Synbiotic therapy (Bifidobacterium longum/Synergy 1) initiates resolution of inflammation in patients with active ulcerative colitis: a randomised controlled pilot trial

E Furrie, S Macfarlane, A Kennedy, J H Cummings, S V Walsh, D A O’Neil, G T Macfarlane

Background and aims: Ulcerative colitis (UC) is an acute and chronic inflammatory disease of the large bowel with unknown aetiology. The immune response against normal commensal microorganisms is believed to drive inflammatory processes associated with UC. Therefore, modulation of bacterial communities on the gut mucosa, through the use of probiotics and prebiotics, may be used to modify the disease state.

Methods: A synbiotic was developed for use in UC patients combining a probiotic, Bifidobacterium longum, isolated from healthy rectal epithelium, and a prebiotic (Synergy 1), a preferential inulin-oligosaccharide growth substrate for the probiotic strain. Treatment was employed in a double blinded randomised controlled trial using 18 patients with active UC for a period of one month. Clinical status was scored and rectal biopsies were collected before and after treatment, and transcription levels of epithelium related immune markers were measured.

Results: Sigmoidoscopy scores (scale 0–6) were reduced in the test group (start 4.5 (1.4), end 3.1 (2.5)) compared with placebo (start 2.6 (2.1), end 3.2 (2.2)) (p = 0.06). mRNA levels for human beta defensins 2, 3, and 4, which are strongly upregulated in active UC, were significantly reduced in the test group after treatment (p = 0.016, 0.038, and 0.008, respectively). Tumour necrosis factor α and interleukin 1α, which are inflammatory cytokines that drive inflammation and induce defensin expression, were also significantly reduced after treatment (p = 0.018 and 0.023, respectively). Biopsies in the test group had reduced inflammation and regeneration of epithelial tissue.

Conclusions: Short term synbiotic treatment of active UC resulted in improvement of the full clinical appearance of chronic inflammation in patients receiving this therapy.

Ulcerative colitis (UC) is a relapsing inflammatory disease of the colon with unknown aetiology. Evidence from animal models suggests that an altered immune response towards the commensal gut microbiota plays a key role in the development and maintenance of this condition.11 For example, knockout or transgenic mice with genetic susceptibilities to IBD only acquire characteristic lesions when their colon is populated with normal commensal bacteria while germ free animals do not manifest an inflammatory response.12–14 Evidence from human studies has also suggested that mucosal bacterial populations in UC may be altered towards a more proinflammatory phenotype.2–4 Manipulation of the mucosal microbiota to reduce the inflammatory potential of colonising bacteria is therefore an attractive therapy for UC. One option is to use antibiotics to remove species involved in inducing the inflammatory response. However, antibiotic therapy has had limited success in UC, possibly due to the fact that treatment needs to be customised for individual patients.10–11 An alternative is to use probiotic bacteria that interact with the host epithelium to resolve inflammation. Probiotics have been defined as live microbial feed supplements that beneficially affect the host by improving its intestinal microbial balance.15 The most widely used probiotics in humans are bifidobacteria and lactobacilli but other organisms such as Escherichia coli and the yeast Saccharomyces boulardii have been reported to have some beneficial effects in maintaining remission in inflammatory bowel disease (IBD).16–20

Probiotic therapy can be potentially improved through combination with a prebiotic (usually a non-digestible oligosaccharide that is not absorbed in the upper gut) that promotes growth of the probiotic in the large bowel. This combination is known as a synbiotic. A further advantage of using a synbiotic is that the probiotic component would promote the growth of indigenous organisms in the gut with probiotic properties.21 Few clinical trials have been performed with probiotics to treat or maintain remission in IBD and the resulting success has been variable.22–25 The best known product, VSL#3, uses a mixture of eight different bacterial strains that have been reported to prevent the onset of pouchitis after pouch formation in UC patients.26–27 Assessments of inflammation in UC have traditionally been done by the gastroenterologist on examination of the bowel at endoscopy and by a pathologist who examines microscopic inflammatory responses in biopsies. However, new sensitive methods of measuring changes in gene expression in inflamed tissue have been developed. Proinflammatory cytokines have been measured before in probiotic therapy in ex vivo colonic biopsy specimens from Crohn’s patients where decreased expression of tumour necrosis factor α (TNF-α) occurred,28 but these cytokines can be produced by a variety of infiltrating cells and give little indication of the immune responsiveness of the epithelium.29–30 A newly described group of antimicrobial peptides known as human beta defensins (hBD) are expressed uniquely by epithelial cells. Six hBD (hBD1–6) are currently recognised. hBD1 is constitutively expressed and its promoter has no functioning...
response elements; therefore, the level of expression of hBD1 mRNA directly allows absolute determination of the number of epithelial cells in each biopsy. 11-13 hBD2 and 3 have been shown to be significantly upregulated in UC but not in Crohn’s disease 14-16 and are consequently excellent target genes for assessing inflammatory responses in UC epithelia.

In this pilot study, we have developed a synbiotic for treating UC patients and, to our knowledge, this is the first report of synbiotic therapy in the treatment of UC. To investigate the effectiveness of the synbiotic, we used traditional methods of assessing inflammation, in conjunction with new techniques that directly measure the inflammatory state of the gut epithelium in a quantitative way.

The principal aim of this work was to induce clinical improvement in UC patients receiving synbiotic therapy in comparison with the placebo group. The second objective was to assess the effects of synbiotic feeding on the mucosa, with reference to inflammatory mediators induced in active colitis.

METHODS

Patients
Consecutive patients with active UC attending the Gastroenterology Outpatients Clinic, Ninewells Hospital, were asked to give written consent to take part in this investigation. Eighteen patients accepted the invitation. Eligible patients were aged 24-67 years who had not received antibiotics in the last three months and were not taking commercially available probiotic preparations. Normal healthy control biopsies were obtained from other patients attending the clinic who had been shown by sigmoidoscopy and histology to have no evidence of inflammatory bowel disease. These studies were approved by the Tayside Committee on Medical Research Ethics, Dundee (study number 205/01).

Study design
Eighteen study numbers were assigned and randomised using a table of random digits. 37 Nine patient numbers were assigned to the test group and nine to the placebo group. The 18 patients were randomly assigned to either group, and given a study number (SO1 to 18). This assignment was not divulged to the clinician, patient, or inhouse researcher who carried out the experimental measurements. A description of patient involvement is given in table 1. Two patients did not attend the first study visit because they had taken antibiotics after recruitment, and therefore 16 patients entered the study. Their characteristics are given in table 2. Patients were recruited on the basis of active inflammation; time lag between recruitment and initiation of the study was up to two months. In one case (table 2) a placebo patient had attended the first study visit because they had taken antibiotics in the last three months and were not taking antibiotics in the last three months and were not taking

synbiotic therapy

improvement in UC patients receiving synbiotic therapy in

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### Table 1

<table>
<thead>
<tr>
<th>Symbiotic (8 starting patients)</th>
<th>Placebo (8 starting patients)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pretreatment</td>
<td></td>
</tr>
<tr>
<td>During study</td>
<td>One removed (antibiotics)</td>
</tr>
<tr>
<td></td>
<td>No adverse responses</td>
</tr>
<tr>
<td>Post-treatment</td>
<td>Remaining 8 patients completed the study, 7 with full biopsy retrieval</td>
</tr>
<tr>
<td></td>
<td>Remaining 6 patients completed study, 5 with full biopsy retrieval</td>
</tr>
</tbody>
</table>

### Table 2

<table>
<thead>
<tr>
<th>Clinical details of patients on commencement of therapy</th>
</tr>
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<tbody>
<tr>
<td>Symbiotic (8 starting patients)</td>
</tr>
<tr>
<td>Age (y)</td>
</tr>
<tr>
<td>Sex (M:F)</td>
</tr>
<tr>
<td>Duration of disease (y)</td>
</tr>
<tr>
<td>Disease extent*</td>
</tr>
<tr>
<td>Steroids</td>
</tr>
<tr>
<td>Immunosuppressants</td>
</tr>
<tr>
<td>5-ASA</td>
</tr>
<tr>
<td>Initial clinical activity index</td>
</tr>
<tr>
<td>Initial sigmoidoscopy score</td>
</tr>
<tr>
<td>Initial CRP (mg/l)</td>
</tr>
</tbody>
</table>

Values are mean (range) or number.
*Disease extent unknown for one placebo patient.
†Ulcerative colitis related drugs only.

CRP, C reactive protein.

 Diary, previously validated by Cummings and colleagues, 40 in which details of the amount and consistency of each stool were recorded, together with the presence of blood or mucus, and whether the subject experienced abdominal pain. Rectal biopsies were also taken for histology scoring by the Department of Pathology, Ninewells Hospital (no inflammation = 0, mild inflammation = 1, moderate inflammation = 2, and severe inflammation = 3). Venous bloods were taken for measurement of C reactive protein (CRP; Biochemical Medicine, Ninewells Hospital, UK). Further biopsies were used for inhouse assessment of mucosal inflammatory mediators.

Production of the probiotic

Nineteen different bifidobacterial isolates were assessed for suitability as a probiotic strain (10 were isolated from healthy colonic mucosa, five were obtained from the faeces of healthy donors, and four were culture collection strains). They comprised six strains of B adolescentis, two B bifidum, two B infantis (DSM 86184, ATCC 15617), two B longum, and one

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conditions at 37 ˚C in Wilkins-Chalgren (WC) broth. Bacteria
subsequently grown overnight in batch culture under anaerobic
tests were checked by two independent microbiologists
maximise contaminant detection. All quantitation and purity
plates. These were incubated aerobically and anaerobically to
weigh, and in order to determine cell viability and check
pellet was frozen overnight at
80˚C and then lyophilised
Mucosa. Its identity was confirmed by partial sequenc-
ing (500 of 1500 bp) of the variable region of the 16S rRNA
gene. Further analysis of the ability of the selected organism
to alter the profile of cytokine expression in a model
epithelial cell line (HT29) was investigated. Proinflammatory
cytokine interleukin 1α (IL-1α) was measured by both real
time polymerase chain reaction (PCR) for mRNA levels and
cytokine ELISA for secreted proteins, either alone or in
coculture with the selected B longum, B bifidum, and a
commensal Escherichia coli. The selected probiotic was found
to significantly reduce IL-1α produced by HT-29 cells with
respect to mRNA levels (p < 0.05) and protein (p < 0.05),
unlike the other two organisms which increased expression
of this proinflammatory cytokine. The organism was subse-
sequently grown overnight in batch culture under anaerobic
conditions at 37˚C in Wilkins-Chalgren (WC) broth. Bacteria
were harvested by centrifugation (14 000 g, 30 minutes) and
washed twice in anaerobic phosphate buffered saline. The
pellet was frozen overnight at −80˚C and then lyophilised
(Modulyo Bench Top Freeze Dryer; Thermo Savant,
Holbrook, New York, USA). The resulting powder was
weighed, and in order to determine cell viability and check
purity, serial dilutions were made and plated onto WC agar
plates. These were incubated aerobically and anaerobically to
maximise contaminant detection. All quantitation and purity
tests were checked by two independent microbiologists
during production and packaging of the probiotic.

Table 3 System used to score inflammation in the colon at sigmoidoscopy (modified from Baron and colleagues9)

<table>
<thead>
<tr>
<th></th>
<th>0</th>
<th>1</th>
<th>2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Friability</td>
<td>None</td>
<td>On touching mucosa</td>
<td>–</td>
</tr>
<tr>
<td>Bleeding</td>
<td>No blood</td>
<td>Blood in lumen</td>
<td>–</td>
</tr>
<tr>
<td>Vessel pattern</td>
<td>Normal</td>
<td>Some patchy changes</td>
<td>All vessels obscured</td>
</tr>
<tr>
<td>Overall appearance</td>
<td>Normal</td>
<td>Moderate change</td>
<td>Severe active disease</td>
</tr>
</tbody>
</table>

Each of B angulatum, B breve, B catenulatum, B dentium, B lactis
(DSM 10140), B pseudocatenulatum, and B suis (ATCC 17533).
The organisms were assessed for aerotolerance, acid toler-
ance, bile salt resistance, adhesion to epithelial cells, and
their abilities to utilise oligofructose as an energy source.

Preparation of standards for quantitation of DNA
Standard amounts of DNA corresponding to the target
sequences are needed to carry out real time PCR. This was
achieved by making purified plasmid DNA containing the
target sequences. Briefly, cDNA from normal healthy colon,
or mononuclear blood cells, was amplified using the specific
PCR primer pair (see table 4). Product of correct size and
sequence was purified using the Qiagen PCR purification
kit (Qiagen) and ligated into a vector using the pGEM-T easy
vector system 1 (Promega). JM109 competent E coli
(Promega) were transformed with each ligated vector, and
after overnight incubation positive colonies were chosen.
From each selected colony the plasmid was purified using the
Wizard plus SV miniprep system (Promega). Concentration of the plasmid preparation was determined by agarose gel
electrophoresis with known standards (New England
Biolabs, Beverly, Massachusetts, USA). Samples were diluted to 10^10 molecules/μl, aliquoted, and stored (−80˚C).

Table 4 Real time polymerase chain reaction primers used to quantitate mRNA/rRNA
levels in rectal biopsies

<table>
<thead>
<tr>
<th>Gene target</th>
<th>Primer pair 5’ to 3’</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glyceraldehyde-3-phosphate dehydrogenase (GAPDH)</td>
<td>GGAAGGTGAAAGGTCCGAGTC</td>
</tr>
<tr>
<td>Human beta defensin 1 (hBD1)</td>
<td>CTCAGCTTCGAGGTCATCGT</td>
</tr>
<tr>
<td>Human beta defensin 2 (hBD2)</td>
<td>CTCAGCTTCGAGGTCATCGT</td>
</tr>
<tr>
<td>Human beta defensin 3 (hBD3)</td>
<td>CTCAGCTTCGAGGTCATCGT</td>
</tr>
<tr>
<td>Human beta defensin 4 (hBD4)</td>
<td>CTCAGCTTCGAGGTCATCGT</td>
</tr>
<tr>
<td>Interleukin 1α (IL-1α)</td>
<td>TCTCGAACCCCGAGTGACAA</td>
</tr>
<tr>
<td>Interleukin 10 (IL-10)</td>
<td>CTTGCTTCCATCTGGGAGCT</td>
</tr>
<tr>
<td>Tumour necrosis factor 1α (TNF-α)</td>
<td>TCTCGAACCCCGAGTGACAA</td>
</tr>
<tr>
<td>Bifidobacterial genus specific</td>
<td>AGGGTTCGATTCTGGCTCAG</td>
</tr>
</tbody>
</table>

RNA and cDNA preparation
Biopsies were macerated using liquid nitrogen snap freezing
and mechanical grinding. RNA was purified using the RNA
easy kit (Qiagen, Hilden, Germany) with an initial clean-up
stage using a Qiashredder column (Qiagen) and an additional
step of DNA digestion to ensure no genomic DNA contamina-
tion. Samples were reverse transcribed using the AMV RT
kit (Promega, Madison, Wisconsin, USA) according to the
manufacturers instructions and aliquoted before storage at
−80˚C.
Synbiotic therapy improves active ulcerative colitis

**RESULTS**

**Inflammatory markers in healthy and UC tissues**

All results shown for inducible hBD (2–4) were normalised for epithelial cell numbers, as determined by levels of hBD1, the constitutive epithelial cell housekeeping gene, and cytokine levels were normalised for total cells per biopsy using glyceraldehyde-3-phosphate dehydrogenase. Significance between the pre- and post-synbiotic groups were hBD2 (*p = 0.0156), hBD3 (*p = 0.0079), hBD4 (*p = 0.0078), TNF-α (*p = 0.0175), IL-1α (*p = 0.0379), and IL-10 (NS). Significance between the post-synbiotic and post-placebo groups were hBD2 (NS), hBD3 (NS), hBD4 (NS), TNF-α (tp = 0.0177), IL-1α (tp = 0.0051), and IL-10 (NS).

**Effect of synbiotic on inflammatory markers in the gut**

Figure 2 shows the results from the two trial groups, before and after the four week feeding period. Significant differences in expression of inducible hBD occurred in the synbiotic patients (hBD2 p = 0.0156, hBD3 p = 0.0379, hBD4 p = 0.0078). No significant differences between the post-synbiotic group and the post-placebo were observed for hBD

**Statistics**

Significant differences between the pre-synbiotic and post-synbiotic therapy groups and between the post-synbiotic and post-placebo groups were assessed for hBD and cytokine results using the Mann-Whitney test for non-parametric analysis as the data were not normally distributed. Significant difference in clinical outcome was assessed using the difference in sigmoidoscopy score (SS) in the synbiotic group over four weeks compared with the difference in SS in the placebo group using a two sample t test with equal variances. Significance was given for p values less than 0.05.

**Histopathology**

Tissue specimens were fixed in formalin and embedded in paraffin. Sections (4 μm thick) were cut using a microtome (Leica RM 2135) and mounted onto clean glass slides which had been coated with poly-L-lysine (Superfrost plus; VWR International Ltd, Poole, Dorset, UK) to improve tissue adherence. Four serial sections were stained with haematoyxlin and eosin, using standard methods, to visualise tissue morphologies.

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was carried out using an iCycler and the iQ SYBR Green Supermix (BioRad, Hercules, California, USA). Test samples were added in triplicate at 2 μl/well in a 20 μl total reaction volume.
due to the small number of placebos who completed the trial. Similar results were obtained for the proinflammatory cytokines TNF-α and IL-1α, with significant reductions in expression in the post-feeding synbiotic patients compared with the pre-synbiotic group (p = 0.0175 and p = 0.0379, respectively). On comparison of the levels of inflammatory cytokines for the post-synbiotic group versus the post-placebo group, we found a significant reduction for both TNF-α and IL-1α levels (p = 0.0177 and p = 0.0051, respectively). The most marked reduction was seen with IL-1α, which returned to levels found in normal healthy tissue (see fig 1). No significant difference was seen in IL-10 between the placebo and synbiotic groups.

**Presence of bifidobacteria on the mucosa**

Levels of bifidobacterial specific total rRNA were determined using real time PCR in mucosal biopsies pre- and post-treatment. The synbiotic group had a starting mean of 1406 molecules of bifidobacterial rRNA per biopsy increasing to 16285 over the same four week interval, a 42-fold increase. The placebo group had a mean starting level of 3527 molecules of bifidobacterial rRNA per biopsy increasing to 58699 at the end of the study, a 422-fold increase. Samples were normalised for number of epithelial cells by hBD1 levels.

**Clinical outcome**

Clinical outcome for patients was assessed by sigmoidoscopy score (SS), clinical activity index (CAI), and bowel habit index (BHI). Results are expressed as the difference in values recorded on day 1 and day 28 of the trial for SS and CAI, and as the percentage change over the four week period for BHI. SS (maximum change possible = 6) was decreased by a mean of 1.3 points over the four week period in the synbiotic group (n = 8); five patients improved, two remained the same, and one increased from 4 to 5. The placebo group had an increase of 0.58 over the same period (n = 6); three had increased scores and three had decreased scores. SS was reduced markedly in the synbiotic group after treatment (start 4.5 (1.4), end 3.1 (2.5)) compared with the placebo group (n = 6; start 2.6 (2.1), end 3.2 (2.2)). The difference in pre- and post-feeding between the synbiotic and placebo groups was borderline significant (p = 0.06). CAI (maximum score 19) was reduced in five patients in the synbiotic group with three showing an increase in score (start 5.6 (3.7), end 5.3 (3.4)). The placebo group had three patients with improving scores, one which remained the same and two with increased CAI (start 4.9 (3.2), finish 3.1 (2.5)). Mean BHI increased in the placebo group by 70.4% compared with a reduction of 20.4% in the synbiotic group.

**Correlation of hBD levels with sigmoidoscopy scores (SS)**

Analysis of the relationship between SS and levels of individual hBD showed that there was no correlation between hBD1 with SS, before or after treatment, in either the test or placebo group (results not shown). However, in both synbiotic and placebo groups pretreatment, there was a positive correlation between SS and mRNA for hBD2–4 whereby in more active disease higher levels of inducible hBD were detected (fig 3A). At the end of the feeding study, comparison of SS and hBD showed that this positive correlation remained only in the placebo group whereas hBD2 and hBD4 manifested a loss of correlation with SS score, and hBD3 had an inverse relationship in the synbiotic group.

**Correlation of hBD levels with histology scores (HS)**

A positive correlation was evident in the relationship between HS and hBD2–4 in both synbiotic and placebo groups, before
the feeding study (fig 3B). After treatment, the placebos retained a positive correlation for all three inducible beta defensins, as occurred with SS. In the symbiotic group, a positive correlation for hBD2 and 4 remained when comparing HS with mRNA concentrations but hBD3 levels related poorly with HS in this group (fig 3B).

**C reactive protein (CRP) levels in blood**

Only five of the eight symbiotic patients had elevated CRP (mean 6.0 (SD 6.5)), and only one of six patients in the placebo group who completed the study had elevated CRP (mean 1.6 (SD 3.6)). After treatment, the symbiotic group showed a reduction in CRP (mean 1.8 (SD 3.9)) after four weeks. None of the placebo patients had raised circulating levels of CRP.

**Histopathology of rectal biopsies**

Histology scoring produced results similar to SS; four individuals showed a reduction in score and two remained the same, with a mean reduction in HS in the symbiotic group over the four week period (start 1.7 (1.4), end 1.1 (1.2)) compared with the mean increase in the placebo group (start 0.9 (0.9), end 1.9 (1.1)), with three showing an increase in score and two remaining the same. Two representative paired biopsies are shown in fig 4: A (SS = 6, HS = 3) and C (SS = 3.5, HS = 1.5) were from a symbiotic patient, pre- and post-feeding, respectively, while B (SS = 1.5, HS = 1.5) and D (SS = 5.0, HS = 2.5) were from a placebo treated patient, at the beginning and end of the study. Comparison of fig 4A and C shows resolution of acute inflammatory activity following symbiotic consumption. The crypt abscesses in fig 4A have disappeared and the epithelium shows a more regenerative appearance in fig 4C. Small crypt abscesses are evident in the rectal biopsy from the placebo patient (fig 4B). This inflammation has been exacerbated in fig 4D, with a larger abscess visible in the middle of the section, with crypt rupture, and a general increase in the numbers of infiltrating cells.

**DISCUSSION**

Consumption of symbiotic twice daily over four weeks significantly reduced mucosal inflammatory markers in active UC. This was concurrent with a reduction in colitis at the macroscopic and microscopic level. However, although lowered SS were observed in the symbiotic group, they were not as marked as the reductions in HS and inflammatory markers. This may indicate that changes in inflammatory mediators at the molecular level could precede gross clinical changes scored through sigmoidoscopy by several weeks. This contention is supported by the relationship seen in HS and not SS after symbiotic feeding, with both hBD2 and 4 showing that although local inflammation was resolving in these patients, more general changes in colonic appearance, such as vessel pattern, take longer to return to normal. Alternatively, this therapy may be resolving the local inflammation through direct contact with the epithelium but is insufficient in all cases of UC (three of eight patients receiving symbiotic therapy had no improvement in SS) in reducing the systemic chronic inflammatory condition. It is possible that these patients may improve their clinical scores if this treatment is given in conjunction with other therapies or through selection of a different probiotic organism. Unfortunately, no significant difference in SS or CAI between the symbiotic and placebo groups was found. This may be due to the small number of placebos who completed the study. Two placebos requested withdrawal because of exacerbation of symptoms, and refused to undergo sigmoidoscopy or provide a final biopsy due to discomfort. There were no withdrawals from the symbiotic group after commencement of the study and no reports of adverse reactions.

Markers of active disease in biopsies from UC patients include proinflammatory cytokines such as TNF-α, IL-8, IL-1α, and IL-1β. These molecules are upregulated in active UC but they are not specific for the epithelium as the large numbers of infiltrating leucocytes in the mucosa contribute greatly to their formation. When using a probiotic or symbiotic, it is important to be able to assess directly its effects on the epithelial barrier as well as the underlying
immune system as the epithelium is the first point of host contact for the organisms. Human beta defensins are good indicators of mucosal immunity because they are uniquely expressed by epithelial cells. Two types of expression occur: hBD1 is a non-inducible constitutively expressed molecule while hBD2-4 are upregulated by bacterial challenge and proinflammatory cytokines. These molecules are produced by the inflammatory infiltrate and by the epithelium itself directly affecting expression levels of inducible hBD in epithelial cells. IL-1 has been shown to induce upregulation of hBD2 in gastric and colonic epithelial cell lines.

In this study, a direct relation between mRNA synthesis for inducible hBD and severity of UC (SS) has been shown for the first time, together with the relationship between inducible hBD and histology scores in UC patients. We were unable to demonstrate a significant difference between the inducible hBD in the post-synbiotic group versus the post-placebo group but the proinflammatory cytokines TNF-α and IL-1α were significantly reduced in the synbiotic group compared with the placebo group. This can be explained by the relationship between SS and levels of inducible hBD. As our placebo group had lower clinical scores than the synbiotic group, resulting from the small sample size and random assignment to each group, we have inadvertently biased against finding a significant difference in inducible hBD between the two groups due to the direct linear relationship between SS and hBD. This is not true for the inflammatory cytokines that have a more all or nothing expression pattern and can be produced by any infiltrating cell in the biopsy. Furthermore, they are compared against the usual housekeeping genes (GAPDH/β-actin) for biopsy size standardisation, unlike hBD which are only produced by colonic epithelial cells and epithelial cell numbers can be accurately standardised by hBD1 levels.

The functions of these induced hBDs are unknown in UC disease progression. However, hBDs are antimicrobial peptides that are particularly good at destroying Gram negative bacteria so their production may be an attempt by the epithelium to modify the composition of the mucosal biofilm. The bifidobacterial probiotic strain used in this study was found to be insensitive to hBD mediated killing by recombinant hBD1, 2, and 3 (results not shown), and therefore, as part of a therapy for UC this organism would exhibit better survival characteristics on the inflamed mucosa and interact directly with the epithelial surface and modify the immune response. We detected higher numbers of total bifidobacteria on the mucosal surface in patients fed the synbiotic compared with those taking placebo. We cannot separately identify the probiotic organism from the indigenous bifidobacteria that may have been stimulated by inclusion of a prebiotic in the feeding regimen. Bifidobacterium longum is a very complicated organism with at least five different subgroups contained within the species when 16S rDNA sequences are compared (data not shown). With the present technology it is impossible to distinguish the probiotic strain from other Bifidobacterium longum in a small pinch biopsy. It is unknown what direct effects probiotic bacteria have on the inflamed epithelium. They could simply be out competing microorganisms involved in disease aetiology for microcolonies on the gut wall, thereby removing inflammatory stimuli. However, they may also be directly influencing the expression of inflammatory cytokines by the epithelium through ligation of particular pattern recognition receptors on the luminal cell surface, as supported by studies using HT29 cells in preliminary screening experiments with the probiotic. Alternatively, the probiotic could be directly influencing dendritic cells that are present in high numbers in the lamina propria with access to the epithelial surface, by downregulating proinflammatory cytokine pathways and stimulating a more immunomodulatory and tolerant immune response.

In conclusion, this short term pilot study has provided the first evidence that synbiotics have the potential to be developed into acceptable therapies for patients suffering from active UC. A large scale clinical trial is now needed to investigate the long term effects of synbiotic use in inducing and maintaining remission in patients with active disease.

ACKNOWLEDGEMENTS
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AUTHORS’ AFFILIATIONS
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Conflict of interest: None declared.

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4 Butcher EC. Leukocyte-endothelial cell recognition: three (or more) steps to specificity and diversity. Cell 1991;67:1033–4.
EDITOR’S QUIZ: GI SNAPSHOT

Answer

From question on page 206

Computed tomography scan of abdomen revealed a large collection of intrahpereal gas peripherally (fig 1) and dilated loops of intestine with pneumatosis (fig 2). Hypotension persisted despite aggressive volume resuscitation. Exploratory laparotomy revealed necrotic intestine from the proximal jejunum to the hepatic flexure. The necrotic intestine was resected, and jejunotransversostomy was performed.

Intrahpereal gas may represent either an incidental finding or a life threatening condition. Intrahpereal gas may be present in either the biliary or portal venous systems. Computed tomography scan may help differentiate gas in the biliary system from that of the portal venous vasculature. Biliary gas tends to be located centrally in the hiliar region in the larger bile ducts. In contrast, gas in the portal system extends to the smaller portal venous branches, resulting in a less localised peripheral distribution. Mucosal and submucosal bowel wall damage may allow some intraluminal air to enter the damaged or dissected bowel wall, leading to pneumatosis, and also to enter some intramural veins, leading to portal venous gas. Therefore, portal venous gas may be observed in any patient with severe inflammation of the intestinal walls—for example, necrotising enterocolitis, ulcerative colitis, Crohn’s disease, large ulcerating gastric carcinoma, appendicitis, and diverticulitis.

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Figure 2. Computed tomography scan demonstrating gas collections in the intestinal wall (black arrows), and a thickened oedematous intestinal wall (arrowhead). White arrow represents luminal air.