INTESTINAL MICROFLORA AND IMMUNITY

Gut microbiota composition and development of atopic manifestations in infancy: the KOALA Birth Cohort Study

John Penders, Carel Thijs, Piet A van den Brandt, Ischa Kummeling, Bianca Snijders, Foekje Stelma, Hanne Adams, Ronald van Ree, Ellen E Stobberingh

Innocuous antigens later in life. However, an alternative during childhood results in aberrant immune responses to associated with a Western lifestyle are thought to be involved in the atopic diseases. Although genetic susceptibility plays an interleukin (IL)-4, IL-13 and IL-5 contribute to the induction of immune response and the elaboration of cytokines such as...
and atopIC manifestations were collected for all members of the cohort by repeated questionnaires at 34 weeks of gestation and at 3, 7, 12 and 24 months post partum.

Participants (with both conventional and alternative lifestyles) recruited from January 2002 onwards were asked to sample the infant’s faeces at the age of 1 month post partum (n = 1176). Subjects received a faeces tube with a spoon attached to the lid (Sarstedt, Nümbrecht, Germany), together with a sanitary napkin, an instruction form and a brief questionnaire (faeces questionnaire). Parents placed a sanitary napkin in the diaper (to prevent absorption of the faeces by the diaper), collected the faeces out of the napkin into the collection tube and sent it immediately to our laboratory by post. The short time was minimised by asking the parents to collect the faeces on a Monday, Tuesday or Wednesday, so that the samples did not remain in the mail service over the weekend.

The study cohort comprised 957 infants after exclusion of premature infants, infants who received antimicrobial agents during their first month of life, infants from whom insufficient amounts of faeces (< 1 g) were collected, infants whose faeces were not collected between 3 and 6 weeks of age and infants for whom the faeces questionnaire was missing.

Home visits were made by trained nurses when the infant was 2 years of age (n = 607). During these visits a clinical diagnosis of atopIC dermatitis was determined using the UK-Working Party (UK-WP) criteria.22–24 Venous blood was also collected from the infants in order to determine total (n = 590) and specific (n = 583) serum immunoglobulin (IgE).

The KOALA study was approved by the Ethics Committee of the University Hospital of Maastricht and all parents signed informed consent for the study.

**DNA purification from faeces**

At the laboratory faecal samples were diluted tenfold in peptone-water (Oxoid CM0009) containing 20% v/v glycerol (Merck, Darmstadt, Germany) and stored at −20 °C until analysis. For DNA isolation, 0.2 ml of the diluted faeces was added to a 2 ml vial containing approximately 300 mg glass beads (diameter 0.1 mm) and 1.4 ml of ASL-buffer from the QIAamp DNA Stool Mini Kit (Qiagen, Hilden, Germany) and the samples disrupted in a mini-bead beater (Biospec Products, Bartlesville, OK) at 5000 rpm for 3 min. Subsequently, bacterial DNA was isolated from the samples using the QIAamp DNA Stool Mini Kit according to the instructions of the manufacturer. The DNA was eluted in a final volume of 200 μl.

**Microbial analysis of faecal samples**

Faecal samples were subjected to real-time PCR for counting of *Bifidobacterium* spp, *Escherichia coli*, *Clostridium difficile*, *Bacteroides fragilis* group, *Lactobacillus* spp, and total bacterial counts as described previously25 (primers and probes are listed in table 1). The log_{10} colony forming units (CFU) per gram of the bacterial groups and species were calculated for each stool sample from the threshold cycle values using the constructed standard curves. The prevalence of colonisation was expressed as the percentage of infants colonised with a specific bacterial group or species.

**Determination of total and specific IgE**

Blood samples collected during home visits when the infant was 2 years of age were analysed for total and specific serum IgE by Sanquin Research (Amsterdam). For total IgE levels, samples were analysed as described earlier.33 34 A sandwich RIA was used for values <150 IU/ml and a competitive RIA for values >150 IU/ml.35 The detection limit for total serum IgE was <0.5 IU/ml. Blood samples were analysed for specific IgE against ben’s egg, cow’s milk, peanuts, birch pollen, grass pollen, cat, dog and house dust mite using RAST as described previously.33 Calculation was performed by means of a standard curve that was obtained by RAST with a dilution series of a chimeric monoclonal IgE antibody against the major house dust mite allergen Der-p2 and Sepharose-coupled recombinant Der-p2.33 A RAST value of >0.3 IU/ml was regarded as positive.

**Definition of atopIC manifestations and sensitisation**

Our definitions of eczema and wheeze were based on questions adapted from the ISAAC questionnaires.26 In the 7, 12 and 24 months’ postpartum questionnaires, parents were asked: “Has your child ever had an itchy rash that came and went over the past months?”. If this question was answered affirmatively, infants were defined as having developed eczema in the first 2 years of life. Infants for whom only diaper rash, rash around the eyes and/or scalp scaling was reported were not regarded as having developed eczema.

“Recurrent wheezing” was defined as the reported presence of wheezing with at least four attacks between 0 and 7 months mentioned in the 7 months’ postpartum questionnaire and/or between 7 and 12 months of life mentioned in the 12 months’ postpartum questionnaire and/or between 13 and 24 months mentioned in the 24 months’ postpartum questionnaire.

Besides the eczema reported by the parents, a diagnosis of atopIC dermatitis could be made using the UK-WP criteria for those infants visited at home.22–24 Briefly, the presence of atopIC dermatitis was determined according to four clinical symptoms: (i) the presence of an itchy rash, (ii) a history of flexural dermatitis, (iii) visible flexural dermatitis, and (iv) onset before the age of 2 years. In this study, infants with a probability of atopIC dermatitis of >0.9 were considered to have atopIC dermatitis.

Infants with specific serum IgE levels >0.3 IU/ml against one or more of the tested food or inhalant allergens were considered to be sensitised.

**Definition of potential confounders**

The following variables were included as potential confounders: subcohort (conventional; alternative), parenteral atopIC history defined as self-reported doctor-diagnosed eczema, hay fever, asthma, and pet and/or house dust mite allergy (none; at least one parent), siblings with an atopIC history defined as parent-reported doctor-diagnosed food allergy, eczema, hay fever, asthma, and pet and/or house dust mite allergy (no siblings; ≥1 sibling, none atopIC; or ≥1 sibling, at least one atopIC), age at collection of faecal sample (age in days) and infant’s gender (boy; girl). Maternal probiotic use during the last month of pregnancy (never/sporadic; several times a month; several times a week; daily), place and mode of delivery (vaginal delivery at home; vaginal delivery in hospital; artificial delivery in hospital; caesarean section in hospital), type of infant feeding during the first month (exclusively breast-fed; exclusively formula-fed or a combination) were included as a separate set of confounders. These latter variables may be more distal determinants in the association between the gut microbiota and atopIC manifestations instead of true confounders, and adjusted analyses were therefore performed with and without adjustment for these variables.

**Statistical analyses**

Logistic regression analyses were used to test for (unadjusted) associations between colonisation with the gut bacteria (colonised or uncolonised) under study and the development of atopIC outcomes (eczema (reported by parent), atopIC dermatitis (UK-WP criteria), recurrent wheeze and/or atopIC sensitisation). Adjusted associations were tested by incorporating...
the potential confounders into the logistic regression models. Two different sets of confounders were used. The first set included the variables subcohort, parental and sibling atopic history, age at collection of the faecal sample and infant’s gender, and the second set included these variables as well as maternal probiotic use during pregnancy, place and mode of delivery and type of infant feeding. Since almost all infants were colonised, the association between colonisation with bifidobacteria and atopic outcomes was analysed as “low” (<10.68 log_{10} CFU/g) versus “high” (≥10.68 log_{10} CFU/g) bifidobacterial counts; the few uncolonised infants (n = 12) were added to the low count groups. The cut-off point of 10.68 log_{10} CFU/g was chosen to create two equal groups.

Logistic regression analyses were also used to test for associations between the concentration (counts) of gut bacteria and atopic outcomes. We additionally adjusted for total bacterial counts to account for differences in the consistency of faecal samples in the analyses on bacterial numbers, in addition to the confounders described above. To test for trend, bacterial counts were categorised (uncolonised infants were used as the reference category and the remaining colonised infants were accommodated in two (C difficile, lactobacilli) or three equal groups (bifidobacteria, E coli, B fragilis group)).

Linear regression analyses were used to test for associations between the gut bacteria under study and total serum IgE levels, controlling for the same confounders mentioned above. Since separate analyses of the conventional and alternative subcohorts showed that the key findings were similar within these two subcohorts, in the final analyses we combined the two groups adjusting for “subcohort”.

To examine the possibility of selection bias in those infants visited at home, we performed non-response analyses. Using logistic regression analyses, infants visited at home (n = 607) were compared with infants not visited at home (n = 305) regarding gut microbiota composition and the prevalence of eczema and recurrent wheeze.

**RESULTS**

Of the 957 infants participating in this study, almost all were colonised with bifidobacteria (98.7%) at the age of 1 month (table 2). Most of the infants were also colonised with E coli (88.6%) and members of the B fragilis group (81.6%), whereas colonisation with lactobacilli (32.2%) and C difficile (25.1%) was less common. Bifidobacteria were detected in the highest numbers, followed by E coli and B fragilis group species.

More than half of the infants had at least one parent with a positive history of doctor-diagnosed atopic manifestations (table 2). Almost one third of the infants who had older siblings, had an atopic sibling. Over 30% of the infants had developed eczema at the age of 2 years and recurrent wheeze was reported in approximately 10%. Over a quarter of the infants had circulating IgE antibodies (>0.3 IU/ml) in their blood against one or more of the food and/or inhalant allergens and thus were regarded as sensitised.

Non-response analyses showed that there were no differences in the composition of gut microbiota or in the prevalence of eczema and recurrent wheeze between infants who were visited at home and those who were not (data not shown).

Separate analyses of the conventional and alternative sets of confounders were used. The first set included these variables as well as maternal probiotic use during pregnancy, place and mode of delivery and type of infant feeding. Since separate analyses of the conventional and alternative subcohorts showed that the key findings were similar within these two subcohorts, in the final analyses we combined the two subcohorts showing that the key findings were similar within these two subcohorts, in the final analyses we combined the two groups adjusting for “subcohort”.

**Table 1 Primes and probes used in this study**

<table>
<thead>
<tr>
<th>Target organisms (amplicon size)</th>
<th>Primer/probe</th>
<th>Sequence(5’-3’)</th>
<th>Tm (°C)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bifidobacterium spp (126 bp)</td>
<td>Forward primer</td>
<td>GCCGCTCTAAACATCAGAACGC</td>
<td>59</td>
<td>Penders et al</td>
</tr>
<tr>
<td></td>
<td>Reverse primer</td>
<td>CACCGGTTCCCGAGGCTATT</td>
<td>59</td>
<td>Penders et al</td>
</tr>
<tr>
<td></td>
<td>Probe</td>
<td>TCAAGCATTTCTACCGTCGCGC</td>
<td>70</td>
<td>Huijdenk et al</td>
</tr>
<tr>
<td>Escherichia coli (96 bp)</td>
<td>Forward primer</td>
<td>CAGCCGCGCCTGATGAAAGA</td>
<td>59</td>
<td>Huijdenk et al</td>
</tr>
<tr>
<td></td>
<td>Reverse primer</td>
<td>CGGTAAATCATGACGCCCCAAA</td>
<td>69</td>
<td>Huijdenk et al</td>
</tr>
<tr>
<td></td>
<td>Probe</td>
<td>TTAATACCTACTTGCCCTCCGCGTGAA</td>
<td>68</td>
<td>Huijdenk et al</td>
</tr>
<tr>
<td>Clostridium difficile (114 bp)</td>
<td>Forward primer</td>
<td>TTGACGATTATCCCTGGTAAAGA</td>
<td>58</td>
<td>Penders et al</td>
</tr>
<tr>
<td></td>
<td>Reverse primer</td>
<td>TGTACTGCGCTACCTTGTATATC</td>
<td>59</td>
<td>Penders et al</td>
</tr>
<tr>
<td>Bacteroides fragilis group (92 bp)</td>
<td>Forward primer</td>
<td>CACACCTCGTACACCCGTCGG</td>
<td>69</td>
<td>Penders et al</td>
</tr>
<tr>
<td></td>
<td>Reverse primer</td>
<td>CGGCAAAATCTACAACGTACTA</td>
<td>59</td>
<td>Liu et al</td>
</tr>
<tr>
<td></td>
<td>Probe</td>
<td>CGCTCCCTTAAAACCCGAAAACCCTGGG</td>
<td>70</td>
<td>Liu et al</td>
</tr>
<tr>
<td>Lactobacillus spp (341 bp)</td>
<td>Forward primer</td>
<td>AGGACGTAGGGGATCTCCTGCA</td>
<td>59</td>
<td>Wolfle et al</td>
</tr>
<tr>
<td></td>
<td>Reverse primer</td>
<td>CAGCCTGACCTGAGGCGACGAG</td>
<td>59</td>
<td>Rintila et al</td>
</tr>
<tr>
<td></td>
<td>Probe</td>
<td>TCTACCCGGAGGGCAGCAGT</td>
<td>59</td>
<td>Huijdenk et al</td>
</tr>
<tr>
<td>Total count (467 bp)</td>
<td>Forward primer</td>
<td>CGGCTACGGGGTATCTAACCTTGTT</td>
<td>58</td>
<td>Nadkarni et al</td>
</tr>
<tr>
<td></td>
<td>Reverse primer</td>
<td>GGACTAACGGGGTATACCTTGTT</td>
<td>58</td>
<td>Nadkarni et al</td>
</tr>
</tbody>
</table>
eczema, atopic dermatitis, recurrent wheeze and atopic sensitisation was not influenced by the concentration of this bacterium in the faecal samples. Furthermore, counts of bifidobacteria, *B. fragilis* group species and lactobacilli were not associated with atopic disease manifestation or sensitisation.

Total serum IgE level at the age of 2 years was not associated with colonisation rates or counts of gut bacteria.

**DISCUSSION**

This prospective study demonstrates that differences in gut microbiota composition in early infancy are associated with the subsequent development of atopic disease manifestation and sensitisation. The presence of *E. coli* was associated with a higher risk of developing eczema, this risk being increased with increasing numbers of *E. coli*. However, this association was not found when atopic dermatitis was defined according to the UK-WP criteria. Colonisation with *C. difficile* in early infancy was associated with an increased risk of all atopic outcomes (eczema, atopic dermatitis, recurrent wheeze and atopic sensitisation), independent of the concentration of this bacterium.

This is the first large-scale prospective study on gut microbiota composition in relation to atopic manifestations. Although previous epidemiological studies were very informative and were the first to suggest a potential role of the gut microbiota in the aetiology of atopic diseases, they were often based on small populations and were not able to adjust for potential confounders. Furthermore, only two previous studies were prospective and therefore able to determine if differences in the gut microbiota precede the development of atopic symptoms. Another strength of our study is the molecular techniques used, which overcome many of the problems associated with traditional bacteriological culture. Analyses of the gut microbiota using bacteriological culture are biased, since many selective culture media are not absolutely selective. Furthermore, these media do not equally support the growth of the different species comprising a population, and not all bacteria are cultivatable. The real-time PCR assays used in the present study are a quantitative culture-independent approach suitable for high-throughput analyses of both fresh and frozen samples.

At present, faeces is the only realistic sample available in large non-invasive epidemiological studies on the gut microbiota. However, a limitation of using faecal samples is that the bacterial composition in the lumen does not reflect the composition of bacteria adhering to the mucosa, and furthermore the composition of bacteria differs throughout the intestinal tract. Nevertheless, it can be assumed that although the proportions and activities of the microbiota change with passage through the intestinal tract, most viable and non-viable

---

**Table 2** Characteristics of the participants in this study

<table>
<thead>
<tr>
<th></th>
<th>Conventional subcohort, n = 652*</th>
<th>Alternative subcohort, n = 305*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Prevalence of colonisation with intestinal bacteria, %</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Bifidobacteria</em></td>
<td>98.3</td>
<td>99.7</td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>89.5</td>
<td>86.6</td>
</tr>
<tr>
<td><em>Clostridium difficile</em></td>
<td>24.7</td>
<td>25.9</td>
</tr>
<tr>
<td><em>Bacteroides fragilis</em></td>
<td>83.0</td>
<td>78.7</td>
</tr>
<tr>
<td><em>Lactobacilli</em></td>
<td>33.1</td>
<td>30.2</td>
</tr>
<tr>
<td>Counts of intestinal bacteria (log10 CFU/g), median (range)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Bifidobacteria</em></td>
<td>10.71 (6.84–11.56)</td>
<td>10.68 (6.85–11.49)</td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>9.45 (5.91–10.79)</td>
<td>9.12 (5.92–10.62)</td>
</tr>
<tr>
<td><em>Clostridium difficile</em></td>
<td>5.12 (2.70–8.41)</td>
<td>5.70 (2.85–8.81)</td>
</tr>
<tr>
<td><em>Bacteroides fragilis</em></td>
<td>9.40 (5.74–10.36)</td>
<td>9.07 (5.79–10.33)</td>
</tr>
<tr>
<td><em>Lactobacilli</em></td>
<td>8.70 (7.92–10.73)</td>
<td>8.58 (7.95–10.33)</td>
</tr>
<tr>
<td><em>E. coli</em></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Overall numbers are not always 652 for the conventional subcohort and 305 for the alternative subcohort due to missing bacterial count data or outcome data; consumed during the last month of pregnancy; forceps or vacuum extraction; totally available for those infants visited at home, total numbers are 391 for conventional subcohort and 192 for alternative subcohort; *only available for those infants visited at home, total numbers are 396 for conventional subcohort and 194 for alternative subcohort.*
intestinal bacteria will still be detectable in the faeces by molecular methods. 90 A drawback of this study is the time second, the age of 1 month post partum. There are several major importance. Second, at the age of 1 month colonisation is complete and although the composition may fluctuate, large shifts in the composition do not occur until weaning. 23 Finally, as we wanted to exclude the chance of reverse causation, the gut microbiota composition had to be analysed prior to the manifestation of atopic symptoms.

Since manifestation of atopic symptoms and sensitisation to allergens do not always occur together (sensitised infants do not always show symptoms and infants with symptoms are not always sensitised), we chose to report the manifestation of atopic symptoms and sensitisation as separate outcome parameters. As a consequence we decided to use the term eczema instead of atopic eczema or atopic dermatitis. However, we did use the term atopic dermatitis for those infants visited at home who fulfilled the UK-WP criteria, since Williams and colleagues when introducing these criteria originally used the term atopic dermatitis. 24

The positive association we found between Escherichia coli and eczema is difficult to compare with previous reports, since several of these studies did not determine E coli or measured this bacterium as part of total coliforms, enterobacteriaceae or Gram-negative rods. 13

In contrast to the association between E coli and eczema (based on parents’ reports in questionnaires), we did not find an association between E coli and atopic dermatitis based on the UK-WP criteria. Selection bias could not explain this difference since non-response analyses showed that the gut microbiota and the prevalence of eczema and recurrent wheeze was similar for those infants who were visited at home and those who were not. The percentage of infants with atopic dermatitis according to the UK-WP criteria is much lower than the percentage of infants with eczema reported by parents. This can be explained by the fact that eczema reported by parents is based upon the presence of this condition at any time during the infants’ first 2 years of life, whereas many of these infants are probably in remission or have already outgrown this condition at time of the home visit. Therefore, it is possible that E coli is only associated with a milder eczematous condition that has already disappeared by the time of the home visit. Another explanation for these discrepant findings is that the UK-WP criteria included specific predilection sites of atopic dermatitis (flexural involvement), whereas the questionnaire data were based on the presence of an itchy rash anywhere (except diaper rash, rash around the eyes and scalp scaling). The increased risk of

---

Table 3  The adjusted association between colonisation with gut bacteria at 1 month of age and atopic sensitisation and atopic disease manifestation at 2 years of age (n = 957)

<table>
<thead>
<tr>
<th>Intestinal bacteria</th>
<th>Eczema</th>
<th>Recurrent wheeze</th>
<th>Atopic dermatitis (UK-WP)</th>
<th>Sensitisation</th>
<th>Odds ratio (95% CI)</th>
<th>p for trend</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Prevalence [n/N]</td>
<td>ORadj (95% CI)</td>
<td>Prevalence [n/N]</td>
<td>ORadj (95% CI)</td>
<td>Prevalence [n/N]</td>
<td>ORadj (95% CI)</td>
</tr>
<tr>
<td>Hib *alo</td>
<td>Low</td>
<td>32.4% (148/457)</td>
<td>9.4% (41/434)</td>
<td>1.0</td>
<td>17.9% (54/302)</td>
<td>1.0</td>
</tr>
<tr>
<td>High</td>
<td>22.2% (20/91)</td>
<td>5.8% (6/104)</td>
<td>1.0</td>
<td>15.9% (10/63)</td>
<td>1.0</td>
<td>30.6% (19/62)</td>
</tr>
<tr>
<td>E coli</td>
<td>No</td>
<td>32.5% (156/480)</td>
<td>1.02 (0.77 to 1.35)</td>
<td>11.8% (55/466)</td>
<td>1.32 (0.85 to 2.06)</td>
<td>14.4% (44/303)</td>
</tr>
<tr>
<td>Yes</td>
<td>33.8% (279/826)</td>
<td>1.87 (1.05 to 3.36)</td>
<td>11.1% (88/793)</td>
<td>1.92 (0.80 to 4.59)</td>
<td>16.2% (88/542)</td>
<td>1.02 (0.49 to 2.10)</td>
</tr>
<tr>
<td>C difficile</td>
<td>No</td>
<td>30.3% (213/702)</td>
<td>1.0</td>
<td>9.4% (63/671)</td>
<td>1.0</td>
<td>14.1% (63/446)</td>
</tr>
<tr>
<td>Yes</td>
<td>38.7% (91/235)</td>
<td>1.40 (1.02 to 1.91)</td>
<td>14.8% (30/207)</td>
<td>1.75 (1.18 to 2.57)</td>
<td>22.0% (34/153)</td>
<td>1.73 (1.08 to 2.78)</td>
</tr>
<tr>
<td>Lactobacilli</td>
<td>F grade</td>
<td>32.0% (55/172)</td>
<td>1.0</td>
<td>9.0% (15/166)</td>
<td>1.0</td>
<td>12.5% (14/112)</td>
</tr>
<tr>
<td>Yes</td>
<td>22.5% (249/765)</td>
<td>1.02 (0.71 to 1.47)</td>
<td>11.0% (81/734)</td>
<td>1.20 (0.66 to 2.18)</td>
<td>17.0% (84/493)</td>
<td>1.41 (0.76 to 2.60)</td>
</tr>
<tr>
<td>E coli</td>
<td>No</td>
<td>31.2% (199/637)</td>
<td>1.0</td>
<td>10.3% (63/613)</td>
<td>1.0</td>
<td>15.7% (64/408)</td>
</tr>
<tr>
<td>Yes</td>
<td>35.0% (105/300)</td>
<td>1.23 (0.91 to 1.65)</td>
<td>11.5% (33/287)</td>
<td>1.22 (0.77 to 1.93)</td>
<td>17.1% (34/199)</td>
<td>1.14 (0.72 to 1.81)</td>
</tr>
</tbody>
</table>

*Based on parents’ reports in 7, 12 and/or 24 months’ questionnaires, 16% determined by trained nurses during home visits, 16% specific IgE antibodies to at least one allergen (cow’s milk, hen’s egg, peanut, birch pollen, grass pollen, cat, dog, house dust mite). Numbers in table do not always add up to 957 due to missing bacterial count data or outcome data. 16 From logistic regression analysis: adjusted for subcohort, parental history of atopy, sibling history of atopy, age at collection of faecal sample and infant’s gender; 10 colonisation with bifidobacteria in association with atopic sensitisation and disease manifestation presented as low (<10^6 CFU/g), high (10^6–10^7 CFU/g) counts since almost all infants were colonised; uncolonised infants (n=12) were added to the low counts group.

---

Figure 1  Adjusted association between numbers of Escherichia coli (log_{10} CFU/g faeces) in faecal samples of 1 month old infants and the development of eczema in the first 2 years of life (adjusted for subcohort, parental history of atopy, sibling history of atopy, age at collection of faecal sample, infant’s gender and total bacterial count). Dots represent the odds ratios, bars represent the 95% confidence intervals. Low: <8.86 log_{10} CFU; middle: 8.86 to 9.75 log_{10} CFU; high: >9.75 log_{10} CFU.
eczema in infants colonised with *E. coli* may therefore also be limited to eczema other than at the specific predilection sites for atopic eczema/dermatitis and thus may be non-atopic. This idea is supported by the fact that we also found no association between *E. coli* and sensitisation.

In contrast, *C. difficile* appears to be associated with a higher risk of atopic eczema since a positive association was also found for atopic dermatitis according to UK-WP criteria and for sensitisation. Our findings of an association between *C. difficile* and atopy are in agreement with several previous studies. In a study on microflora-associated characteristics, allergic infants had higher levels of i-caproic acid in their stools compared with non-allergic infants. This short chain fatty acid is suggested to indicate the presence of *C. difficile*. Two studies used IgG serology against *C. difficile*. Woodcock and colleagues found increased specific IgG against *C. difficile* in sensitised wheezy infants compared with non-sensitised non-wheezy infants. Linneberg and colleagues found a positive association between IgG seropositivity against *C. difficile* and both allergic rhinitis and sensitisation.

Furthermore, several studies found an association between high numbers of the genus *Clostridium* and atopic dermatitis and/or sensitisation, although others did not find such an association. The genus *Clostridium* is a very heterogeneous group comprising several different clusters. It seems therefore unlikely that the members of such a phylogenetically diverse genus like *Clostridium* all have the same effect on the human host. It is more likely that certain species (such as *C. difficile*) or a certain cluster of species within this genus is responsible for the increased risk of developing atopic manifestations.

In contrast to several other studies, we did not find a negative association between allergies and bifidobacteria. A possible explanation is the lack of contrast in our study with respect to bifidobacterial counts, because almost all infants were colonised with high numbers of bifidobacteria. This is probably the consequence of the very young age of our population, an age at which bifidobacteria are known to dominate the gut microbiota.

Altogether our results support a role of the gut microbiota in the aetiology of atopic diseases. There are several hypotheses by which the associations we found between *C. difficile* and *E. coli* and atopic manifestations could be explained. First of all it should be noted that many of the bacteria in the gut are still unknown; differences in *E. coli* and *C. difficile* colonisation found in our study could therefore also reflect differences in other unknown bacteria.

The presence of *E. coli* and *C. difficile* could be associated with a decrease in other (unknown) beneficial bacteria. This could result in reduced induction of Treg cells by these beneficial bacteria leading to immune dysregulation. In the absence of optimal levels of immune regulation, an individual may develop a Th1 (such as *Crohn’s disease* or autoimmunity) or Th2 (such as atopic diseases) mediated inflammatory disorder depending on their own Th1/Th2 bias, immunological history and genetic background. Secondly, *E. coli* and *C. difficile* could have a direct effect on the production of cytokines by antigen-presenting cells, thereby affecting the differentiation of T cells. Another hypothesis is that *E. coli* and/or *C. difficile* increase the intestinal permeability (for instance by the production of toxins). This increased permeability of the intestinal barrier could facilitate the penetration of innocuous antigens and subsequent sensitisation. Indeed it has been shown that *C. difficile* toxins A and B compromise the intestinal cell barrier. Furthermore, increased intestinal permeability has been described in patients with food allergies, eczema and asthma compared with healthy subjects. Finally, it has also been suggested that infants susceptible to the development of allergies are also susceptible to aberrant colonisation of the gut. However, this explanation seems less likely since we have controlled for familial history of atopy. Additionally, the fact that differences in the gut microbiota are already present at such a young age preceding atopic symptoms makes this hypothesis less likely.

The consistent findings of a positive association between *C. difficile* and all atopic symptoms as well as sensitisation strengthen the probability of a causal relationship between the gut microbiota and atopy, and support the potential role of probiotics in the prevention and treatment of these diseases. We have previously examined the external factors influencing the composition of the gut microbiota in early infancy. Caesarean section, antibiotic therapy, hospitalisation and formula feeding all caused perturbations in the gut microbiota by increasing colonisation rates and counts of *E. coli* and *C. difficile* and/or by decreasing numbers of bifidobacteria. In particular, when the gut microbiota is disrupted by one or more of these external factors, probiotics may be effective in the treatment or prevention of atopic diseases.

Perturbations in the gut microbiota may also be related to other atopic outcomes which manifest at older ages such as asthma, rhinoconjunctivitis and persistent food allergies; long term follow-up of cohort studies is necessary to examine whether perturbations are also related to these outcomes.

In conclusion, we demonstrated that differences in the gut microbiota composition precede the manifestation of atopic symptoms and atopic sensitisation. In particular, *C. difficile* was associated with all atopic symptoms and sensitisation, whereas *E. coli* appeared to be only associated with (non-atopic) eczema. Different immunological mechanisms may underlie the effects of *E. coli* and *C. difficile*. This calls for further research on the mechanisms by which intestinal microbes interfere with our (gastrointestinal) immune system.

ACKNOWLEDGEMENTS
We thank Chantal Delnoy, Brigitte Winants and Karen Groot for home visits, Cobie Martens and Willeke Hendrikx for assistance with data collection questionnaires, Astrid van Leeuwen (Sanquin) for IgE determination, and last but not least, all mothers and their infants participating in the KOALA study.

Authors’ affiliations
J Penders, P. A van den Brandt, I Kummeling, B Snijders, Department of Epidemiology, Nutrition and Toxicology Research Institute Maastricht (NUTRIM), Maastricht University, Maastricht, the Netherlands
C Thijs, Department of Epidemiology, Care and Public Health Research Institute (Caphri), Maastricht University, Maastricht, the Netherlands
R van Ree, Department of Experimental Immunology, Academic Medical Center, Amsterdam, the Netherlands
F Stelma, H Adams, E E Stobberingh, Department of Medical Microbiology, University Hospital of Maastricht, Maastricht, the Netherlands

Funding: This study was supported by grants from the Dutch Asthma Foundation (grant 3.2.03.48) and Royal Friesland Foods (the Netherlands).

Competing interests: None declared.

REFERENCES
Gut microbiota composition and atopy