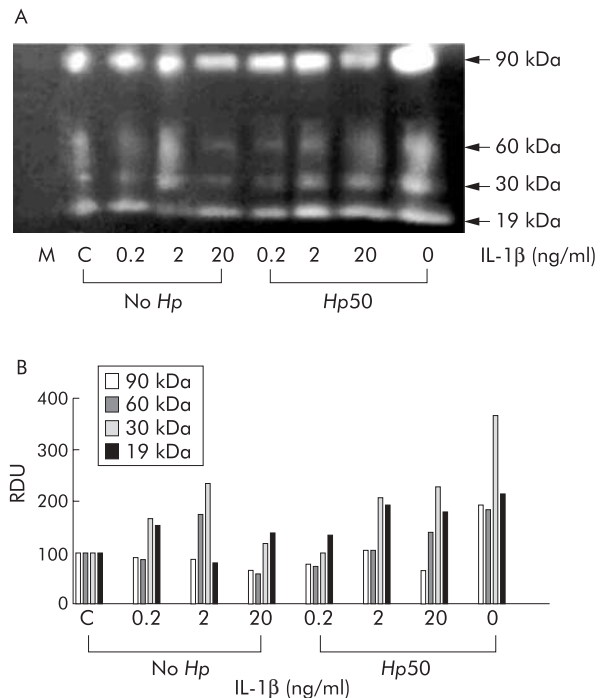


Figure 2 (A) Caseinolytic activity of interleukin (IL)-1 β treated (0.2–20 ng/ml, No Hp) and/or *Helicobacter pylori* (multiplicity of infection (MOI)= 50, Hp50) infected AGS cell conditioned media, as measured by casein zymography. Lane M, cell culture medium alone; lane C, supernatant of untreated AGS cells. Calculated molecular masses of caseinolytic bands are shown on the right. (B) Densitometric analysis of 90 kDa, 60 kDa, 30 kDa, and 19 kDa caseinolytic bands. The zymogram is a representative replicate selected from three experiments. RDU, relative densitometric units.

between IL-1 β and *H pylori*, we included cell supernatants from IL-1 β treated (0.2–20 ng/ml) and *H pylori* infected (MOI=50) cells. Figure 2 shows a typical caseinolytic zymogram together with densitometric analysis of caseinolytic gel bands. Caseinolytic activities in unconditioned medium and in medium conditioned by untreated AGS cells are shown in lanes M and C, respectively. No caseinolytic activity was detected in unconditioned medium. Culture medium from untreated AGS cells showed bands of caseinolytic activity at ~90 kDa and 19 kDa and traces of activity at ~60 kDa and 30 kDa. These data are consistent with our previous findings.³ IL-1 β (0.2–20 ng/ml) treatment dose dependently increased caseinolytic activity at 60 kDa and 30 kDa and decreased activity at 19 kDa. However, cell treatment with 20 ng/ml IL-1 β decreased caseinolytic activity at 90 kDa, 60 kDa, and 30 kDa. Densitometric analysis of caseinolytic bands in fig 2 indicated that IL-1 β and *H pylori* concentrations inducing comparable IL-8 secretory responses (2 ng/ml and MOI=50, respectively) induced comparable increases in AGS cell MMP activity at 60 kDa (175 v 183 relative densitometric units). Comparable caseinolytic activities were induced by *H pylori* at MOI of 100 and at MOI of 50 (data not shown). Medium conditioned by *H pylori* alone showed weak caseinolytic activity at 60 kDa³ (data not shown).

MMP-3 secretion by IL-1 β treated or *H pylori* infected AGS cells

We have previously shown that MMP-3 secretion by AGS cells is increased during *H pylori* infection.¹⁹ In the present study, we addressed the role of IL-1 β in induction of MMP-3 secretion. Concentrated serum free media were analysed by immunoblotting using an MMP-3 monoclonal antibody. Figure 3 shows a typical MMP-3 immunoblot together with densitometric

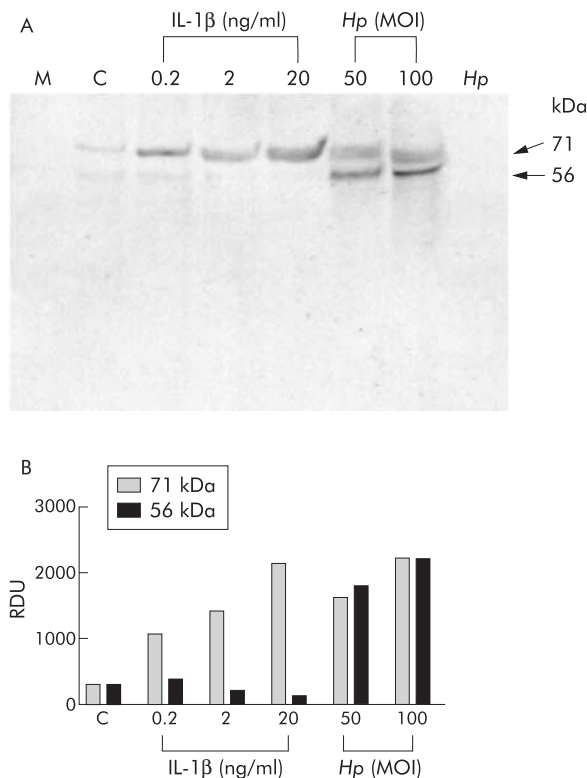


Figure 3 (A) Matrix metalloproteinase 3 (MMP-3) expression in serum free culture media conditioned by interleukin (IL)-1 β treated or *H pylori* infected AGS cells, as measured by non-reducing sodium dodecyl sulphate-polyacrylamide gel electrophoresis immunoblot analysis using MMP-3 specific monoclonal antibody. Lane M, cell culture medium alone; lane C, medium from untreated/uninfected AGS cells. (B) Densitometric analysis of immunoreactive bands. The blot is a representative replicate selected from three experiments. Hp, *Helicobacter pylori*; MOI, multiplicity of infection; RDU, relative densitometric units.

analysis of immunoreactive bands. MMP-3 immunoreactivities of unconditioned medium and in medium conditioned by untreated AGS cells are shown in lanes M and C, respectively.

In media conditioned by IL-1 β or *H pylori* treated AGS cells, immunoreactivity was detected at 56 kDa and 71 kDa. Densitometric analysis of the immunoreactive bands showed that *H pylori* treatment (MOI=50 and 100) of AGS cells induced dose dependent increases in both 56 kDa and 71 kDa bands whereas IL-1 β (0.2–2 ng/ml) treatment induced a dose dependent increase only of the 71 kDa band. Concentrated serum free media conditioned by *H pylori* alone did not show any MMP-3 immunoreactivity. Because the molecular mass of TIMP-3 is 24 kDa and because TIMP-3 is known to form SDS stable heterodimers with MMP-3,³ we hypothesised that the 71 kDa band represented an MMP-3/TIMP-3 heterodimer, and that IL-1 β treatment, like *H pylori* infection, increased both MMP-3 and TIMP-3 secretion by AGS cells. In the case of IL-1 β treatment alone, however, MMP-3 is immediately complexed with TIMP-3 and so is present mainly as a 71 kDa band on the gel. Thus the traces of 56 kDa immunoreactivity in IL-1 β treated lanes (0.2–2ng/ml) reflect the presence of uncomplexed MMP-3.

TIMP-3 secretion by IL-1 β treated or *H pylori* infected AGS cells

As a test of this hypothesis, the TIMP-3 content of culture media conditioned by IL-1 β treated or *H pylori* infected AGS cells was assessed by immunoblotting with a polyclonal TIMP-3 antibody. As shown in fig 4, TIMP-3 antibody labelled a 71 kDa band in media conditioned by IL-1 β treated or

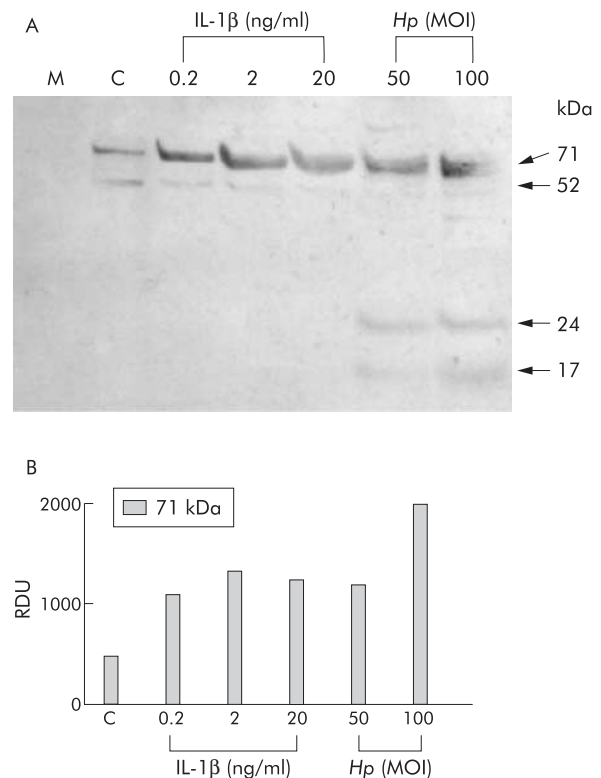


Figure 4 (A) Tissue inhibitor of metalloproteinase 3 (TIMP-3) expression in serum free culture media conditioned by interleukin (IL)-1 β treated or *H pylori* (Hp) infected AGS cells, as measured by non-reducing sodium dodecyl sulphate-polyacrylamide gel electrophoresis immunoblot analysis using polyclonal TIMP-3 antibody. Lane M, cell culture medium alone; lane C, medium from untreated/uninfected AGS cells. (B) Densitometric analysis of immunoreactive bands. The blot is a representative replicate selected from three experiments. MOI, multiplicity of infection; RDU, relative densitometric units.

pylori infected AGS cells. Densitometric analysis revealed dose dependent increases in TIMP-3 immunoreactivity of the 71 kDa band induced by IL-1 β (0.2–20 ng/ml) treatment and *H pylori* infection (MOI of 50 and 100). We have previously shown that this TIMP-3 antibody also recognises a bacterial TIMP-3-like factor.¹⁹ Thus the 71 kDa band detected in media conditioned by *H pylori* infected AGS cells reflects both human and bacterial TIMP-3 complexes. The 24 kDa and 17 kDa bands represent uncomplexed bacterial and/or human TIMP-3 immunoreactivity, and the 52 kDa band is most probably a TIMP-3 dimer. The data indicate that IL-1 β at 2 ng/ml and *H pylori* at MOI of 50, and IL-1 β at 20 ng/ml and *H pylori* at MOI of 100, induce comparable secretion of TIMP-3. This contrasts with the significantly higher secretion of MMP-3 induced by *H pylori* compared with IL-1 β (fig 2).

Co-immunoprecipitation of MMP-3 and TIMP-3

Further confirmation that the 71 kDa TIMP-3 immunoreactive bands are MMP-3/TIMP-3 complexes was obtained from co-immunoprecipitation experiments. MMP-3 complexes were immunoprecipitated from media conditioned by IL-1 β treated (2 ng/ml) or *H pylori* infected (MOI=50) AGS cells showing comparable TIMP-3 immunoreactivity. The immunoprecipitates were analysed by non-denaturing TIMP-3 immunoblotting. As shown in fig 5, more TIMP-3 immunoreactivity was detected in *H pylori* infected (MOI=50) than in IL-1 β treated (2 ng/ml) AGS cell conditioned media. This result confirms the data in fig 3 and supports the hypothesis that *H pylori* induces formation of more MMP-3/TIMP-3 complexes than IL-1 β . Because comparable TIMP-3 complexes were formed in

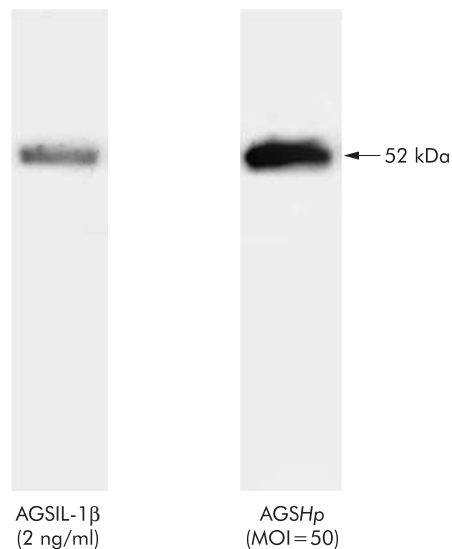


Figure 5 Co-precipitation of tissue inhibitor of metalloproteinase 3 (TIMP-3) with matrix metalloproteinase 3 (MMP-3) immunoprecipitates from interleukin (IL)-1 β treated (2 ng/ml) or *H pylori* infected (*Hp* multiplicity of infection (MOI)=50) AGS cell conditioned serum free media, as measured by non-reducing sodium dodecyl sulphate-polyacrylamide gel electrophoresis immunoblot analysis of MMP-3 monoclonal antibody immunoprecipitates using TIMP-3 polyclonal antibody. The blot is a representative replicate selected from two experiments.

both treatment conditions (fig 4), it is possible that during IL-1 β treatment enzymes other than MMP-3 (MMPs or other matrix proteases) may form complexes with TIMP-3.

MMP-3 enzyme activity in AGS cell conditioned media

The enzymatic activity of MMP-3 secreted into media conditioned by IL-1 β treated or *H pylori* infected AGS cells was measured using the synthetic fluorogenic MMP-3 specific substrate M2110. Media were concentrated 100-fold by ammonium sulphate precipitation and incubated with M2110. The rate of M2110 hydrolysis was measured as changes in emitted fluorescence as a function of time (fluorescence units/s) and was expressed as a percentage of the initial reaction velocity of MMP-3 specific protease in untreated AGS cell conditioned media (table 1). Both IL-1 β and *H pylori* elicited dose dependent increases in evoked fluorescence: however, ~24 times more MMP-3 activity was expressed in *H pylori* treated compared with IL-1 β treated AGS cell conditioned media. The combination of IL-1 β treatment with *H pylori* infection decreased the MMP-3 activity compared with *H pylori* infection alone. Statistical analysis revealed a significant ($p < 0.001$) interaction between IL-1 β treatment and *H pylori* infection.

We showed previously that *H pylori* alone secreted MMP-3-like activity and that host-cell interaction was needed for significantly increased MMP-3 activity in the cell medium.³ The latter result is consistent with data in the present study—that is, *H pylori* induced MMP-3 activity is significantly higher than IL-1 β induced MMP-3 activity. Furthermore, these data were consistent with immunoblot data showing higher MMP-3 immunoreactivity in *H pylori* infected compared with IL-1 β treated AGS cell conditioned media.

DISCUSSION

In this study, we tested the hypothesis that activation of gastric epithelial matrix metalloproteinases by *H pylori* infection is mediated by the proinflammatory cytokine IL-1 β . In an earlier study, we showed that gastric epithelial cells secrete MMP-1 and MMP-3 and that *H pylori* infection increases

Table 1 Effects of interleukin (IL)-1 β and *Helicobacter pylori* on matrix metalloproteinase 3 (MMP-3) activity of AGS cell conditioned media

Sample	Initial reaction velocity (% of AGS)
AGS	100 (5)
AGS+IL-1 β (0.2 ng/ml)	144 (26)**
AGS+IL-1 β (2 ng/ml)	149 (20)**
AGS+IL-1 β (20 ng/ml)	199 (11)**
AGS+ <i>Hp</i>	2375 (11)**
AGS+ <i>Hp</i> +IL-1 β (0.2 ng/ml)	2042 (12)**
AGS+ <i>Hp</i> +IL-1 β (2 ng/ml)	1662 (6)**
AGS+ <i>Hp</i> +IL-1 β (20 ng/ml)	1944 (7)**
<i>Helicobacter pylori</i>	291 (39)**
Medium	11 (8)**

Hydrolysis of M2110, an MMP-3 specific synthetic fluorogenic substrate, by concentrated serum free culture media of IL-1 β treated (0.2–20 ng/ml) and/or *H pylori* infected (MOI=50) AGS cells and bacterial supernatant alone (*Hp*).

Data are expressed as a percentage of the initial reaction velocity of MMP-3 activity in untreated AGS cell conditioned medium (mean (SEM)).

Medium refers to cell culture medium alone and AGS refers to untreated/uninfected AGS cell conditioned medium (n=3).

MMP-3 activity of every sample was significantly different from AGS (** $p < 0.01$). Interaction between IL-1 β and *H pylori*, as calculated by two way ANOVA, was $p < 0.001$.

secretion of both metalloproteinases.¹⁹ *H pylori* infection is known to induce IL-8 secretion by gastric epithelial cells, a response which is also elicited by cell stimulation with IL-1 β .²³ In the present study, gastric epithelial cell MMP secretion was measured in response to an IL-1 β stimulus inducing the same amplitude of IL-8 secretory response as the *H pylori* MOIs required to activate gastric epithelial MMP secretion. This IL-1 β stimulus was within the range of IL-1 β concentrations reported in studies of transcriptional factor nuclear factor κ B activation of gastric cancer cells during *H pylori* infection²⁴ or MMP activation by cytokines.^{25–27}

Induction of MMP-3 secretion was investigated because although gastric epithelial cell MMP-1 secretion was increased during *H pylori* infection,¹⁹ the significantly higher rates of MMP-3 over MMP-1 secretion suggested that stromelysin-1 (MMP-3) and not MMP-1 is the major contributor to gastric epithelial degradation. Stromelysin-1 displays a wide spectrum of proteolytic activity against extracellular matrix proteins, and so increased MMP-3 activity would be expected to accelerate stromal degradation.²⁸ The present study comparing caseinolytic activity of *H pylori* or IL-1 β treated gastric cell conditioned media showed that comparable MMP-1/MMP-3 activity ratios are induced by both treatments. The MMP-3 specific immunoblot data, immunoprecipitation data, and enzyme activity data all indicate that IL-1 β dose dependently increased gastric epithelial cell MMP-3 secretion, providing a mechanistic basis for augmented epithelial degradation.

Several previous studies have documented regulation of TIMP-3 expression by IL-1 β . Gene expression of TIMP-3 was induced by IL-1 β in human synovial lining cells²⁹ and in rat granulosa cells.²⁵ In contrast, IL-1 β downregulated TIMP-3 expression in neonatal rat ventricular cells,²⁶ in brain microvascular endothelial cells,²⁷ and in human endometrial stromal cells.³⁰ The present study provides evidence that IL-1 β regulates TIMP-3 expression in human gastric epithelial cells. In addition to increasing MMP-3 secretion, IL-1 β also elicited a dose dependent increase in TIMP-3 secretion by gastric epithelial cells. Most of the secreted MMP-3 was complexed by TIMP-3, leading to inhibition of stromelysin-1 activity.²⁸ In the present study, *H pylori* infected AGS cell conditioned media expressed higher levels of TIMP-3 than IL-1 β treated AGS cell conditioned media. The polyclonal TIMP-3 antibody used in

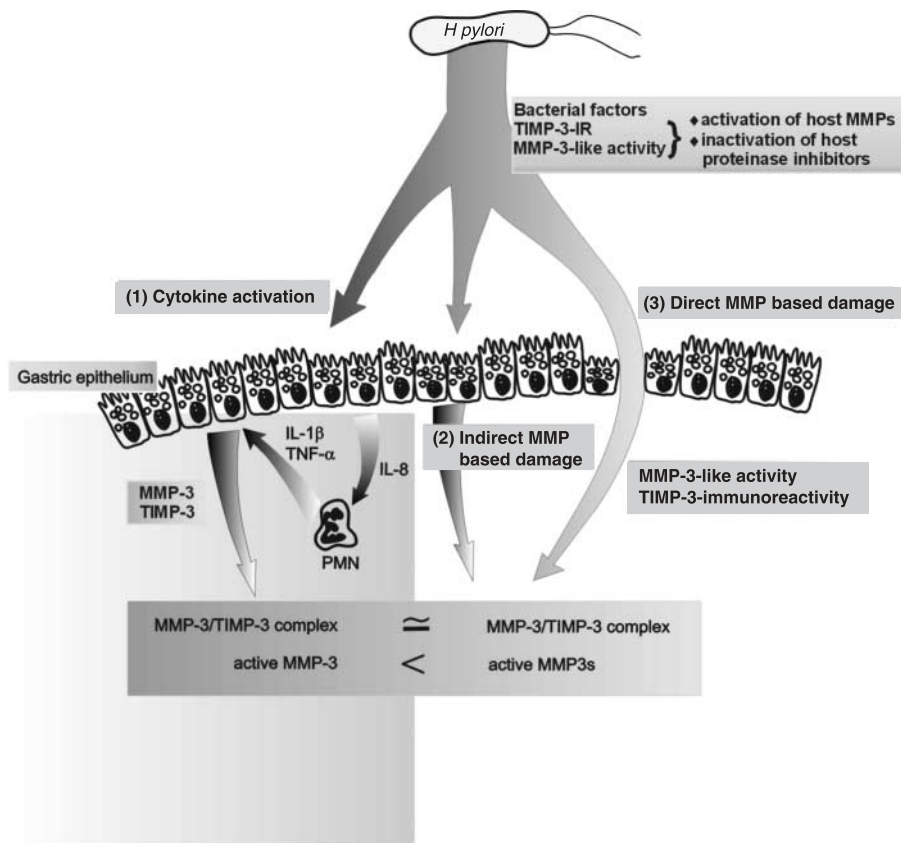


Figure 6 A model of gastric mucosal interactions between matrix metalloproteinases (MMPs) and their inhibitors (TIMPs) with the cytokine cascade during *Helicobacter pylori* infection. *H. pylori* induces epithelial MMP-3 secretion by: (1) interleukin (IL)-8, tumour necrosis factor α (TNF- α), and IL-1 β dependent mechanisms; (2) cytokine independent mechanisms; and (3) is directly enhanced by *H. pylori* metalloproteinases. Epithelial MMP-3 activity stimulated by IL-1 β alone is effectively titrated by MMP-3/TIMP-3 complex formation. However, the increased MMP-3 tissue level induced by *H. pylori* infection is incompletely titrated by increased TIMP-3 secretion, indicating that IL-1 β alone is necessary but insufficient to mediate the full gastric epithelial cell MMP activation response induced by *H. pylori* infection.

this study was previously shown to cross react with *H. pylori* proteins.¹⁹ However, as shown by our immunoblot and enzyme activity data, the increased amount of TIMP-3 did not complex all available human immunoreactive MMP-3, and did not inhibit the increased human and bacterial MMP-3 enzymatic activity. Because MMPs and TIMPs are critical for tissue remodelling, and disturbances in MMP/TIMP balance play important roles in pathophysiological processes such as stromal degradation, our data showing IL-1 β and *H. pylori* modulation of MMP-3 and TIMP-3 expression in gastric epithelial cells indicate potential *H. pylori* mediated mechanisms of gastric epithelial damage. Figure 6 is a schematic diagram incorporating the present MMP and TIMP data into the gastric mucosal cytokine cascade during *H. pylori* infection.

At least two mechanisms may account for the observed higher efficiency of *H. pylori* in stimulating MMP-3 activity compared with IL-1 β . Firstly, the high MMP-3 activity detected in *H. pylori* treated AGS cell conditioned media reflects both human MMP-3 and *H. pylori* MMP-3-like activity. As the MMP-3 monoclonal antibody used in this study does not cross react with any *H. pylori* proteins,¹⁹ the immunoreactive bands represent only MMP-3 secreted by AGS cells; however, *H. pylori* MMP-3-like activity is also measured in the specific fluorogenic substrate based activity assay. The *H. pylori* MMP-3-like proteinase potentially contributes to activation of host MMPs and activation of chemotactic pathways, while the *H. pylori* TIMP-3-like activity¹⁹ inactivates host protease inhibitors. Alternatively, or concomitantly, MMP-3 activation may be promoted by *H. pylori* induced activation of cytokines other than IL-1 β , including GRO/CINC-1,³¹ and TNF- α .³² As a putative *H. pylori* pathogenicity factor, MMP-3 complements two

previously described *H. pylori* metalloproteinases, one of which is similar to *Vibrio cholerae* mucinase³³ and the other a Zn dependent 200 kDa metalloproteinase.³⁴

The cytokine-chemokine network and matrix metalloproteinases are increasingly understood to converge and cooperate in tumour cell metastasis and cancer cell resistance. Although MMPs are recognised as important regulatory proteases of chemokines,^{16,35} the relative contributions of inflammatory cytokines to activation of host MMPs are complex and as yet poorly defined.^{24,32,36} In addition to inducing IL-8 secretion, IL-1 β and *H. pylori* also affect proliferation of gastric epithelial cells in culture,³⁷ although data concerning the direct effects of IL-1 β in *H. pylori* infection in vivo are conflicting.^{38,39}

In summary, the present study provides further evidence of interaction between cytokines and MMPs in promoting host responses to *H. pylori* infection. The proinflammatory cytokine IL-1 β , secretion of which is activated during gastric *H. pylori* infection as part of the cytokine cascade, contributes to enhanced MMP-3 and TIMP-3 secretion by *H. pylori* infected gastric epithelial cells. Although most of the secreted MMP-3 is complexed by endogenous tissue inhibitor of MMP-3, the remaining balance of active MMP-3 may contribute to the mucosal damage during *H. pylori* infection. In addition to IL-1 β , other bacterial factors including bacterial MMP-3 are needed to mediate the full gastric epithelial cell MMP activation response induced by *H. pylori* infection.

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Authors' affiliations

M Gööz, M Shaker, A J Smolka, Gastroenterology and Hepatology Division, Department of Medicine, Medical University of South Carolina,

Charleston, SC, USA

P Gööz, Rheumatology and Immunology Division, Department of Medicine, Medical University of South Carolina, Charleston, SC, USA

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