ORIGINAL ARTICLE

The oesophageal string test: a novel, minimally invasive method measures mucosal inflammation in eosinophilic oesophagitis

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ABSTRACT

Objective Eosinophil predominant inflammation characterises histological features of eosinophilic oesophagitis (EoE). Endoscopy with biopsy is currently the only method to assess oesophageal mucosal inflammation in EoE. We hypothesised that measurements of luminal eosinophil-derived proteins would correlate with oesophageal mucosal inflammation in children with EoE.

Design The Enterotest diagnostic device was used to develop an oesophageal string test (EST) as a minimally invasive clinical device. EST samples and oesophageal mucosal biopsies were obtained from children undergoing upper endoscopy for clinically defined indications. Eosinophil-derived proteins including eosinophil secondary granule proteins (major basic protein-1, eosinophil-derived neurotoxin, eosinophil cationic protein, eosinophil peroxidase) and Charcot–Leyden crystal protein/galectin-10 were measured by ELISA in luminal effluents eluted from ESTs and extracts of mucosal biopsies.

Results ESTs were performed in 41 children with active EoE (n=14), EoE in remission (n=8), gastro-oesophageal reflux disease (n=4) and controls with normal oesophagus (n=15). EST measurement of eosinophil-derived protein biomarkers significantly distinguished between children with active EoE, treated EoE in remission, gastro-oesophageal reflux disease and normal oesophagus. Levels of luminal eosinophil-derived proteins in EST samples significantly correlated with peak and mean oesophageal eosinophils/high power field (HPF), eosinophil peroxidase indices and levels of the same eosinophil-derived proteins in extracts of oesophageal biopsies.

Conclusions The presence of eosinophil-derived proteins in luminal secretions is reflective of mucosal inflammation in children with EoE. The EST is a novel, minimally invasive device for measuring oesophageal eosinophilic inflammation in children with EoE.

BACKGROUND

Eosinophilic oesophagitis (EoE) is a chronic oesophageal disease characterised by a constellation of symptoms and eosinophil predominant inflammation of the oesophageal epithelium.1 2 A significant problem besetting the understanding of pathogenic mechanisms and the care of patients with EoE is the identification and capture of biomarkers of oesophageal inflammation. Except for enumeration of eosinophils in oesophageal...
biopsies, no validated measurements of oesophageal inflammation or disease activity have been identified. Previous studies suggest that measurements of eosinophils as well as extracellular eosinophil secondary granule protein (ESGP) deposition in the oesophageal mucosa may serve as valuable measures of disease activity in EoE.3–6

The ESGPs include four cationic proteins present in the eosinophil’s large specific (secondary) granule known as major basic protein-1 (MBP1), eosinophil peroxidase (EPX), eosinophil-derived neurotoxin (EDN) and eosinophil cationic protein (ECP). A fifth protein, the Charcot–Leyden crystal protein/galectin-10 (CLC/Gal-10),7 is primarily a cytosolic protein, but is also present in the eosinophil’s primary (coreless) granule.8 Each granule-associated protein carries different biochemical and functional properties relevant to proposed pathogenetic mechanisms for oesophageal dysfunction and remodelling in EoE.9–12 Thus, measurement of eosinophil-derived proteins, including ESGPs and CLC/Gal-10, within the oesophageal mucosal microenvironment could provide both clinically relevant information regarding EoE disease activity and novel pathophysiological mechanisms.

To date, the only methodology to assess mucosal inflammation in patients with EoE is endoscopy and mucosal biopsy. Correlation of symptoms with eosinophilic inflammation is variable and peripheral measurements of eosinophil-associated biomarkers from serum and stool have been of limited value13–15 in which some demonstrate significant correlation whereas others do not. As an alternative to endoscopy and mucosal biopsy and peripheral measurements, we have used the proximal portion of the Enterotest device16–17 to measure luminal eosinophil-derived proteins and termed this section of the Enterotest the oesophageal string test (EST). We hypothesised that measurements of ESGPs and CLC/Gal10 directly in oesophageal luminal secretions would reflect mucosal inflammation in EoE.

MATERIALS AND METHODS
Enterotest description
The Enterotest (HDC Corporation, Pilpitas, CA, USA), a minimally invasive string-based technology composed of a capsule filled with approximately 90 cm of string, was originally designed to detect gastric and small intestine pathogens, sample bile and assess for gastro-oesophageal reflux disease (GORD) (online supplementary figure S1A).17–19 In this study, one end of the string (~10 cm) was pulled from the capsule and wound around the index finger (online supplementary figure S1B–D). The capsule was swallowed, the proximal string taped to the cheek (online supplementary figure S1E) and the remaining string in the capsule deployed to end in the duodenal lumen, where the capsule was dislodged.

We sampled luminal secretions from the proximal part of the Enterotest string and termed this the EST. Locations of oesophageal and gastric segments of the Enterotest were determined using pH indicator sticks (online supplementary figure S1F) supplied with tests, and if subjects were on antiacid medications, by using pH indicator sticks (online supplementary figure S1F). Subjects could drink liquids until 3 h before their endoscopic procedure. If families or subjects desired, the subject was admitted to the Research Center (Clinical and Translational Research Center) at the Children’s Hospital Colorado or Ann and Robert H Lurie Children’s Hospital of Chicago the night before the procedure.

The day after swallowing the Enterotests, subjects had upper endoscopy performed. After anaesthesia was administered, the Enterotest was removed, adherent secretions were eluted from the proximal oesophageal section of the Enterotest and the sample was immediately frozen for later batch analysis of the ESGPs and CLC/Gal-10. The oesophageal string location was determined as noted below (online supplementary figure S1F). Mucosal pinch biopsies were then taken from oesophageal mucosal surfaces and immediately snap frozen for eosinophil-derived protein extraction and measurement. Diagnostic biopsies were placed in neutral buffered formalin for processing and routine H&E (eosinophil enumeration) and immunohistochemical (EPX Staining Index) staining.

This study was approved by the Colorado Multi-institutional IRB (COMIRB) and IRBs of the University of Illinois at Chicago and the Children’s Memorial Hospital.

Processing EST samples and biopsies for ESGPs and CLC/-10 biomarker assays
See online supplementary methods.

Quantitative measurement of ESGP and CLC/Gal-10 biomarkers
ESGP and CLC/Gal-10 levels present in EST samples and biopsy extracts were determined using commercial (EDN, ECP; MBL International, Woburn, MA, USA) or inhouse developed (MBP1, CLC/Gal-10, EPX) ELISAs. Assays for EDN and ECP were performed according to the manufacturer’s instructions. Double-antibody sandwich ELISAs for MBP1 and CLC/Gal-10 were developed and standardised at the Ackerman laboratory.
(Chicago, IL, USA) and for EPX at the Lee laboratory (Scottsdale, AZ, USA). MBP1 and CLC/Gal-10 ELISAs used standard curves generated using the purified eosinophil-derived protein, prepared as previously described.7 20–22 For MBP1, mouse monoclonal antibodies were used for both capture and detection (mouse anti-human MBP1-clones, J14-S2A and J13-6B6, respectively, kindly provided by Dr Hirohito Kita, Mayo Clinic, Rochester, Minnesota, USA), the detection antibody being biotinylated and detected using streptavidin-conjugated horseradish peroxidase (ExtaAvidin Peroxidase, E2886; Sigma-Aldrich, St. Louis, Missouri, USA). In the detection antibody being biotinylated and detected using streptavidin-conjugated horseradish peroxidase (ExtaAvidin Peroxidase, E2886; Sigma-Aldrich, St. Louis, Missouri, USA), the detection antibody was a CLC/Gal-10 affinity purified rabbit polyclonal antibody prepared as previously described.7 Detection antibodies for the MBP1 and CLC/Gal-10 ELISAs were biotinylated using a Biotin-XX Microscale Protein Labelling Kit (B30010; Molecular Probes, Invitrogen, Eugene, Oregon, USA) according to the manufacturer’s instructions. MBP1 and CLC/Gal-10 ELISA assays detected these eosinophil-derived proteins in the ranges of 11.8–750 and 0.125–16 ng/ml, respectively, with recoveries of 80%–100% and 100%–120%, signal to noise ratios >5 and coefficient of variation (CV) of 15%.

Specific parameters and detailed protocol and methods associated with the EPX-based ELISA in mice have been recently described.23 EPX ELISA detected EPX in the range of 8–1024 ng/ml, with recovery of 90%, signal to noise ratios >15 and an assay CV of 13%.

**Histological analysis**

Eosinophils were enumerated in formalin-fixed, H&E stained tissue sections as previously described (KC, SS, JM, Denver; HM, Chicago).24 Eosinophils were defined as having at least one nuclear lobe identified in association with intracellular eosin stained granules. Only epithelial eosinophils were enumerated (surface area 0.26 mm²). Results were reported as peak (in one mucosal sample from most densely populated section) and mean eosinophils/HPF (from 5 HPFs counted). Analytical variability of histological assessment was <5% and discrepancies between investigators were resolved.

**EPX immunohistochemistry and EPX scoring**

Immunohistochemistry for EPX and determination of the EPX Staining Index score for proximal and distal oesophageal biopsies was performed in formalin-fixed, paraffin-embedded tissue sections as previously described.6

**Statistical analysis**

SAS V1 9.2 was used for statistical analysis. Biomarker levels were compared across groups using ANOVA. Since these outcome variables were not normally distributed, logarithmic transformation to the natural base was applied to the data, where the value of 0 in biomarker level was coded as 10⁻⁶. Spearman correlation coefficient (r) was used to assess the correlations between biomarkers. Fisher’s z transformation was applied when testing statistical significance of r. C-statistics (ie, the area under receiver operating characteristic (ROC) curve) were used to quantify the discriminating ability of a biomarker. C value>0.7 was considered acceptable discrimination. ROC model was estimated from the logistic regression model with the level of biomarkers being a continuous predictor.

**RESULTS**

**Eosinophil-derived proteins bind to Enterotest nylon strings in vitro**

As determined by western blot analyses, Enterotest strings detected a concentration dependent increase in MBP1 protein from eosinophil lysates (online supplementary figure S2); EPX and MBP1 were detected on Enterotest nylon strings as early as 1 h after incubation with lysates (online supplementary figure S3). Strings were also able to detect EPX and MBP1 from IL-5 activated intact eosinophils (online supplementary figure S4) as early as 1 h following incubations. Strings were also able to capture another eosinophil-derived protein, EDN, from activated eosinophils (1×10⁶) (online supplementary figure S5). Together, these in vitro results supported the feasibility of using Enterotest nylon strings in situ to capture eosinophil-derived proteins.

**Subject characterisation and tolerance of procedure**

Forty-one subjects were recruited; no significant differences in recruitment were identified between the two sites. Clinical characteristics of these subjects are summarised in online supplementary table S1. No complications or endoscopic mucosal findings were identified that were related to the performance of Enterotests and a gagging sensation during swallowing the capsule was the only problem reported.

**Mucosal biopsy concentrations of ESGPs and CLC/Gal-10 differentiate subjects with active EoE from those with treated EoE, GORD and normal subjects without inflammation**

Eosinophil enumeration has been the ‘gold standard’ biomarker used for assessing mucosal inflammation. Thus, we first compared the number of mucosal eosinophils (enumeration and EPX Staining Index) with quantitative measurements of eosinophil-derived proteins in biopsy extracts. Consistent with previous studies, both eosinophil numbers and EPX Staining Indices were significantly greater in subjects with active EoE compared with healthy controls (table 1). Eosinophil-derived protein concentrations (figure 1; (A1) MBP1, (A2) EDN, (A3) ECP (A4) EPX and (A5) CLC/Gal-10) in oesophageal biopsy extracts were significantly increased in subjects with active EoE compared with treated EoE (MBP1, p<0.0001; EDN, p<0.0001; ECP, p<0.001; EPX, p<0.05; CLC/Gal-10, p<0.01; active vs treated), GORD (MBP1, p<0.001; EDN, p<0.0001; ECP, p<0.001; EPX, p<0.05; active vs GORD) and to those with normal mucosa (MBP1, p<0.0001; EDN, p<0.0001; ECP, p<0.001; EPX, p<0.001; CLC/Gal-10, p<0.0001 active vs normal).

Next we wanted to determine whether mucosal biopsy concentrations of eosinophil-derived proteins correlated with traditional and newer measurements of eosinophil burden. All eosinophil-derived proteins tested correlated significantly with peak eosinophil counts, mean eosinophil counts and EPX Staining Indices measured in the above subject groups (p<0.0001 for all eosinophil proteins vs peak eosinophil, mean eosinophil and EPX Staining Index) (table 2A).

Together, these results determined that mucosal biopsy concentrations of eosinophil-derived proteins, similar to eosinophil enumeration, serve as reliable measures of mucosal eosinophil burden. In addition, measurements of biopsy extracts were significantly different between disease states suggesting they were reflective of disease activity.
EST detection of ESGPs and CLC/Gal-10 differentiates subjects with active EoE from those with treated EoE, GORD and normal subjects without inflammation

To determine whether we could capture evidence of mucosal inflammation by using an intraluminal device, we next measured levels of luminal oesophageal eosinophil-derived proteins with ESTs in subjects with active EoE, treated EoE, GORD and normal oesophagus. Eosinophil-derived proteins (figure 1; (B1) MBP1, (B2) EDN, (B3) ECR (B4) EPX and (B5) CLC/Gal-10) were significantly increased in EST samples obtained from subjects with active EoE compared with those with treated EoE (MBP1, p<0.001; EPX, p<0.01; CLC/Gal-10, p<0.0001; active vs treated), GORD (MBP1, p<0.0001; EDN, p<0.0001; CLC/Gal-10, p<0.01; active vs GORD) as well as subjects with no histopathological evidence of oesophageal inflammation (MBP1, p<0.0001; EDN, p<0.0001; ECR p<0.01; CLC/Gal-10 p<0.0001; active vs normal).

Luminal measurements of eosinophil-derived proteins correlate with tissue measurements of mucosal eosinophilic inflammation

To determine whether luminal concentrations of eosinophil-derived proteins were reflective of tissue measurements of eosinophil burdens, we performed correlational analyses of eosinophil-derived protein concentrations obtained with ESTs with three measures of eosinophilic inflammation present in the tissue: (1) traditional gold standard measurement1 2—eosinophil counts/HPF (peak and mean), (2) newer measurement6—EPX Staining Index and (3) newest measurement (this study)—eosinophil-derived protein levels in mucosal biopsy extracts determined by ELISA. Concentrations of MBP1, EDN, EPX and CLC/Gal-10 measured in EST samples obtained from all subjects correlated significantly with eosinophil counts (figure 2; A, peak eosinophils; B, mean eosinophils) and EPX indices (table 2B). In addition, EST assessments of all eosinophil-derived proteins correlated significantly with measures of the same biomarkers measured in extracts of matched mucosal biopsies (MBP1, p<0.001; EDN, p<0.0001; ECP, p<0.05 and CLC/Gal-10, p<0.0001) (figure 3). Together, these results demonstrated that capture of luminal eosinophil-derived biomarkers by ESTs provides an accurate reflection of oesophageal mucosal eosinophilic inflammation.

ESTs can detect eosinophilic inflammation without evidence of superficial inflammation

We next determined whether evidence of superficial mucosal inflammation was necessary to detect luminal inflammation with the EST. Of the 14 active EoE subjects, white exudates, microabscess or superficial layering were seen in all but four subjects despite each having detectable eosinophil-derived protein levels in either their biopsy or EST samples. Except for one condition, ECP versus microabscess (table 3, values in bold), no significant correlations were identified between gross or histological measurements of oesophageal mucosal inflammation with respect to

### Table 1 Summary of histological and gross features of mucosal eosinophilia

<table>
<thead>
<tr>
<th></th>
<th>EoE-active (untreated), n=14</th>
<th>EoE-remission (treated), n=8</th>
<th>GORD, n=4</th>
<th>Normal oesophagus, n=15</th>
<th>p Values untreated EoE versus treated, GORD, normal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Eosinophil peak count (±SEM)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Proximal</td>
<td>30.1±6.0</td>
<td>0.4±0.3</td>
<td>0.3±0.2</td>
<td>0</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td>Distal</td>
<td>45.4±9.9</td>
<td>0</td>
<td>0.8±0.4</td>
<td>0</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td>Eosinophil average of five random fields (±SEM)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Proximal</td>
<td>17.4±3.9</td>
<td>0.1±0.1</td>
<td>0.3±0.2</td>
<td>0</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td>Distal</td>
<td>28.9±6.0</td>
<td>0</td>
<td>0.3±0.2</td>
<td>0</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td>EPX Staining Index*</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Proximal</td>
<td>34.2±5.3</td>
<td>4.4±2.3</td>
<td>0</td>
<td>2.8±2.8</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td>Distal</td>
<td>41.1±1.9</td>
<td>6.5±3.5</td>
<td>11.3±6.0</td>
<td>5.9±3.3</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td>Endoscopic evidence of exudate (% of total patients)</td>
<td>6 (43%)</td>
<td>2 (25%)</td>
<td>1 (25%)</td>
<td>0</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td>Microabscess† (% of total patients)</td>
<td>11 (79%)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>p&lt;0.001</td>
</tr>
<tr>
<td>Superficial layering‡ (% of total patients)</td>
<td>9 (64%)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>p&lt;0.001</td>
</tr>
</tbody>
</table>

*Panels show representative images of immunohistochemical staining for EPX (red reaction product) in tissue sections from subjects with normal oesophagus, untreated EoE, treated EoE in remission and GORD that were used to generate the EPX Staining Index in this study.

†Microabscess: four or more eosinophils adjacent to each other in the superficial epithelia.

‡Superficial layering: eosinophils within the luminal surface of the oesophageal epithelium.

EoE, eosinophilic oesophagitis; EPX, eosinophil peroxidase; GORD, gastro-oesophageal reflux disease.
Eosinophil-derived protein concentrations measured in mucosal biopsies.

No significant correlations were identified between gross measurements (white exudate) of oesophageal mucosal inflammation with respect to eosinophil-derived protein concentrations measured from biopsy or EST samples (table 3). Except for two conditions, MBP1 versus superficial layering and microabscess (table 3, values in bold), no significant correlations were detected when comparing histological evidence of superficial epithelial inflammation (eosinophil microabscess and superficial layering) with eosinophil-derived protein concentrations measured from ESTs. These results suggest that ESTs can capture eosinophilic inflammation even if there is no evidence of superficial mucosal inflammation.

EST-based assessment of eosinophil-derived proteins represents a sensitive and specific test for oesophageal eosinophil inflammation

We next sought to determine whether assessments of subjects using mucosal biopsies and EST-based measurements of eosinophil specific proteins achieved diagnostic/therapeutic thresholds that would permit their use to diagnose and monitor children with EoE. ROC curves for eosinophil-derived protein concentrations of MBP1 and CLC measured in mucosal biopsies and ESTs were used to establish diagnostic thresholds.
ESTs indicated a high predictive power with $C=1.00$ and $C=0.99$ for mucosal biopsy sampling, and $C=0.97$ and $C=0.97$ for EST sampling, respectively (figure 4A). Significant predictive values were also true for each of the other eosinophil-derived proteins (figure 4B and online supplementary figure S6).

**DISCUSSION**

EoE is a chronic disease characterised histologically by epithelial eosinophilia. To date, oesophageal eosinophilia can only be fully assessed by procuring mucosal biopsies during endoscopic procedures. While endoscopy with biopsy is a safe, efficacious and commonly performed procedure, it is also costly, time consuming and carries potential complications. In an effort to determine a less expensive, quicker and safer alternative to endoscopic biopsy, we wondered whether mucosal eosinophilia associated with EoE could be assessed in a less invasive manner using an FDA-approved device, the Enterotest. Other measures of oesophageal function such as ultrasound, endoflip, stationary and mobile motility monitoring and pH/impedance monitoring are invaluable to document anatomy and real-time oesophageal function, but they cannot analyse biochemical features of oesophageal inflammation. Therefore, we first tested the ability of the Enterotest to capture eosinophil proteins from the oesophageal lumen. Consistent with previous studies that used the Enterotest to geographically target different sections of the gastrointestinal tract, we used the proximal end of Enterotest to directly measure oesophageal inflammation in children with EoE. Our results demonstrate that oesophageal portions of the Enterotests or the EST detected luminal concentrations of eosinophil-derived proteins which correlated significantly with the gold standard eosinophil enumeration (peak and mean), the recently defined methodology, EPX Staining Index and the herein described measurements of eosinophil-derived proteins in biopsy extracts. The high degree of correlation of eosinophil-derived proteins from EST samples with these metrics provides very strong support for the use of the EST to biochemically interrogate the oesophageal mucosa.

Our findings document the use of the EST to capture eosinophil-derived proteins and its potential use as a device to assist in monitoring therapeutic efficacy in patients with EoE. In this regard, therapeutic efficacy could be monitored on a more regular basis following changes in treatments or during exacerbations of disease. However, the small number of GORD patients studied to date, and the lack of commercially available standardised ELISAS for a number of eosinophil biomarker, currently obviate use of the EST as a diagnostic device for EoE pending further studies.

Table 2 Correlations of eosinophil-derived protein concentrations from biopsy extracts and ESTs with histological measures of mucosal eosinophilia

<table>
<thead>
<tr>
<th>A. Biopsies</th>
<th>Peak eosinophils*</th>
<th>Mean eosinophils*</th>
<th>EPX Staining Index*</th>
</tr>
</thead>
<tbody>
<tr>
<td>MBP1</td>
<td>0.72559</td>
<td>&lt;0.0001</td>
<td>0.72003</td>
</tr>
<tr>
<td>EDN</td>
<td>0.78673</td>
<td>&lt;0.0001</td>
<td>0.79306</td>
</tr>
<tr>
<td>ECP</td>
<td>0.70621</td>
<td>&lt;0.0001</td>
<td>0.70804</td>
</tr>
<tr>
<td>EPX</td>
<td>0.61117</td>
<td>&lt;0.0001</td>
<td>0.61596</td>
</tr>
<tr>
<td>CLC/Gal-10</td>
<td>0.66360</td>
<td>&lt;0.0001</td>
<td>0.66329</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>B. ESTs</th>
<th>r Value†</th>
<th>p Value</th>
<th>r Value†</th>
</tr>
</thead>
<tbody>
<tr>
<td>MBP1</td>
<td>0.72170</td>
<td>&lt;0.0001</td>
<td>0.72003</td>
</tr>
<tr>
<td>EDN</td>
<td>0.47688</td>
<td>&lt;0.002</td>
<td>0.47688</td>
</tr>
<tr>
<td>ECP</td>
<td>0.20519</td>
<td>NS</td>
<td>0.20519</td>
</tr>
<tr>
<td>EPX</td>
<td>0.54145</td>
<td>&lt;0.0002</td>
<td>0.54145</td>
</tr>
<tr>
<td>CLC/Gal-10</td>
<td>0.72735</td>
<td>&lt;0.0001</td>
<td>0.72735</td>
</tr>
</tbody>
</table>

*n=41 subjects for peak and mean eosinophil numbers and EPX index. †Spearman’s rank correlation coefficient (r) for values from n=41 subjects.
such as asthma and allergic conjunctivitis revealed that eosinophils and their products are deposited generously within affected mucosal surfaces and these can be captured in fluids obtained from the lung, tears and now the oesophagus.\textsuperscript{31–36}

Eosinophilic inflammation of the oesophageal mucosa may or may not contain evidence of superficial eosinophilia as characterised by endoscopic findings of white exudates, histological patterns of microabscess formation and/or superficial layering.\textsuperscript{5, 6, 37, 38} Our results are highly suggestive that eosinophil-derived proteins can be captured regardless of whether gross or histological evidence of surface eosinophilia is present, but further studies will need to be completed to validate this finding.

Eosinophil-derived proteins are biologically potent molecules that carry significant functional activities relevant to the mechanisms of pathogenesis in EoE. In animal models, MBP...
has been shown to contribute to increased epithelial permeability, smooth muscle contraction and liberation of molecules related to tissue remodelling and fibrosis. These functional properties bear particular relevance to patients with EoE.

Pathogenetic mechanisms related to EoE include alterations in barrier function with dysregulated filaggrin expression, altered oesophageal motility and oesophageal stricture formation/long segment narrowing. Thus, ongoing identification and measurement of eosinophil-derived proteins may provide additional insights into pathogenetic mechanistic pathways.

In this study, ESTs captured eosinophil-derived proteins successfully, but a few technical and biological issues remain to be addressed. Whereas this study focused on measurements of eosinophil-derived proteins as a biomarker of EoE, future studies will need to identify novel intraluminal biomarkers that will allow for the differentiation of EoE from other forms of oesophagitis. Identification and validation of these biomarkers will then offer increased diagnostic capacity of the device, especially in the assessment of patients with dysphagia and other symptoms referable to the oesophagus. To maximise the ability of ESTs to harvest measureable levels of eosinophil-derived proteins from oesophageal secretions, this study was performed leaving ESTs in the oesophagus overnight. The eventual use of ESTs will be best suited to an office setting, and ongoing studies are testing shorter incubation times. Contamination from swallowed oropharyngeal or refluxed gastric secretions may alter oesophageal measurements, but our data (not shown) related to gastric contamination suggest low inconsequential eosinophil-derived protein levels in the stomach. Minimally detectable levels of eosinophil-derived proteins in normal patients could have been from undetected oesophageal inflammation in swallowed nasal/oral secretions; these levels did not reach those detected in active EoE patients. The primary sources of proteins measured here are eosinophils, but basophils, mast cells, neutrophils and T lymphocytes also express or sequester low levels of some of these proteins. These sources may also help to explain the non-zero values of ESGPs in control and treated subjects.
In summary, we have demonstrated the use of a minimally invasive test, the EST, to document mucosal inflammation in children with EoE. Although we do not suggest that the EST will replace endoscopy and biopsy as a critical tool for analyses of EoE, it certainly has the potential to significantly improve the evaluation and treatment of patients with EoE who may require repeated assessments of their oesophageal mucosae. In addition, as future biomarkers are identified and validated for EoE, ESTs may be able to differentiate patient phenotypes that are more predisposed to complications or responsive to various treatments.

Acknowledgements  We thank the physicians (Robert Kramer, Edward Hoffenberg, Edwin Liu, Edwin de Zoeten, David Brumbaugh, Christine Waasdorp, Shikha Sundaram, Cara Mack, Michael Narkewicz, Ronald Sokol, Jason Soden, Deborah Neigut, Barry K Wershil, Lee Bass, Jennifer Strople) and nurses (Tammy

Figure 4  Receiver operating characteristic (ROC) curves for the diagnosis of eosinophilic oesophagitis with biopsies and oesophageal string test (EST) sampling of eosinophil-derived proteins. ROC sensitivity versus specificity curves (A) are shown for biopsies (left panels) and ESTs (right panels) for measurements of major basic protein-1 (MBP1) and Charcot–Leyden crystal protein/galectin-10 (CLC/Gal-10) in biopsy extracts and EST supernatants. C-statistics (the area under the ROC curve) are indicated as a measure of the discriminating ability of each biomarker. (B) C-values for the ROC curves for biopsies and EST samples for other eosinophil secondary granule proteins are presented here and graphically in online supplementary figure 6. ECP, eosinophil cationic protein; EDN, eosinophil-derived neurotoxin; EPX, eosinophil peroxidase.

Table

<table>
<thead>
<tr>
<th>Eosinophil Protein</th>
<th>ROC for Biopsies (C-values)</th>
<th>ROC for EST (C-values)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MBP1</td>
<td>0.97</td>
<td>0.97</td>
</tr>
<tr>
<td>CLC/Gal-10</td>
<td>0.99</td>
<td>0.97</td>
</tr>
<tr>
<td>EDN</td>
<td>0.99</td>
<td>0.88</td>
</tr>
<tr>
<td>EPX</td>
<td>0.88</td>
<td>0.82</td>
</tr>
<tr>
<td>ECP</td>
<td>0.98</td>
<td>0.73</td>
</tr>
</tbody>
</table>

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