Translocation of p21\textsuperscript{Cip1/WAF1} from the nucleus to the cytoplasm correlates with pancreatic myofibroblast to fibroblast cell conversion

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Background and aims: In the pancreas, myofibroblasts (MFBs) were shown to play an important role in the cellular response during inflammation and injury. However, there is only fragmentary information concerning the fate of these cells in pancreas regeneration and fibrosis development.

Methods: Explant cultures of rat pancreatic tissue were used as a model to follow cellular dynamics and phenotype conversion of pancreatic MFBs in vitro. For detailed biochemical analyses a pancreatic fibroblast cell line (long culture fibroblast (LCF)) was generated from MFBs in a long term culture. Cerulein induced acute pancreatitis and dibutyltin dichloride induced pancreas fibrosis were used as experimental models for acute and chronic fibrogenic reactions, respectively.

Results: In the explant culture, pancreatic MFBs which derived from fat storing fibroblastic cells underwent apoptosis or converted again to fibroblasts. The phenotype switch to fibroblasts was associated with translocation of p21\textsuperscript{Cip1/WAF1} from the nucleus into the cytoplasm. Molecular analyses in LCFs revealed subsequent binding to and inhibition of the activities of Rho kinase 2 and apoptosis signal regulating kinase 1. In the experimentally established pancreas fibrosis, fibroblasts with cytoplasmic expression of p21\textsuperscript{Cip1/WAF1} were distributed throughout fibrotic bands whereas in experimental acute pancreatitis MFBs with nuclear expression of p21\textsuperscript{Cip1/WAF1} dominated.

Conclusions: The results indicate that pancreatic MFBs are transient and suggest that intracellular localisation of p21\textsuperscript{Cip1/WAF1} can contribute to the phenotype conversion of these cells to fibroblasts in culture and experimental injury.

METHODS

Antibodies and reagents

Mouse monoclonal anti-\(\alpha\)-SMA (clone 1A4), anti-vimentin (clone V9), anti-desmin (clone DEU 10), anti-glial fibrillary acidic protein (GFAP, clone GA5), and anti-myosin light chain (MLC, clone MY21), and rabbit polyclonal anti-antibodies with anti-Rock2 (H85), and anti-ASK1 (H300), and mouse monoclonal anti-\(\alpha\)-SMA (clone 1A4), anti-vimentin (clone V9), anti-desmin (clone DEU 10), anti-glial fibrillary acidic protein (GFAP, clone GA5), and anti-myosin light chain (MLC, clone MY21), and rabbit polyclonal anti-antibodies with anti-Rock2 (H85), and anti-ASK1 (H300), and mouse monoclonal anti-pp38 (D8), anti-p21 Cip1/WAF1 (F5), and anti-myosin light chain (MLC, clone MY21), and rabbit polyclonal anti-antibodies with anti-Rock2 (H85), and anti-ASK1 (H300), and mouse monoclonal anti-pp38 (D8), anti-p21 Cip1/WAF1 (F5), and anti-death receptor 5 (DR5) (M20) antibody were from Santa Cruz Biotechnology (Santa Cruz, California, USA). Mouse monoclonal anti-\(\alpha\)-SMA (clone 1A4), anti-vimentin (clone V9), anti-desmin (clone DEU 10), anti-glial fibrillary acidic protein (GFAP, clone GA5), and anti-myosin light chain (MLC, clone MY21), and rabbit polyclonal anti-antibodies with anti-Rock2 (H85), and anti-ASK1 (H300), and mouse monoclonal anti-pp38 (D8), anti-p21 Cip1/WAF1 (F5), and anti-death receptor 5 (DR5) (M20) antibody were from Santa Cruz Biotechnology (Santa Cruz, California, USA).
monoclonal anti-CD 95L (F 37720) was from Calbiochem (San Diego, California, USA). Rabbit polyclonal anti-collagen I (Col 1) (11346) was from Rockland (Gilbertsville, Pennsylvania, USA). Rabbit polyclonal anti-pMLC 2 (Nr 3674) and mouse monoclonal anti-pSAPK/JNK (No 9255) were from Cell Signaling (Beverly, Massachusetts, USA). For immunofluorescence, Alexa Fluor 488 conjugated goat anti-mouse IgG and Alexa 594 conjugated chicken anti-rabbit Ig (Molecular Probes, Eugene, Oregon, USA) were used as secondary antibodies. Monoclonal anti-rabbit Ig and monoclonal anti-mouse Ig, conjugated with AP (Dako A/S, Glostrup, Denmark) were used as secondary antibodies for immunoblotting. BrdU labelling and detection kit 1 and the in situ cell death detection kit were from Roche (Mannheim, Germany). All trans-retinol (cat No R 7632), myelin basic protein (M 2016), and trypsin blue (T 8154) were from Sigma. JNK inhibitor I (form, cell permeable, No 420116) was from Calbiochem.

**Animals**

Male Wistar rats (90 days old, weighing 260–270 g, water ad libitum) were obtained from an outbreeding colony of the University of Rostock.

**Explant culture**

Rats were anaesthetised using 5-ethyl-5-isooamylbarbituric acid (sodium salts), the pancreas was removed, and after mechanical cutting of the tissue into small blocks the pieces were placed into Col 1 (Sigma) coated Petri dishes (size 94 mm) and cultured in Dulbecco’s modified Eagle’s medium (Invitrogen, Karlsruhe, Germany) supplemented with 10% fetal calf serum at 37 ˚C in a 5% CO₂ atmosphere.

**Long term subculture and generation of a fibroblast cell line (LCF)**

Myofibroblasts which were obtained on day 9 of the explant cultures were further cultivated at low density on plastic under the same conditions during a period of one year (25 passages) to generate a cell line with a stable fibroblast phenotype. These cells were termed long culture fibroblasts (LCFs). Soft agar cloning of these cells was negative and excluded malignant transformation.

**Morphological analysis and detection of vitamin A uptake by LCFs**

Cells of three independent explant cultures were morphologically examined daily over a period of 21 days under an inverted microscope (Axiovert 35) using phase contrast illumination. The presence of vitamin A in cells outgrowing from explants was detected by the fast fading of green fluorescence of retinoids excited with 328 nm UV light. In the LCFs, the presence of vitamin A was detected after incubation with 5 μM of all trans-retinol over four days prior to the test.

**Immunofluorescence analyses**

Immunostaining of α-SMA, vimentin, desmin, GFAP, PDGFRB, p21cip1/WAF1, ROCK2, and ASK1 was performed as described previously. Fluorescence was analysed using a confocal laser scanning microscope (LSM-410, Carl Zeiss, Jena, Germany) equipped with a 63× oil immersion objective. Immunostaining for each detected protein was repeated three times in three independent explant and LCF cell cultures.

**Proliferation and apoptosis assays**

For analyses of proliferation and apoptosis, the BrdU incorporation assay and the TUNEL reaction were used, respectively. BrdU labelling and detection kit 1 and the in situ cell death detection kit were used according to the manufacturer’s instructions.

**Western analyses**

Immunoblots were performed from total cell lysates and from immunoprecipitates of cells on day 9 of the explant cultures (MFBS) and LCFs. Analyses were performed for Fn, Col I, PDGFRB, α-SMA, desmin, GFAP, CD 95L, DR-5, p21cip1/WAF1, phospho-MLC, MLC, phospho-p38, p38, phospho-JNK1/2, JNK1/2, Rock2, and ASK1. For total cell lysates, adherent cells were lysed using a detergent containing buffer (62.5 mM Tris HCl, pH 6.8, 5 mM EDTA, 2% sodium dodecyl sulphate, 10% glycerol, 2% β-mercaptoethanol). Equal amounts of total cellular protein or of immunoprecipitates were separated by sodium dodecyl sulphate-polyacrylamide gel electrophoresis and then transblotted to PVDF membranes. Membranes were incubated with appropriate primary antibodies overnight at 4°C followed by AP conjugated secondary antibody. Immunoblotting for each detected protein was repeated three times using lyses from three independent explants and LCF cell cultures.

**Cellular fractionation**

Briefly, cells were lysed in buffer A (10 mM HEPES, 1.5 mM MgCl₂, 50 mM KCl, 0.2 mM phenylmethylsulphonyl fluoride, and protease inhibitor cocktail) and after incubation on ice for 10 minutes cells were sonicated by ultrasound. Cellular fractionation was carried out as described previously. ²

**Immuono- and coimmunoprecipitation**

Immuono- and coimmunoprecipitation were performed using cytoplasmic and nuclear fractions. Briefly, lysates containing 350 μg of total cellular protein were precleared by incubation with protein A/G agarose for 10 minutes at 4°C and proteins were immunoprecipitated by incubation with appropriate antibodies (α antibodies per 350 μg of total cellular protein) overnight at 4°C followed by protein A/G agarose for two hours at 4°C. Pellets were washed twice with lysis buffer and then with kinase buffer (50 mM Tris, 50 mM NaCl, 10 mM MgCl₂, 25 mM β-glycerophosphate, 2 mM DTT, 0.2 mM PMSF). Complexes were disrupted in 2× Laemmli buffer and resolved in 10% gels. Immunoblotting for each detected protein was repeated three times using lyses from three independent cytoplasmic and nuclear fractions of MFB and LCF cells.

**In vitro kinase assay**

Rock2 and ASK1 proteins were immunoprecipitated from cytoplasmic fractions as described above. Beads were washed three times in kinase buffer and incubated in the presence of 10 μCi ³²P-ATP and 2.5 μg myelin basic protein (MBP). Reactions were incubated for 30 minutes at 30°C. The reaction products were spotted onto P81 phosphocellulose paper discs, dried overnight at room temperature, and quantified using a scintillation counter. Reactions for each immunoprecipitated protein were repeated three times from three independent cytoplasmic and nuclear fractions of MFB and LCF cells.

**Induction of oxidative stress and trypan blue staining for plasma membrane integrity**

MFBS and LCFs (0.5×10⁶) were cultured for 48 hours and then incubated with 200 μM H₂O₂ in culture medium for six and 12 hours followed by further culture for 12 hours without H₂O₂. To test the role of JNK activity in the apoptotic sensitivity to oxidative stress, cells were incubated with JNK inhibitor (1 μM) for three hours and then washed, followed by incubation with H₂O₂. MFBS and LCFs without H₂O₂ were used as controls.
treatment or treated only with JNK inhibitor were used as controls. Detached cells as well as adherent cells were stained with trypan blue solution, as described previously.  

In vivo models and histochemistry

Cerulein induced pancreatitis in rats was used as a model of acute pancreatic disease and generated as described previously. DBTC induced pancreatic fibrosis in rats was used as a model for established pancreatitis, and was generated as described previously.  

For immunohistochemistry, cryosections were fixed with 4% paraformaldehyde and then permeabilised with 0.1% Triton X-100 followed by incubation with anti-p21Cip1/WAF1 and anti-α-SMA antibodies for two hours. As secondary antibody, a rabbit anti-mouse AP conjugated antibody was used. After washing, FAST RED visualisation agent was added. Finally, the tissue was counterstained with haematoxylin. Cryosections incubated with a secondary antibody alone were used as negative controls (not shown).

RESULTS

Phenotype conversions of pancreatic fibroblast cells in primary explant cultures

Cells that grew out from the explants were examined daily over a period of 21 days, with particular interest in cellular dynamics and phenotype differentiation. After 24 hours in culture, the first cells were growing out from the explants (fig 1A–D). These cells had a fibroblastic shape and contained numerous lipid droplets in cytoplasm. During the following days, proliferating cells which incorporated BrdU in the S phase of the cell cycle were distributed throughout the population (data not shown). In the immunofluorescence experiments, fibroblasts on day 4 weakly expressed α-SMA in a granular pattern, mainly localised in the perinuclear region (fig 3A, 3G). After day 6, on the periphery, flattened cells with a great volume of cytoplasm and without processes dominated (fig 1D). These cells were characterised as MFBs due to α-SMA filaments in cytoplasm (see below), which were organised in parallel and distributed along the cell axis.

In MFBs analysed in the immunofluorescence experiments on day 9, α-SMA was expressed in stress fibres and colocalised with F-actin (fig 3B). Cells expressed filamentous vimentin and desmin (fig 3D–F) and PDGFRα was upregulated (fig 3K). From day 12 to 21, we again observed a change in the cellular phenotype on the periphery of the cell...
population (fig 1E-H). MFBs converted to cells that revealed a fibroblast phenotype with a decreased volume of cytoplasm and gradual package of stress fibres into 2–3 cytoplasmic processes (fig 1E, F). These fibroblastic cells appeared morphologically identical to the pancreatic fibroblasts on day 4 but did not contain lipid droplets (fig 1H). When analysed in the immunofluorescence experiments, these cells on day 19 showed weak and granular expression of \( \alpha \)-SMA (fig 3C), GFAP (fig 3I), and decreased expression of PDGFRB (fig 3L). In parallel, the majority of MFBs rounded up, condensed, and could be characterised as apoptotic cells on day 17 by the TUNEL assay (fig 1G, fig 2). These findings demonstrate recovery of pancreatic fibroblasts from MFBs in explant culture, which was accompanied by MFB apoptosis.

Long term subculture of pancreatic MFBs and generation of LCFs

To evaluate phenotype stability of the cells in vitro, pancreatic MFBs and fibroblasts were transferred from explant culture into tissue culture flasks on day 9 and 19, respectively. Due to the massive apoptosis of MFBs, only a few fibroblasts were generated on day 19 that could be harvested which maintained their phenotype on plastic for two days (fig 4B) followed by activation and conversion to MFBs. MFBs from day 9 (fig 4A) maintained their phenotype for 24 passages and then converted to fibroblasts (fig 4C). These fibroblasts

Figure 3  Differential expression of cytoskeletal proteins and platelet derived growth factor receptor type beta (PDGFRB) in the explant culture. (A) Weak and dot-like distribution of \( \alpha \) smooth muscle actin (\( \alpha \)-SMA) on day 4 and on day 19 (C). (B) High expression of \( \alpha \)-SMA in stress fibres on day 9 (control as in (F)). Filamentous organisation of vimentin (D) and desmin (E) in myofibroblast cells on day 9 (F) control with a secondary Alexa Fluor 488 labelled antibody alone. (G) Granular expression of glial fibrillary acidic protein (GFAP) on day 9 (control as in (F)). Expression of PDGFRB on days 3 (J), 9 (K), and 19 (L). Note the enhanced expression on day 9 (control as in (F)).

Figure 4  Generation of fibroblastic cells (long culture fibroblast (LCF) cell line) after passaging and subculture of myofibroblast (MFB) cells on plastic. (A) MFBs on day 9 of the explant culture transferred on plastic. (B) Fibroblasts on day 19 of the explant culture transferred on plastic. (C) Morphology of LCF cells which were generated in passage 25 of the MFBs isolated from the explant culture on day 9. All magnifications 100\( \times \). (D) Storage of vitamin A in the form of fat droplets in cytoplasm of the LCF cells after two days of incubation with 5 \( \mu \)M retinol. Magnification 320\( \times \) was used to emphasise fat droplet formation.
(LCF) exhibited a stable phenotype during further subcultivation. In LCFs, uptake of vitamin A and its storage in fat droplets were detected after incubation with trans-retinol (fig 4D). Immunofluorescence revealed the same characteristics as found in fibroblasts of the explant culture: granular expression of \( \alpha \)-smooth muscle actin (A), glial fibrillary acidic protein (B), desmin (C), and PDGFRB (D) were assessed (control as in fig 3F).

Pancreatic MFBs display nuclear while fibroblasts show cytoplasmic localisation of \( \alpha \)-SMA

To further evaluate possible mechanisms that could be responsible for inhibition of apoptosis and phenotype conversion of pancreatic MFBs to fibroblasts, we hypothesised a pathway that involves \( p21^{\text{Cip1/WAF1}} \) and could negatively regulate stress fibre formation and apoptotic sensitivity.\(^{22-24} \)

Using immunofluorescence, we found that MFBs expressed \( p21^{\text{Cip1/WAF1}} \) exclusively in the nucleus whereas both fibroblasts on day 19 in the explant culture and LCFs contained \( p21^{\text{Cip1/WAF1}} \) also in the cytoplasm (fig 7). In LCFs, cytoplasmic localisation was confirmed in western blots (fig 7). These findings suggest that phenotype conversion of MFBs to fibroblasts was associated with translocation of \( p21^{\text{Cip1/WAF1}} \) from the nucleus into the cytoplasm.

Cytoplasmic \( p21^{\text{Cip1/WAF1}} \) interacts with Rock2 and ASK1 in LCFs which correlates with inhibition of the activities of Rock2 and ASK1

To further support a role for cytoplasmic \( p21^{\text{Cip1/WAF1}} \) in the conversion of MFBs to fibroblasts, we tested its association with Rock2 and ASK1. Rock2 is one of the three downstream mediators of GTP-Rho and plays an important role in stress fibres and focal adhesion formation through phosphorylation of the MLC.\(^{30} \) ASK1 is a mitogen activated protein kinase kinase and involved in the cellular response to Fas, tumour necrosis factor receptor, and oxidative stress stimulation through activation of the JNK1/2 and p38 pathways.\(^{31-33} \) As detected by immunofluorescence, Rock2 and ASK1 were localised together with \( p21^{\text{Cip1/WAF1}} \) in the cytoplasm of both LCFs and fibroblasts on day 19 of the explant culture but not in MFBs (fig 7). In LCFs, cytoplasmic localisation of both proteins was confirmed in western blots (fig 7). These findings suggest that phenotype conversion of MFBs to fibroblasts was associated with translocation of cytoplasmic \( p21^{\text{Cip1/WAF1}} \) from the nucleus to the cytoplasm.

In LCFs, \( p21^{\text{Cip1/WAF1}} \) was detected in immunoprecipitates of Rock2 and ASK1 in the cytoplasm of both LCFs and fibroblasts (fig 7). Furthermore, \( p21^{\text{Cip1/WAF1}} \) was only detected in immunoprecipitates of Rock2 and ASK1 from cytoplasmic fractions of LCFs and not from MFBs (fig 8) which shows that \( p21^{\text{Cip1/WAF1}} \) binds to Rock2 and ASK1 in the cytoplasm of LCFs. To investigate the effect of \( p21^{\text{Cip1/WAF1}} \) on phosphorylation activity, cytoplasmic immunoprecipitates of Rock2 and ASK1 were subjected to a kinase assay using MBP, a previously reported suitable substrate for Rock2 and ASK1.\(^{22-24} \) We measured decreased kinase activities of both Rock2 and ASK1 in LCFs compared with MFBs (fig 8). These results suggest that cytoplasmic \( p21^{\text{Cip1/WAF1}} \) in LCFs forms a complex with Rock2 and ASK1, which correlates with inhibition of phosphorylation activities of both tested serine-threonine kinases.

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**Figure 5** Immunofluorescence analysis of cytoplasmic proteins and platelet derived growth factor receptor type beta (PDGFRB) in the long culture fibroblast (LCF) cells. Weak and granular expression of \( \alpha \)-smooth muscle actin (A), glial fibrillary acidic protein (B), desmin (C), and PDGFRB (D) were assessed (control as in fig 3F).

**Figure 6** Western analysis of protein expression in myofibroblast (MFB) (1) and long culture fibroblast (LCF) cells (2). Note the marked downregulation in expression of cytoskeletal, cell surface, and extracellular matrix proteins in LCF cells compared with MFB cells. Immunoblots for each detected protein were repeated three times using lysates from three independent explant and LCF cell cultures. Col, collagen; DR, death receptor; FN, fibronectin; GFAP, glial fibrillary acidic protein; PDGFRB, platelet derived growth factor receptor type beta; \( \alpha \)-SMA, smooth muscle actin.
Cytoplasmic p21\textsuperscript{Cip1/WAF1} correlates with reduced phosphorylation of MLC, p38 and JNK1/2 in LCFs

To see whether decreased kinase activities of Rock2 and ASK1 immunoprecipitates, which we found in LCFs, were associated with lower activation of downstream proteins in the stress activated pathways, we examined phosphorylation of MLC, p38, and JNK1/2. We revealed decreased phosphorylation of these proteins in LCFs compared with MFBs, whereas levels of protein expression were identical in both cell phenotypes (fig 9). These findings suggest that...
cytoplasmic expression of p21\textsuperscript{Cip1/WAF1} correlates with a decrease in MLC, p38, and JNK1/2 phosphorylation in LCFs.

**Different sensitivities to oxidative stress in MFBs and LCFs which depend on JNK1/2 activation**

To investigate whether the apoptotic sensitivity of pancreatic MFBs in culture can be explained by increased ASK1 activity which is followed by downstream activation of JNK1/2, we incubated MFBs and LCFs with 200 \(\mu\)M \(H_2O_2\) to induce oxidative stress which was shown to induce ASK1 activation and induction of apoptosis in several cell types.\(^{37}\) We found an increased proportion of cells that detached from the substrate due to \(H_2O_2\) in MFBs compared with LCFs (fig 10). Trypan blue staining of cells revealed a permeable membrane. Furthermore, we inhibited activation of JNK1/2 using a concentration of inhibitor proposed by the manufacturer which revealed reduced sensitivity of MFBs to oxidative stress. LCFs remained unaffected. This indicates that this apoptotic sensitivity depends on JNK1/2 activation in MFBs (fig 10). Together, these findings suggest decreased apoptotic sensitivity in LCF cells compared with MFBs due to the low activity of ASK1 and reduced activation of JNK1/2 (see results above).

**Pancratic MFBs and fibroblasts show different p21\textsuperscript{Cip1/WAF1} localisation in experimental acute pancreatitis and pancreas fibrosis**

To test the hypothesis that the phenotype conversion from MFBs to fibroblasts combined with cytoplasmic expression of p21\textsuperscript{Cip1/WAF1} that we have found in cell culture also occurs in pancreatic fibrosis, we examined expression of \(\alpha\)-SMA and p21\textsuperscript{Cip1/WAF1} in cryosections from normal rat pancreas, experimental cerulein induced acute pancreatitis, and DBTC induced established pancreatic fibrosis (fig 11). Consistent with our findings in vitro, interstitial cells in cerulein induced acute pancreatitis demonstrated prominent anti-\(\alpha\)-SMA immunoreactivity and nuclear localisation of p21\textsuperscript{Cip1/WAF1} whereas weak anti-\(\alpha\)-SMA immunoreactivity and cytoplasmic expression of p21\textsuperscript{Cip1/WAF1} were detected in fibroblasts distributed throughout fibrotic bands in DBTC induced established pancreatic fibrosis. In the normal pancreas, only a few fibroblasts were positive for p21\textsuperscript{Cip1/WAF1} which revealed cytoplasmic expression.

**DISCUSSION**

The principle finding of this study was that in a culture model, pancreatic MFBs were transient and disappeared by apoptosis or converted to fibroblasts. This phenotype conversion to fibroblasts was documented by corresponding changes in morphology, expression, and organisation of cytoskeletal proteins, cell surface receptors, and ECM proteins, and by a lower apoptotic sensitivity. Moreover, our histochemical results in the cerulein induced model of acute pancreatic disease and in the DBTC induced model of established pancreatic fibrosis revealed the relevance of our in vitro findings for tissue reactions in vivo. Prominent \(\alpha\)-SMA expression as a main histopathological marker of MFB transdifferentiation was found exclusively in cryosections from experimental acute pancreatitis whereas in established pancreatic fibrosis, weak \(\alpha\)-SMA immunoreactivity or even immunonegativity was a characteristic feature.
Given that recovery of fibroblasts from pancreatic MFBs is characterised by significant alterations in the cytoskeleton and increase in resistance to apoptotic stimuli, we believe that a pathway activated during this phenotype conversion negatively regulates stress fibre formation and apoptotic sensitivity. It was previously reported that p21Cip1/WAF1, an inhibitor of cell cycle progression, exhibits such functions in different cell types through its ectopic cytoplasmic localisation. Moreover, cytoplasmic expression of p21Cip1/WAF1 in MFBs correlated with hypophosphorylation of MLC, JNK1/2, and p21Cip1/WAF1 translocation from the nucleus to the cytoplasm. Investigating the molecular mechanisms in more detail, in LCFs we found a complex of cytoplasmic p21Cip1/WAF1 with Rock2 and ASK1 which correlated with decreased phosphorylation of these proteins in a kinase assay. Moreover, cytoplasmic expression of p21Cip1/WAF1 in MFBs correlated with hypophosphorylation of MLC, JNK1/2, and p21Cip1/WAF1 translocation from the nucleus to the cytoplasm.

REFERENCES


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