Interstitial pneumonia associated with autoimmune pancreatitis

We read with interest the article by Kamisawa et al reporting IgG4 positive plasma cells in peripancreatic tissue, extra-hepatic bile duct, gall bladder, and salivary gland (Gut 2003; 52:683-7). The association of retroperitoneal fibrosis and sclerosing pancreatitis with IgG4 bearing plasma cells in the tissues of both lesions has been also reported.1

We would like to report the first case of interstitial pneumonia associated with autoimmune pancreatitis and IgG4 positive plasma cells in the interstitium.

Hyperamylasemia was detected in a routine blood examination in a 63 year old man who had been treated for duodenal ulcer at a clinic. He was admitted to our hospital for further examination. He did not complain of epigastralgia or back pain. Serum amylase was 323 (39-130 IU/l), IgG was elevated to 2350 (800-1600 mg/dl), and IgG4 was 1690 (<80 mg/dl). Antinuclear antibody, anti-SS-A antibody, anti-SS-B antibody, rheumatoid factor, and antismooth muscle antibody were all negative. Abdominal ultrasonography and computed tomography (CT) showed swelling of the head and tail of the pancreas. Endoscopic retrograde pancreatography showed irregular narrowing of the main pancreatic duct in the head and tail. Magnetic resonance cholangiography showed stenosis of the lower common bile duct. The patient was diagnosed with autoimmune pancreatitis but he refused steroid therapy and was followed as an outpatient.

Three months later, hometocyming of the bilateral lower lung field was detected in a follow up abdominal CT. Chest CT revealed ground glass attenuation in the middle and lower lobe, and hometocyming predominantly at the back of the lower lobe, bilaterally (fig 1A). (Figure 1 (A-D) is available for viewing online at http://gut.bmjournals.com/cgi/eletters/52/5/683 #127.) Retrospectively, a slight reticulon shadow in the lower lung field was detected in the chest roentogenogram taken at the first admission but the lesion had progressed over three months. He was readmitted for further examination. He had a history of smoking 30-40 cigarettes a day for approximately 40 years. IgG was 3934 mg/dl, IgG4 was 2690 mg/dl, KL-6 was 1440 (<350 u/ml), serum amylase was 142, and lipase was 121 (0-49 IU/l). Schirmer’s test indicated a decrease in lacrimal secretion. Swelling of the head and tail of the pancreas were not changed on abdominal ultrasonography and CT.

With gallium scintigraphy, uptake was observed bilaterally at the back of the lower lobe, suggesting active pneumonia. Histology obtained by transbronchial lung biopsy from segment 8a of the right lobe showed marked thickening of the alveolar septum with marked infiltration of plasma cells and lymphocytes (fig 1B).

Immunostaining with IgG4 was performed using the immunoperoxidase method (mouse antihuman IgG4; ICN Biomedicals, Inc, Ohio, USA). Infiltration of IgG4 positive plasma cells was pancreas, peripancreatic tissue, extra-hepatic bile duct, gall bladder, salivary gland, pancreatic duct in the head and tail.

References


Author’s reply

We thank Dr Taniguchi et al for the interesting presentation of interstitial pneumonia associated with autoimmune pancreatitis (AIP).

We have experienced 24 cases of AIP but no cases showed interstitial pneumonia currently. Recently, we histopathologically examined the organs of eight patients with AIP using anti-IgG4 antibody. IgG4 positive plasmacytic infiltration was detected in the portal area of the liver, gastric mucosa, colonic mucosa, and bone marrow as well as in the pancreas, peripancreatic tissue, extra-hepatic bile duct, gall bladder, salivary gland, pancreatic duct in the head and tail.

Macrophages in the alveoli are considered to be due to smoking which often coexist with interstitial pneumonia in smokers.2

Because interstitial pneumonia associated with autoimmune pancreatitis was strongly suggested, prednisolone (40 mg/day) was administered for two weeks and then the dose was tapered. Chest CT taken two weeks after treatment showed that the ground glass attenuation in the middle and lower lobe had disappeared whereas the hometocyming remained (fig 1D).

Abdominal ultrasonography performed two weeks after treatment showed a marked decrease in the swelling the pancreas.

In the present case, infiltration of IgG4 positive plasma cells in the interstitium strongly suggests that the interstitial lung disease was associated with autoimmune pancreatitis. Interstitial pneumonia associated with Sjogren’s syndrome is unlikely in this case although there was decreased lacrimal secretion. Sica syndrome observed in a renal transplant patient is distinguishable from classical Sjogren’s syndrome in that it is negative for anti-SS-A or anti-SS-B antibodies, serum IgG4 is elevated, and infiltration of IgG4 positive plasma cells in the salivary glands is observed.

Autoimmune pancreatitis, in some cases, may be part of a systemic disease associated with IgG4.

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and lymph nodes of patients with AIP. However, few IgG4 positive plasma cells were observed in identical control specimens. From these findings, we proposed a new clinicopathological entity of IgG4 related autoimmune disease, and stressed that AIP is not simply pancreatitis but a pancreatic lesion involved in this systemic autoimmune disease. As IgG4 positive plasmacytic infiltration was observed in the transbronchially biopsied pulmonary specimens of the patient with AIP (unpublished data), it is likely that interstitial pneumonia occurs in association with AIP.

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References


The diagnostic dilemmas in discrimination between pancreatic carcinoma and chronic pancreatitis

In response to the letter of Harlozinska-Szymyka and Strutyńska-Karpinska (Gut 2004;53:469–70) commenting on our study, we agree with the remarks made in relation to the difficulties in discriminating between chronic pancreatitis and adenocarcinoma using currently employed diagnostic imaging and tumour marker analysis. Our study was aimed at determining the risk of cancer development in patients with proven chronic pancreatitis, examining age and sex standardised incidence ratios calculated from the number of observed cases of pancreatic cancer in our cohort of 373 patients with predominantly alcohol related chronic pancreatitis to the number of cases expected in the National Cancer Registry. Our study design did not take into consideration diagnostic dilemmas and focused purely on cancer risk in our cohort of patients using defined stringent criteria. Indeed, we previously underlined the interest of biological markers to this situation (for example, CA19–9 and circulating K-ras)3; however these markers have problems with both sensitivity and specificity.

We acknowledge that given the difficulties in diagnosing cancer in this situation, the establishment of new tumour markers such as tissue polypeptide specific antigen (TPS)4 with proven good sensitivity and specificity should provide for progress in the future. It has to be stressed however that TPS, a marker of proliferation activity, is not specific to pancreatic cancer, and other digestive and non-digestive cancers as well as benign chronic disorders may have high levels of this marker. Thus validated data concerning tumour markers, either alone or in combination, in distinguishing pancreatic cancer from chronic pancreatitis should prove important in diagnostic situations.

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In search of the correct strategy for preventing the spread of HCV infection

Hepatitis C virus (HCV) infection is an emerging global healthcare issue. Apart from affecting approximately 3% of the world population, HCV is also a silent disease—the majority of incidences go unrecognised and are treated unknowingly as sources of infection to others.1-3 Add to that the scant information resulting from past iatrogenic transmission, aggravated by:

- extensive use of glass syringes or non-sterile syringes; and
- social conditions which may indirectly favour the spread of infection:

- general poverty,
- poor education etc.,4,5 and
- especially among females, haemodialysis procedures.

Prisons do play a significant role in the hepatitis C epidemic, especially as the same social conditions mentioned above which may facilitate spread of infection also predict imprisonment.3 In fact, a disproportional share of the burden of HCV infection is found among those who pass through correctional facilities.4 As stated previously, HCV seroprevalence is high among this group. A history of incarceration is one of the strongest associations with HCV seropositivity.5 Yet despite these high levels, reports of HCV transmission in the prison setting are uncommon.4 In fact, studies have revealed that an overwhelming number of infections are being brought into prisons via inmates who are already previously infected; drug users are most likely to become infected with HCV at the beginning of their addiction—long before being imprisoned for the first time.4-6 Yet it must be taken into account the fact that the dynamic movement of people in and out of prisons makes it very difficult to detect transmission.6 While the available data do not prove that infections are acquired in prison, they do indicate prisons as high risk settings for the spread of HCV.6

What is of great concern to us are the implications of the previous and following data:

- HCV is easily transmitted parenterally.

Patterns of transmission in Italy have shown that this infection is predominantly alcohol related chronic pancreatitis and pancreatic adenocarcinoma. Gastroenterol Clin Biol 1998:22:152–9.


Our findings confirm the high prevalence among drug users (31.8%) but the highest prevalence was found among male prisoners (37.4%), particularly those in the 30–49 year age range (47.3%). In fact, we concur with Skipper et al that “those involved with IDU frequently pass through the penal system and spend time in prisons” thus indicating a probable interrelationship between these two groups.

Our other findings from this study shed light on the unusually high prevalence of this disease in the general population of Southern Italy. Previous reports have shown that the prevalence of HCV infection is generally low in the general population of most industrialised countries (<3%). In Southern Italy, the high prevalence of HCV infection may be the result of past iatrogenic transmission, aggragated by:

- extensive use of glass syringes or non-sterile syringes; and
- social conditions which may indirectly favour the spread of infection:

- general poverty,
- poor education etc.,4,5 and
- especially among females, haemodialysis procedures.

Patterns of transmission in Italy have shown that this infection is predominantly alcohol related chronic pancreatitis and pancreatic adenocarcinoma. Gastroenterol Clin Biol 1998:22:152–9.
Our prisons are overcrowded (the Secondaglione prison located in Naples, for example, has 1350 prisoners and only 790 beds).

In Southern Italy, the health system in general is less efficient and less meticulous than that in the north of Italy (and in the rest of the EU and in the USA).2

We have no harm reduction programmes in place.

The limited availability of prevention methods has been linked to the transmission of HCV infection.3 This association, and its relation to inadequate management of a major health problem, surely opened up our National Health System to costly retaliations. If it can be proven that an inmate contracted HCV while incarcerated, due to a lack of sufficient care and prevention on the part of the system, he then has the right to seek judicial indemnification—a costly process for all concerned.

How much more economical to initiate admission screening programmes in our overcrowded prisons where, as detailed above, there is an identifiable elevated risk. By so doing, we move one step closer to correcting a problem that is grossly out of control. As HCV is associated with different kinds of cancers (liver, possibly non-Hodgkins lymphoma) and with autoimmune diseases (crigilobulinaemia, thyroiditis, Hashimoto thyroiditis), which develop after the virus has caused immune system alterations,4 routine health screening on admission to prison presents a unique opportunity to identify health needs and plan health services at an early stage.5 In fact, studies have found that screening provides a preventing efficient management of a crucial problem and sustaining the welfare of its people.

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Cross reactivity due to positive canrenone interference

Canrenone is a selective competitive inhibitor of the aldosterone receptor and a diuretic drug commonly used in the treatment of cirrhotic patients both with and without ascites. The aim of our observation was to determine if canrenone cross reacts with aldosterone in an immunoradiosorbent assay kit used for monitoring the aldosterone secretion system, according to the Child-Pugh classification for liver cirrhosis.6 Canrenone, for in vitro experiments, was a gift from GInEne Pharma (Group Therabel) Diagnostics. Canrenone, potassium canrenoate, and spironolactone are often used with digoxin in clinical practice and can cause false positive test results in common assays for digoxin (that is, Asym MEIA-Abbott) due to negative cross reactivity, and falsely elevated serum digoxin concentrations with the fluorescence polarization immunoassay for digoxin.

Human hepatic stellate cells, isolated from wedge sections of normal human liver unsuited for transplantation, were separated, after digestion with collagenase/protease, from other liver non-parenchymal cells by ultracentrifugation over gradients of stractan. Cells were cultured on plastic culture dishes in Iscove’s modified Dulbecco’s medium, supplemented as described elsewhere.7 Cells were incubated (1 x 10^4 cells in well dishes) and incubated with increasing concentrations of canrenone or with no (SFIF or control conditions), after a 24 hour incubation period in serum free insulin free (SFIF) medium. After 24 hours, medium was replaced by fresh medium, then resuspended in a total volume of 1 ml and aliquots of 200 μl were processed for aldosterone assay (Radim kit, Italy; Kit 517CT, RIA method). The results were evaluated using linearity (0.007 (0.003); 0.018 (0.004); 0.027 (0.003) nmol/l (mean (SD)); n = 3). As values obtained in SFIF samples were below the lower limit of the assay (0.009 (0.001)), it is conceivable that these cells do not produce aldosterone under basal conditions.8 To further validate this observation, we then spiked sera with increasing concentrations of canrenone (10, 50, 100 μM) and, subsequently, aldosterone concentrations were determined. Sera were collected from patients with different degrees of liver disease (from acute hepatitis (n = 1), to non-alcoholic steatohepatitis (n = 1), or chronic active hepatitis with (n = 3) or without (n = 6) cirrhosis). Aldosterone concentrations were as follows: 0.32 (0.18), 0.63 (0.22), 0.85 (0.26), and 1.07 (0.35) nmol/l (n = 11). Comparing these concentrations with the increasing concentrations of canrenone, no cross reactivity was found (from r = 0.874 to r = 0.988; p<0.001).

To date, only positive interferences leading to falsely high digoxin readings, including those due to spironolactone and canrenone, have been reported.9 Negative interference is much more dangerous. Toxic concentrations may remain undetected. Less severe negative interferences or interferences from clinically less significant cross reactions have been reported. Assay manufacturers should assess potential cross reactivity in the presence of the primary ligand. This difficulty may apply to immunoassays and cross reactions other than digoxin and canrenone. Pathologists and clinicians should be aware of negative interference so that intoxication due to drug dosing guided by monitoring of its concentration in serum can be avoided. For positive interference of a low magnitude, such as in our observation, false positive test results for hormonal and electrolyte disturbances during liver cirrhosis can be avoided.

These in vitro results can be considered evidence for measuring aldosterone concentrations in plasma under in vivo conditions using a more cautious approach, as cross
reactivity could hardly affect biological determinations.

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Responses to endothelin-1 in patients with advanced cirrhosis before and after liver transplantation
In response to Helmy’s comments in his recent letter (Gut 2004;53:470–1), we wish to emphasise the following points, many of which were clearly stated in our original paper.

In agreement with the comments on “generalised vasodilatation” in cirrhosis, we made it clear that basal forearm blood flow was normal in our patient cohort despite the presence of a vasodilated circulation, as evidenced by a reduced systemic vascular resistance index. As pointed out in our paper, this observation is consistent with findings of previous studies and suggests that beds other than the forearm circulation, such as those of the splanchic and pulmonary circulation, were dilated in our patients.

With regard to the issues raised about the use of one arm plethysmography, our own results and those of others have shown that under well controlled circumstances the effects of external stimuli on results obtained using this approach are minimal. Indeed, in our study, the results of forearm plethysmography were very consistent across both the control and cirrhotic patient groups.

However, we acknowledge that single arm plethysmography could be affected by changes in systemic haemodynamics (due for example to the effects of drug infusions). However, as stated in the text, neither heart rate nor blood pressure altered significantly throughout the course of the experiment.

In forearm resistance arteries (and elsewhere), ET<sub>A</sub> receptors on vascular smooth muscle and endothelial cells mediate opposing effects on vascular tone. Thus ET<sub>A</sub>-blockade could result in either vasodilatation or vasoconstriction, depending on which receptor subpopulation is most affected. In our hands, preliminary experiments with the ET<sub>A</sub>-blocker BQ788 yielded ambiguous results, even in control subjects, causing vasocstriction in some and vasodilatation in others. Until a selective ET<sub>A</sub> receptor antagonist (for VSMC or endothelium) is available, interpretation of the results remains difficult.

As for the concern that similar vasodilatation was observed with endothelin-1 (ET-1) and BQ123 (an ET<sub>A</sub> receptor antagonist), we wish to re-emphasise these were two very different experiments in two separate groups of patients, asking two different questions. We observed:

(1) that ET-1 infusion in these advanced cirrhotics produced a broad range of stimulations, and
(2) in similar patients, there was no difference between cirhotoths and controls in the effects of BQ123 on vascular tone.

We put these two results together to propose that it is likely that the abnormal response to ET-1 infusion reflects alterations in ET<sub>B</sub> mediated responses in cirrhotics (either via receptor changes or downstream pathways such as changes in nitric oxide synthase, prostaglandin, or endothelin-derived hyperpolarising factor).

As pointed out by Helmy, there was an early dilatory response following ET-1 infusion in cirrhotics. This was not totally unexpected as it has previously been demonstrated that ET-3 (an ET<sub>B</sub> receptor agonist) causes early vasodilatation in control subjects; similarly, a trend towards an early vasodilatory effect of ET-1 has been observed in healthy subjects.

Regarding the use of concomitant drug therapy, all medications were ceased more than 24 hours prior to the experiments. While some residual effect of these agents is possible, more prolonged cessation of drug therapy in these decompensated patients was not considered safe or ethical. With regard to measurement of ET-1, as detailed in our paper, a commercially available assay with cross reactivity between big ET-1 and ET-1 was used. The study was not powered (nor was it designed to) to pick up small differences in brachial artery ET-1 levels.

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Cluster’s last stand
Guthrie and colleagues (Gut 2003;52:1616–22) described the results of cluster analysis in a patient sample with severe irritable bowel syndrome (IBS). Their analysis investigated a broad range of factors in addition to symp- toms; these included psychosocial measures (psychiatric involvement, health service encounters, quality of life) and physiological parameters (rectal thresholds). The authors have demonstrated that severe IBS can be classified according to nosological dif- ferentiation and, in particular, according to the level of psychological distress, service encounters, and rectal sensitivity. They describe three groups which they labelled “distressed high utilisers”, “distressed low utilisers”, and “tolerant low utilisers.” The authors defend their analysis on clinical grounds and point to treatment implications for each of these groups.

We feel that there are some fundamental points about the nature of cluster analysis that readers of this paper should not overlook. Cluster analysis was initially developed to create and/or evaluate classifications. Its application to gastrointestinal research has followed this approach. In recent years, clustering techniques have been applied to confirm that IBS and functional dyspepsia exist as separate clinical entities, and to evaluate specific syndrome subgroups, as described by the current Rome criteria. Following traditional clinical approaches, cluster solutions have generally been derived from symptom based parameters, including frequency, severity, and predominant complaints.

The term “cluster analysis” describes a range of procedures which use empirical methods to form groups of highly similar entities. While the notion that cluster analy- sis is solution seeking, rather than solution imposing, these techniques is essentially solution imposing: that is, clustering methods will always place objects into groups. Furthermore, as there are no formal statistical procedures to evaluate the resulting solution, the reasonableness of any solution is determined only on the basis of personal judgement. This is a problem. Indeed, critics of the approach have argued that cluster analysis encourages naïve empiricism—that is, inclusion of as many variables as possible in the hope that a meaningful structure will come out. However, proponents of cluster analysis have suggested that careful selection of variables on theoreti- cal grounds can overcome these techniques.

It is intuitively obvious that any single entity can be classified according to a broad range of dimensions, and Guthrie et al have certainly demonstrated this with respect to IBS. However, we rarely classify any entity according to all possible dimensions simultaneously; this would lead to a complex set of descriptors which may be unwieldy and contain many redundancies. Rather, we tend to select a subset of meaningful dimensions that best suit our purposes in forming a classification.

There are certainly theoretical grounds for considering psychological involvement when evaluating patients with IBS. However, we rarely classify any entity according to all possible dimensions simultaneously; this would lead to a complex set of descriptors which may be unwieldy and contain many redundancies. Rather, we tend to select a subset of meaningful dimensions that best suit our purposes in forming a classification.

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may stigmatise some patients with this complaint; one of the unfortunate consequences of classification is the tendency to attach labels to the subgroups that emerge.

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Small bowel carcinoma and coeliac disease
We thank Howdle et al for their comments on our study, detailed recently in their letter (Gut 2003;53:470). In their British Society of Gastroenterology (BSG) National Survey,1 Howdle et al relied mainly on gastroenterologists and surgeons to report cases of small bowel carcinoma and whether they were associated with either coeliac or Crohn’s disease. This may have resulted in underestimation of associated coeliac disease. In our series, we had two cases in which the original pathologist had failed to recognise the histological features of coeliac disease in mucosa adjacent to the adenocarcinoma.2 The diagnosis of coeliac disease was made after review of the original resections. This problem has been recognised previously3 and results in the underdiagnosis of coeliac disease and further diagnostic delay for the patient with coeliac disease.

While the individual risk for patients with coeliac disease in developing adenocarcinoma of the small intestine is not great, poor survival should prompt rapid evaluation when symptoms occur. In addition, there should be a consideration of whether there is a subset of patients with coeliac disease who would benefit from screening for these cancers. Because patients with coeliac disease do not have a significantly increased risk of duodenal adenomas,4 the role of video capsule endoscopy of the entire small intestine needs to be explored.

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14th International Workshop of Digestive Endoscopy, Ultrasonography and Radiology
The 14th International Workshop of Digestive Endoscopy, Ultrasonography and Radiology will be held in Marseille on 27–28 May 2004. For further information, please contact: Nathalie Fontant, Atelier Phenic, 41 rue Docteur Morucci, 13006 — Marseille (tel: (33) 04-91-57-15-28; e-mail: nf@aphenix.com).

European Postgraduate Gastro-surgical School (EPGS) Courses 2004
The EPGS at the Academic Medical Center of the University of Amsterdam will be holding the following courses during the year: ‘Benign Hepato-Biliary Disorders’ will be held on 22 & 23 April 2004, ‘Endosonography live in Amsterdam’ will be held on 2, 3 & 4 June 2004, and ‘Update in Coloproctology’ will be held on 28 & 29 October 2004. For further information, please contact: J Goedkoop (tel: (31) 566 3926; fax: (33) 267 5594; e-mail: j.goedkoop@amc.uva.nl; website: www.epgs.nl).

8th Southeast European Symposium of Paediatric Surgery
The 8th Southeast European Symposium of Paediatric Surgery will focus upon ‘Infectious Problems in Paediatric Surgery’. The event will be held between 24–25 September 2004, at the University of Graz, Austria. For further information, please contact: Professor M E Höllwarth, Department of Paediatric Surgery, Medical University of Graz, Austria, Auenbruggerplatz 34, 8036 Graz; tel: +43 34 316 385 3762; fax: +31 316 385 3775; e-mail: kinderchirurgie@uni-graz.at.