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ORIGINAL ARTICLE

XIAP variants in male Crohn's disease

Yvonne Zeissig,^{1,2,3} Britt-Sabina Petersen,³ Snezana Milutinovic,⁴ Esther Bosse,¹ Gabriele Mayr,³ Kenneth Peuker,¹ Jelka Hartwig,¹ Andreas Keller,⁵ Martina Kohl,² Martin W Laass,⁶ Susanne Billmann-Born,³ Heide Brandau,⁷ Alfred C Feller,⁸ Christoph Röcken,⁹ Martin Schrappe,² Philip Rosenstiel,³ John C Reed,⁴ Stefan Schreiber,^{1,3} Andre Franke,³ Sebastian Zeissig¹

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For numbered affiliations see end of article.

Correspondence to

Dr Sebastian Zeissig, Department of Internal Medicine I, University Medical Center Schleswig-Holstein, Kiel, Schittenhelmstr. 12, Kiel 24105, Germany; szeissig@1med.uni-kiel.de and Dr Andre Franke, Institute of Clinical Molecular Biology, Christian-Albrechts-University of Kiel, Schittenhelmstr. 12, 24105 Kiel, Germany; a.franke@mucosa.de

YZ and BSP contributed equally to this work. SZ and AF contributed equally to this work.

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ABSTRACT

Objective The genetic basis of inflammatory bowel disease (IBD) is incompletely understood. The aim of this study was to identify rare genetic variants involved in the pathogenesis of IBD.

Design Exome sequencing and immunological profiling were performed in a patient with early onset Crohn's disease (CD). The coding region of the gene encoding X-linked inhibitor of apoptosis protein (*XIAP*) was sequenced in samples of 275 paediatric IBD and 1047 adult-onset CD patients. *XIAP* genotyping was performed in samples of 2680 IBD patients and 2864 healthy controls. Functional effects of the variants identified were investigated in primary cells and cultured cell lines.

Results Our results demonstrate the frequent occurrence of private variants in *XIAP* in about four percent of male patients with paediatric-onset CD. While *XIAP* mutations are known to be associated with the primary immunodeficiency (PID) X-linked lymphoproliferative disease type 2 (XLP2), CD patients described here exhibited intestinal inflammation in the absence of XLP2 and harboured a spectrum of mutations partially distinct from that observed in XLP2. The majority of *XIAP* variants identified was associated with a selective defect in NOD1/2 signalling, impaired NOD1/2-mediated activation of NF- κ B, and altered NF- κ B-dependent cytokine production.

Conclusions This study reveals the unanticipated, frequent occurrence of *XIAP* variants in male paediatric-onset CD. The link between *XIAP* and NOD1/2, and the association of *XIAP* variants with XLP2, support the concept of PID in a subset of IBD patients. Moreover, these studies provide a rationale for the implementation of *XIAP* sequencing in clinical diagnostics in male patients with severe CD.

INTRODUCTION

Inflammatory bowel disease (IBD) is a heterogeneous group of chronic inflammatory disorders of the intestine with Crohn's disease (CD) and ulcerative colitis (UC) as the two major forms.¹ Although the precise aetiology of IBD is unknown, susceptibility to disease is influenced by a complex interplay of genetic and environmental factors.²⁻⁴ Previously, we and others identified over 163 common genetic variants associated with IBD that are involved in the regulation of intestinal barrier function as well as innate and adaptive immunity.⁵ However, for the majority of genetic variants associated with IBD, a

Significance of this study

What is already known on this subject?

- Immune defects have been described in patients with CD and may contribute to the pathogenesis of intestinal inflammation.
- *XIAP* mutations are associated with X-linked lymphoproliferative disease type 2 (XLP2), a primary immunodeficiency, which can manifest as intestinal inflammation.
- *XIAP* has been implicated in the regulation of several pathways relevant to IBD including NOD1/2 signalling, apoptosis, and natural killer T cell development.

What are the new findings?

- Private variants in *XIAP* are commonly observed in male patients with paediatric-onset CD and affect about 4% of patients in this subset of CD patients in the German cohort described here.
- Novel *XIAP* variants described here are associated with CD as the sole clinical manifestation in the absence of XLP2.
- Lack of functional *XIAP* in primary immune cells is associated with selective defects in NOD1/2 signalling, while other alterations in innate and adaptive immunity were not observed.

How might it impact on clinical practice in the foreseeable future?

- These studies provide a mechanistic basis for the unexplained observation of functional NOD2 defects in the absence of genetic *NOD2* variants, support the concept of immunodeficiency in a subset of CD patients, and provide a rationale for the implementation of *XIAP* sequencing in routine clinical diagnostics in male patients with severe CD.

complete understanding of the mechanisms that contribute to disease pathogenesis is missing. This is related to the fact that these polymorphisms are common, of low to moderate effect size, and are likely to modify disease risk through interaction with environmental factors and other genetic variants found in the same individual.^{3,6}



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Mendelian disorders can provide crucial insight into the mechanisms of disease pathogenesis. Accordingly, the analysis of gene-function relationships in IBD may be facilitated by the study of rare phenotypes such as early onset CD and UC, which are often associated with a severe course of disease, lack of response to immunomodulators and biologicals, and familial segregation.^{7–12} In accordance with this concept, studies in early onset IBD revealed apparent monogenetic variants of IBD and provided unique insight into disease pathogenesis. Thus, mutations in the genes encoding for IL-10 and its receptor subunits IL-10R1 and IL-10R2 were found to be associated with chronic intestinal inflammation and manifestation of disease during the first year of life.^{9–11 13 14} As *IL10* and *IL10rb* deficiency in mice are associated with spontaneous intestinal inflammation, these data suggest that defects in this pathway are sufficient to induce intestinal inflammation in the presence of the intestinal microbiota.^{15 16} Similarly, mutations in a disintegrin and metalloprotease 17 (*ADAM17*) are associated with early onset IBD in accordance with susceptibility to intestinal inflammation observed in mice with *Adam17* deletion.^{17 18} Further, the study of primary immunodeficiencies (PID), such as chronic granulomatous disease (CGD), Wiskott–Aldrich syndrome, and IPEX has revealed monogenetic immune defects associated with chronic intestinal inflammation.^{8 19 20} These studies demonstrate how individual genetic variants can lead to intestinal inflammation and, together with functional studies in CD patients,^{21 22} lend support to the concept of primary immune defects underlying a subset of IBD cases.^{8 23 24} Along these lines, recent studies revealed that mutations in the gene encoding for the X-linked inhibitor of apoptosis protein (*XIAP*) are associated with the PID XLP2 and also with chronic intestinal inflammation.^{25–30} While these observations were originally made in families with XLP2, a recent exome sequencing study identified a novel *XIAP* variant in a child with early onset CD as the sole clinical manifestation.³¹ These results raised the question of whether *XIAP* variants are commonly associated with IBD. Here, we demonstrate the frequent occurrence of private *XIAP* variants in male patients with paediatric-onset CD, which affects about 4% of patients in this CD subset.

METHODS

Patients

The study was approved by the local ethics committees. All patients and, in case of children, both parents provided written informed consent. Control subjects for functional studies included the patient's mother and healthy volunteers (median age 25 years, range 23–33). Neither the patient nor the controls received medication. Genotyping of *XIAP* variants was performed in 1900 CD patients (median age at disease onset 25 years, range 1–73; 58% male, 42% female), 780 UC patients (median age at disease onset 27, range 3–61; 54% male, 46% female) and 2864 healthy controls (52% male, 48% female). Sanger sequencing of *XIAP* was performed in two cohorts: A cohort of 275 paediatric IBD cases including 181 CD cases (95 male patients, 86 female patients, median age at disease onset 13, range 1–16; 37 cases A1a, 144 cases A1b according to the Paris classification¹²) and 94 UC cases (48 male patients, 46 female patients, median age at disease onset 13 years, range 1–16; 24 cases A1a, 70 cases A1b according to the Paris classification¹²). The second cohort consisted of 1047 patients with adult onset CD (median age at disease onset 29 years, range 17–70, 36% male, 64% female).

Exome sequencing

Samples were enriched using Illumina's TruSeq Exome Enrichment Kit. Sequencing of 2×100 bp paired-end reads was performed for the three samples together on one lane of the Illumina HiSeq2000. Reads were mapped against the human reference genome build hg19 using BWA³² v0.5.9, sorted, converted to bam format and indexed with SAMtools³³ v0.1.8, followed by the removal of PCR duplicates with Picard v1.55 (<http://picard.sourceforge.net>). Local realignment around InDels and base quality score recalibration as well as variant calling and quality score recalibration were performed with GATK³⁴ v1.2–60 according to their best practice recommendations. For annotation we applied ANNOVAR's³⁵ summarize_annovar.pl script. For the identification of de novo variants we used a two-step approach involving the somatic mutation command from VarScan³⁶ v2.2.11 for a pairwise comparison of the son and the mother as well as the son and the father. We calculated the overlap of detected differences from both comparisons to find variants present exclusively in the son followed by manual inspection of the alignment at positions of potential de novo mutations.

Sanger sequencing

PCR and Sanger sequencing were performed on an Applied Biosystems 3730xl. Sequences were analysed using novoSNP.³⁷ Potential novel coding variants were validated by resequencing in the forward and reverse directions. The average overall drop-out rate was below 2.3%. PCR primers and conditions are shown in online supplementary table S1.

Protein structure analysis

See online supplementary methods.

Flow cytometry

Flow cytometry was performed as described previously³⁸ and as described in online supplementary methods.

T cell proliferation and apoptosis

For restimulation-induced cell death, peripheral blood mononuclear cells (PBMCs) were stimulated with PHA (2 µg/mL). After 24 h, recombinant hIL-2 (100 U/mL, R&D systems) was added and renewed every 48 h. On day 14, cells were restimulated with 1 µg/mL plate-bound anti-CD3 antibody (OKT3, Biolegend). Apoptosis was determined using the FITC Annexin V kit (BD Biosciences).

T cell proliferation was determined by ³H-thymidine incorporation 5 days after stimulation (PHA or anti-CD3) or restimulation (PHA, followed by plate-bound anti-CD3 after 14 days) as described previously.³⁸ 5-bromo-2'-deoxyuridine (BrdU) incorporation and cell cycle analysis were determined using the FITC BrdU Flow Kit (BD Biosciences).

TLR and NOD signalling

1×10⁵ PBMCs were stimulated in 96-well flat-bottom plates with LPS (Sigma–Aldrich), MDP (InvivoGen), Tri-DAP (Invivogen), Pam3CSK4 (InvivoGen), TNFα (Peprotech), PMA (Sigma–Aldrich), and ionomycin (Sigma–Aldrich). Cytokine secretion was detected by ELISA (R&D) 24 h after stimulation. Where indicated, Z-VAD-fmk or Z-YVAD-fmk (Enzo Life Sciences) was added.

For luciferase assays, an NF-κB-dependent firefly luciferase plasmid (Promega) was transfected (Lipofectamine 2000) along with wildtype or mutant *XIAP* and a Renilla luciferase plasmid

(phRluc, Promega) for normalisation. After 24 h, cells were stimulated as indicated, and luciferase expression was analysed after another 16 h (Dual-Glo-Luciferase Assay System, Promega).

Electrophoretic mobility shift assays were performed as described previously³⁹ using a probe, which contains an NF-κB consensus site (5'-AGT TGA GGG GAC TTT CCC AGG C-3').

Western blotting and immunoprecipitation

Immunoprecipitation and western blotting was performed as described previously,⁴⁰ and as described in online supplementary methods.

Monocyte-derived dendritic cells (DCs) and lentiviral reconstitution

Generation of monocyte-derived DCs and lentiviral infection were performed as described previously³⁸ and as outlined in online supplementary methods.

Statistical analysis

Statistical testing was performed using the unpaired Student t test. p Values were calculated in a two-tailed manner. For multiple comparisons against the same control, one-way analysis of variance (ANOVA) with Dunnett's posthoc test was applied.

RESULTS

A novel nonsense mutation in XIAP revealed by exome sequencing

A male patient presented with bloody diarrhoea at the age of 9 months. Endoscopy and histological examination revealed severe, ulcerative, discontinuous pancolitis with anal fissures and stenosis. Low-grade, chronic, *Helicobacter pylori*-negative gastritis and duodenitis in accordance with upper gastrointestinal CD was demonstrated. The patient was classified as A1a, L2+L4a, B2p according to the Paris classification¹² and the subsequent course of disease was refractory to mesalazine, corticosteroids, antibiotics and infliximab. A loop ileostomy at 36 months of age was associated with clinical improvement despite persistent mild, discontinuous colitis. For further clinical characterisation see the Supplementary Results section, online supplementary tables S2 and S3, and.³⁰ The patient and both parents underwent exome sequencing. The average sequencing coverage ranged between 45-fold and 80-fold with over 90% of the target covered by more than eightfold in all samples (online supplementary table S4). We identified a total of 53 320 on-target variants in the patient with 22 452 coding variants of which 1212 non-synonymous variants (missense, nonsense, cancel-start, read-through, splice-site and frameshift) were not present in dbSNP132. We filtered using a maximum frequency of 1% in the 1000 genomes data and inspected the remaining 995 variants assuming a recessive model with homozygosity or compound heterozygosity in the son and heterozygosity in both parents. This approach yielded one homozygous (*FAM151A*) and two compound-heterozygous (*RAPGEF4*, *DCHS2*) variants (online

supplementary table S5). Neither *RAPGEF4* nor *DCHS2* are expressed in the intestine. Expression of *DCHS2*, encoding dachshous cadherin-related 2, is restricted to brain and testis,⁴¹ while expression of *RAPGEF4*, encoding Rap guanine nucleotide exchange factor 4, is restricted to the central nervous system, adrenal gland and pancreas.^{42 43} Additionally, neither the function nor the distribution of *FAM151A* is known. Given that these variants did not provide sufficient mechanistic explanation for the observed clinical phenotype, we investigated the possibility of a de novo mutation in the affected son. This resulted in the identification of a novel, hemizygous, de novo nonsense mutation in *XIAP* (c.295G>T, E99X (p.Glu99*), table 1, figure 1A), while other de novo mutations were not detectable. The affected residue is located in the second of seven exons and truncates the protein after the first of three baculovirus inhibitor of apoptosis protein repeat (BIR) domains (figure 1A). This is associated with loss of function specific to BIR domains 2 and 3 and the RING domain, but predicted to retain the ability of the BIR1 domain to interact with TAB1 (figure 1B).

Unaltered PBMC subsets and T cell function in the absence of functional XIAP

To investigate the mechanistic basis underlying intestinal inflammation in the presence of XIAP variants, we studied the phenotype and function of immune cells in the patient harbouring XIAP E99X. Full-length XIAP was undetectable in various PBMC subsets (figure 2A) and lymphoblastic B cell lines (figure 2B). In accordance with a de novo mutation, PBMCs of the patient's mother exhibited unaltered XIAP expression (figure 2A). Phenotypic PBMC analyses revealed unaltered absolute and relative numbers of monocytes, T cells and B cells in the patient carrying XIAP E99X (figure 2C–D, see online supplementary table S3, and data not shown). CD4 and CD8⁺ T cells in the affected child contained a higher proportion of naive subsets compared with healthy adults (figure 2E, see online supplementary figure S1), but the distribution of naive and memory T cells reflected that previously found in healthy children.⁴⁴ Similarly, reduced numbers of memory but not total B cells and unaltered numbers of peripheral blood plasma cells and immunoglobulin levels were found (figure 2C, F, see online supplementary table S3). While deficiency in invariant natural killer T (iNKT) cells was described in patients with XIAP variants,²⁵ the patient carrying XIAP E99X exhibited detectable levels of iNKT cells at the lower margin of controls (figure 2G and reference³⁸).

XIAP BIR domains inhibit caspase-3 (BIR2), -7 (BIR2), and -9 (BIR3).²⁸ Accordingly, XIAP deficiency was reported to be associated with increased susceptibility to reactivation-induced T cell death (RICD).^{25 45} However, despite a mutation causing deletion of the BIR2 and BIR3 domains of XIAP, CD4⁺ and CD8⁺ T cells from the XIAP E99X patient did not exhibit alterations in activation-induced or restimulation-induced T cell death (figure 3A and data not shown). Further, T cell proliferation (figure 3B), expansion (figure 3C), and cell cycle

Table 1 Summary of identified XIAP variants

	Nucleotide sequence	Exon location	Protein sequence	Gender	Zygoty	IBD type	Age at disease onset (years)
Patient 1	c.295G>T	2	E99X	Male	Hemizygous	CD	0.75
Patient 2	c.115G>T	2	G39C	Male	Hemizygous	CD	13
Patient 3	c.890A>C	3	K297T	Male	Hemizygous	CD	15
Patient 4	c.968G>A	3	W323X	Male	Hemizygous	CD	16

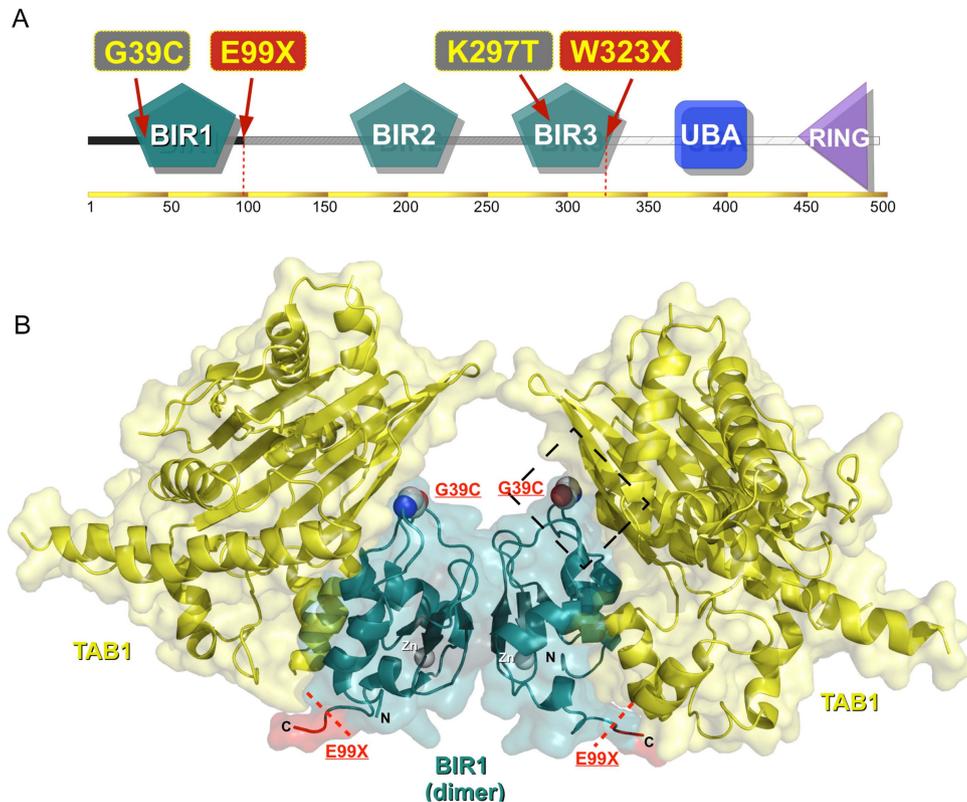


Figure 1 Summary of XIAP variants. A: Domain architecture of the human XIAP protein. Missense mutations are indicated in grey, and nonsense mutations in red boxes. E99X truncates the XIAP protein immediately after the BIR1 domain (residues 26–93), while W323X truncates XIAP near the C-terminus of BIR3 (residues 265–330). B: BIR1/TAB1 complex with XIAP E99X and G39C. Cartoon and surface representation of the XIAP BIR1 protein domain (green) in complex with TAB1 (yellow). The location of E99X is indicated by a red line, and the deleted C-terminal segment is coloured in red. The truncation likely retains the BIR1 domain unaffected with respect to domain interaction and fold stability. G39C is represented as spheres in atomic colours.

progression (figure 3D) induced by phytohemagglutinin (PHA) and anti-CD3 treatment were unaltered (figure 3B–D and data not shown).

XIAP E99X is associated with a severe and selective defect in NOD signalling

XIAP was shown to be critical for NF- κ B-dependent production of IL-6 and IL-8 in response to NOD2 signalling.^{31 40 46–49} In accordance with these findings, PBMCs from the patient carrying XIAP E99X did not exhibit IL-6 or IL-8 secretion in response to the NOD2 ligand muramyl dipeptide (MDP) (figure 4A and see online supplementary figure S2A). Exome sequencing results demonstrated that impaired NOD2 signalling was not due to coding variants in *NOD2*, *BIRC2*, *BIRC3*, *FRMPD2*, *RBCK1*, *RIPK2*, *RNF31*, *SHARPIN* and *TRAF2*, all of which are associated with NOD2 and contribute to NOD2 signalling (data not shown).^{40 47–49} LPS-induced, Pam3CSK4-induced and TNF α -induced IL-6 and IL-8 secretion were unimpaired, thus confirming the absence of general defects in toll-like receptor (TLR) signalling or NF- κ B-mediated IL-6 and IL-8 secretion (figure 4B–D, see online supplementary figure S2B, and data not shown). Additionally, while XIAP has been suggested to act as a negative regulator of the NLRP3 inflammasome,⁵⁰ caspase-1 cleavage and caspase-1-mediated IL-1 β secretion were unaltered in the patient carrying XIAP E99X (see online supplementary figure S3A–B).

XIAP binds receptor-interacting serine-threonine kinase 2 (RIPK2), a downstream mediator of NOD2, in a manner dependent on its BIR2 domain.^{40 49} Since XIAP E99X lacks the

BIR2 domain, we investigated whether the truncated protein fails to interact with RIPK2. While wildtype and E99X myc-XIAP were expressed at similar levels, RIPK2 co-immunoprecipitated only with wildtype but not E99X XIAP in 293 T cells (figure 4E). XIAP E99X thus fails to bind to RIPK2, which provides a mechanistic explanation for the observed NOD2 defect.

To confirm that impaired NOD2 signalling is the consequence of altered XIAP function, XIAP expression was restored by lentiviral reconstitution in primary monocyte-derived dendritic cells (moDCs) of the XIAP E99X patient. Viral transduction rates as determined by lentivirus-derived green fluorescent protein (GFP) were similar in all patients with about 70% GFP⁺ cells among transduced moDCs (see online supplementary figure S4) in accordance with previous results.³⁸ Importantly, XIAP transduction restored baseline and MDP-induced IL-8 secretion in XIAP E99X moDCs (figure 4F). By contrast, moDCs from healthy controls did not exhibit increased baseline or MDP-induced IL-8 secretion upon lentiviral XIAP transduction, thus demonstrating that XIAP expression is not rate-limiting for NOD2 signalling in moDCs from healthy controls (figure 4F). Together, these results demonstrate that impaired NOD2 signalling is the consequence of the XIAP E99X variant.

Targeted sequencing reveals additional CD-associated variants in XIAP

Another case of early onset CD associated with a hemizygous missense mutation (C203Y) in XIAP has recently been

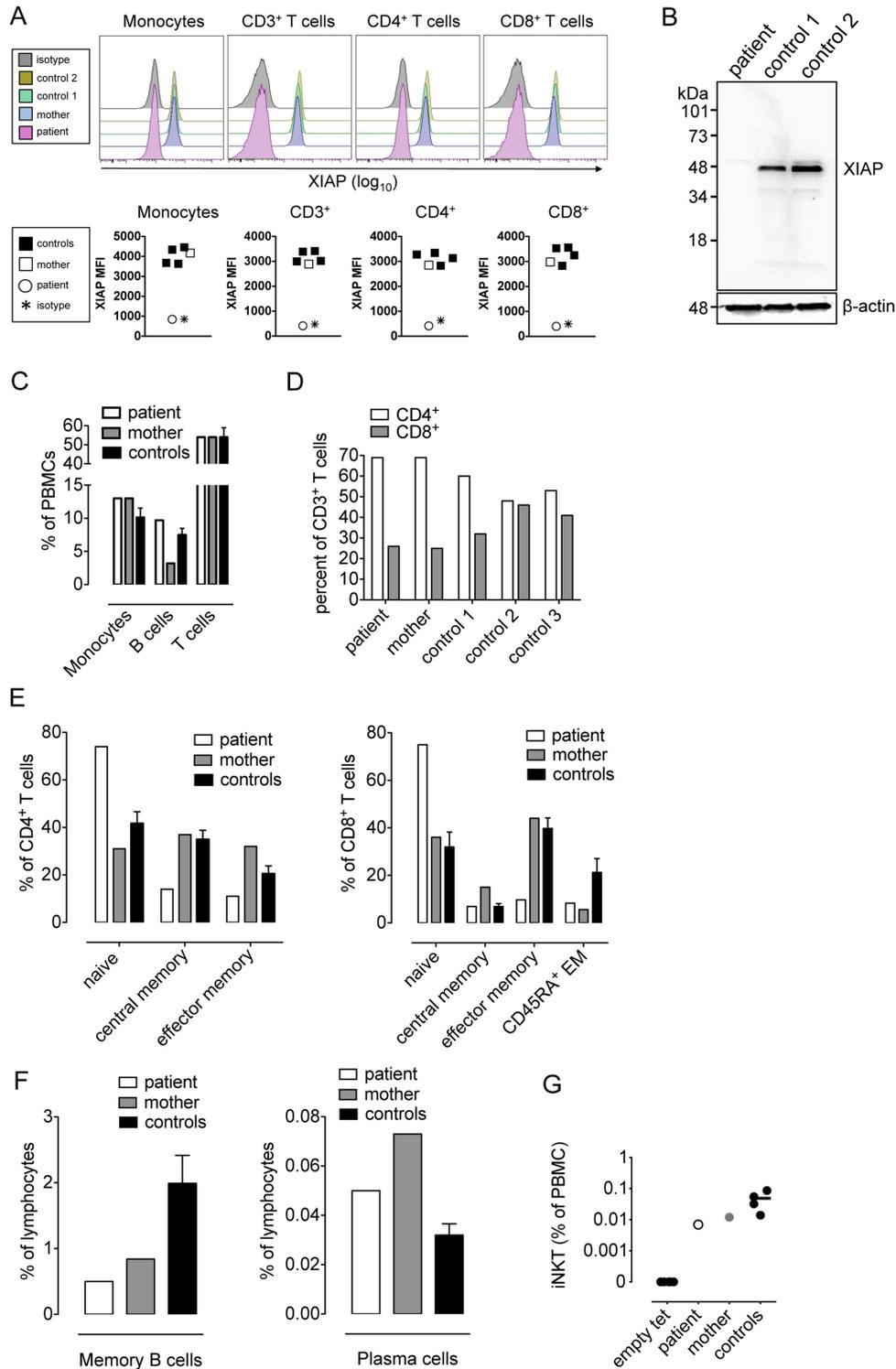
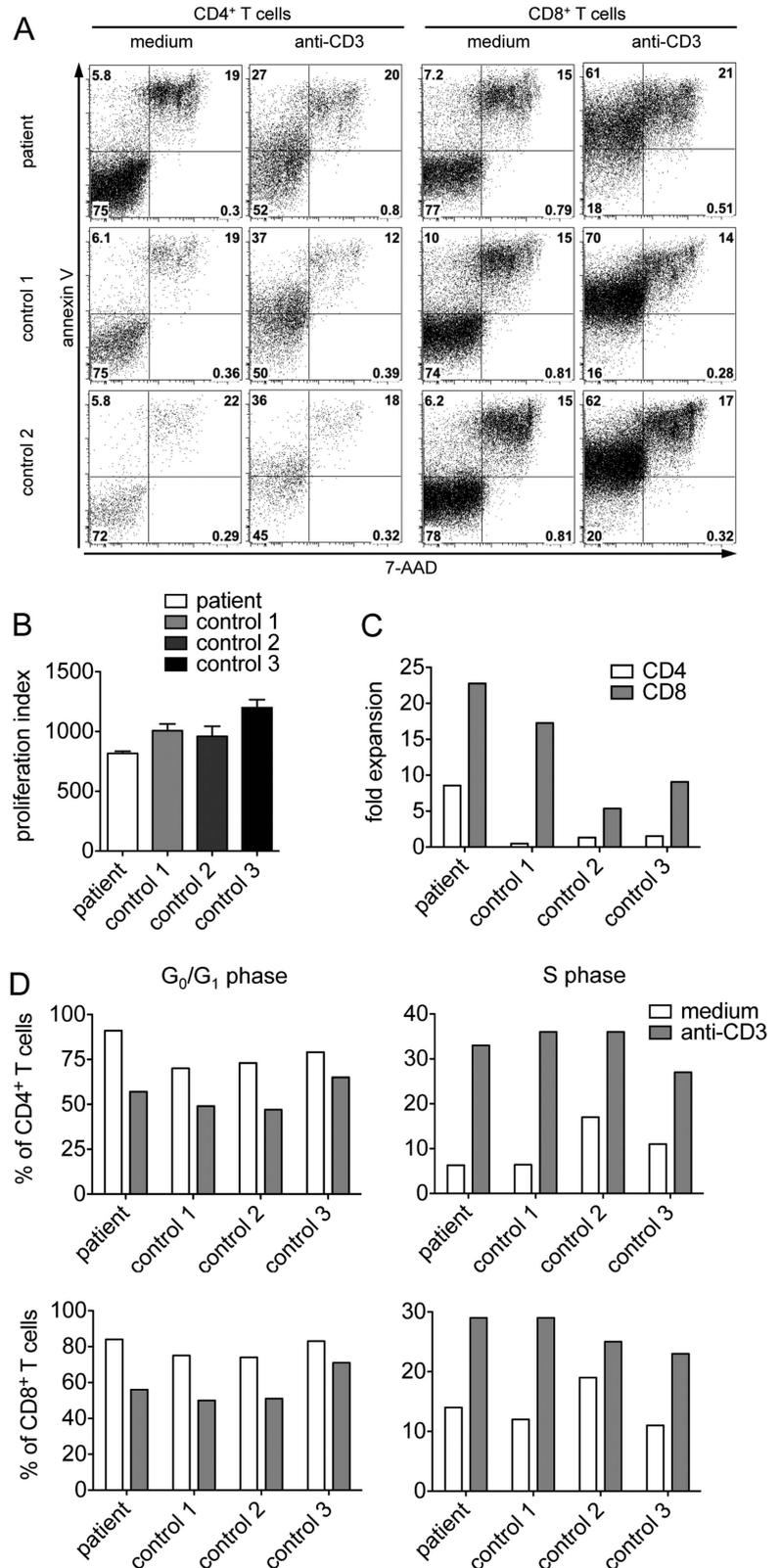


Figure 2 The XIAP E99X variant is not associated with alterations in PBMC frequencies. **A:** XIAP expression of the indicated PBMC subsets as measured by intracellular flow cytometry. Histograms are shown in the upper panel and mean fluorescence intensities (MFI) are given in the lower panel. **B:** XIAP protein expression as determined by western blot analysis in lymphoblastic cell lines. **C–F:** characterisation of PBMC subsets by flow cytometry. **C:** percentage of CD14⁺ monocytes, CD19⁺ B cells, CD3⁺ T cells among PBMCs. **D:** percentage of CD4 and CD8⁺ among CD3⁺ T cells. **E:** naive (CCR7⁺, CD45RA⁺, CD28⁺), central memory (CCR7⁺, CD45RA⁻, CD28⁺), effector memory (CCR7⁻, CD45RA⁻, CD28⁺), and CD45RA⁺ effector memory (CCR7⁻, CD45RA⁺, CD28⁺) subsets of CD4 and CD8⁺ T cells according to ref^{64–66}. **F:** percentage of CD19⁺, CD27⁺, CD38⁻ peripheral memory B cells and CD19⁺, CD27⁺, CD38⁺ plasma cells. **F:** percentage of α -galactosylceramide (PBS57)/CD1d-tetramer-positive CD3⁺ iNKT cells. In **C, E, F**, mean \pm sem of three healthy controls are shown. In all other panels, individual controls are shown. Statistical analysis is not provided, as studies with primary PBMCs from the XIAP E99X patient were performed only once.

identified.³¹ To test for potential recurrence of XIAP variants, we carried out genotyping of the E99X and C203Y mutations in 1900 CD patients, 780 UC patients and 2864 healthy

controls (see Methods). Genotyping results were monomorphic (data not shown), suggesting a private character of these two variants.

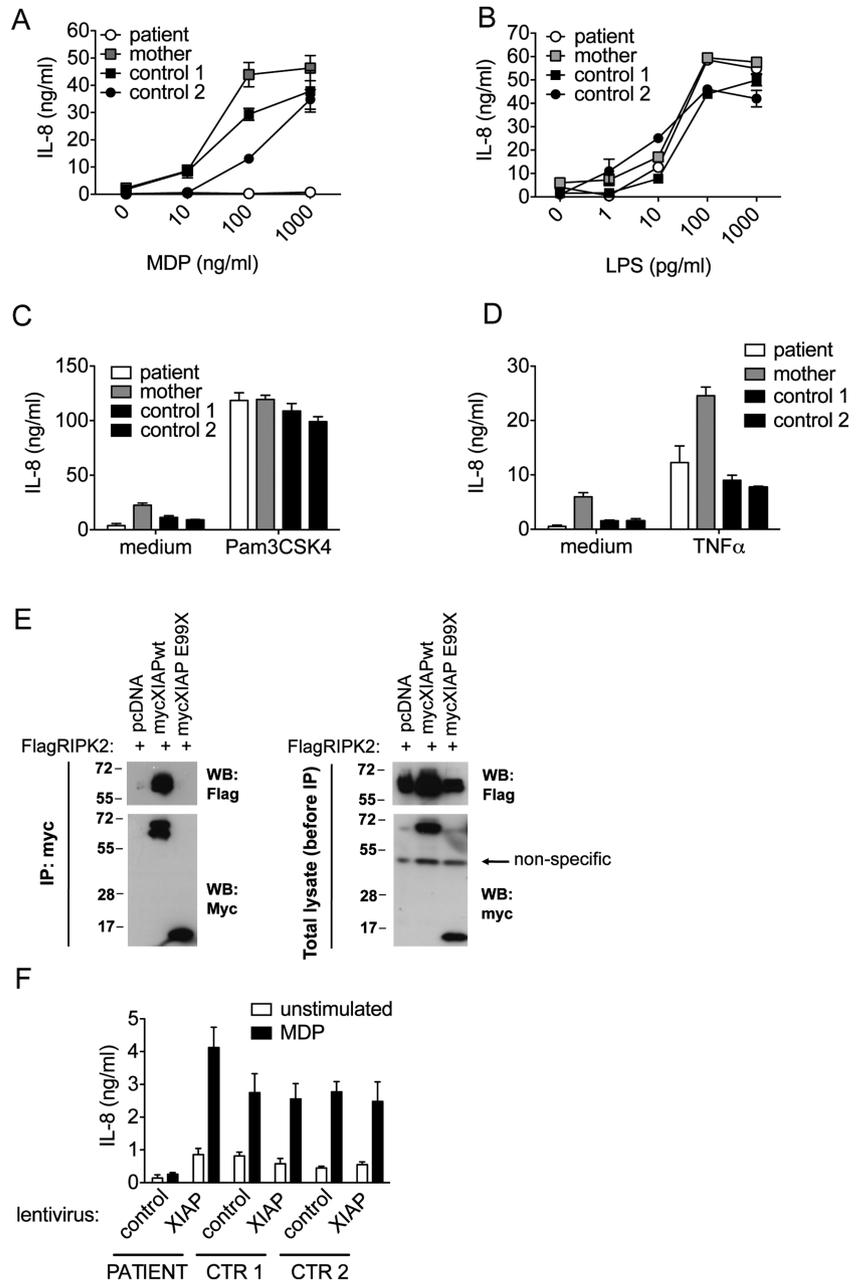
Figure 3 Unaltered apoptosis and proliferation of T cells obtained from the XIAP E99X patient. **A:** PBMCs were stimulated with phytohemagglutinin (PHA) in the presence of IL-2. Reactivation was induced by anti-CD3 cross-linking. Reactivation-induced cell death was determined by Annexin V and 7-AAD staining. **B:** activation-induced T cell proliferation induced by PHA and measured by ³H-thymidine incorporation in total T cells. **C:** fold expansion of the indicated T cell subsets 14 days after PHA treatment. **D:** cell cycle analysis of the indicated T cell subsets two days after anti-CD3 cross-linking as determined by flow cytometric BrdU/7-AAD analysis. CD4⁺ T cells are shown in the upper panel. CD8⁺ T cells are shown in the lower panel. In B, mean ±sem of triplicate cultures is shown. In all other panels, individual controls are shown. Statistical analysis is not provided as studies with primary PBMCs from the XIAP E99X patient were performed only once.



To investigate whether additional XIAP variants can be identified in patients with IBD, targeted Sanger sequencing was performed in a cohort of 275 paediatric-onset IBD cases (age ≤ 16 years according to groups A1a and A1b of the Paris classification,¹²) and 1047 adult-onset CD patients (≥ 17 years according to groups A2 and A3 of the Montreal classification;⁵¹ see Methods for further details). Sanger sequencing revealed four

novel variants in XIAP including one synonymous, two missense and one nonsense variant (table 1). Non-synonymous mutations occurred exclusively in males with paediatric-onset CD (see Suppl. Results), were hemizygous, and located in exons 2 and 3 of the XIAP gene (table 1, figure 1A). Remarkably, missense variants G39C (p.Gly39Cys) and K297T (p.Lys297Thr) were located in the BIR1 and BIR3 domains of XIAP, respectively, and

Figure 4 A severe NOD2 defect in the XIAP E99X patient, impaired interaction of XIAP E99X with RIPK2, and functional NOD2 restoration upon lentiviral XIAP reconstitution. A–D: IL-8 secretion as determined by ELISA in supernatants of PBMCs treated with the indicated stimulants for 24 h (Pam3CSK4 10 µg/mL, TNFα 5 ng/mL). E: the left panel shows coimmunoprecipitation of FLAG-tagged RIPK2 with WT but not mutant myc-tagged XIAP as determined by western blotting in 293 T cells. The right panel shows expression of RIPK2 and XIAP in cell lysates. F: restoration of NOD2 signalling by lentiviral reconstitution with XIAP wildtype protein in monocyte-derived dendritic cells of the XIAP E99X patient. In A–D, F, mean±sem of triplicate cultures are shown. Results in E are representative of two independent experiments. Studies in A–D and F involving primary PBMCs of the XIAP E99X patient were performed only once and statistical analysis is therefore not provided.

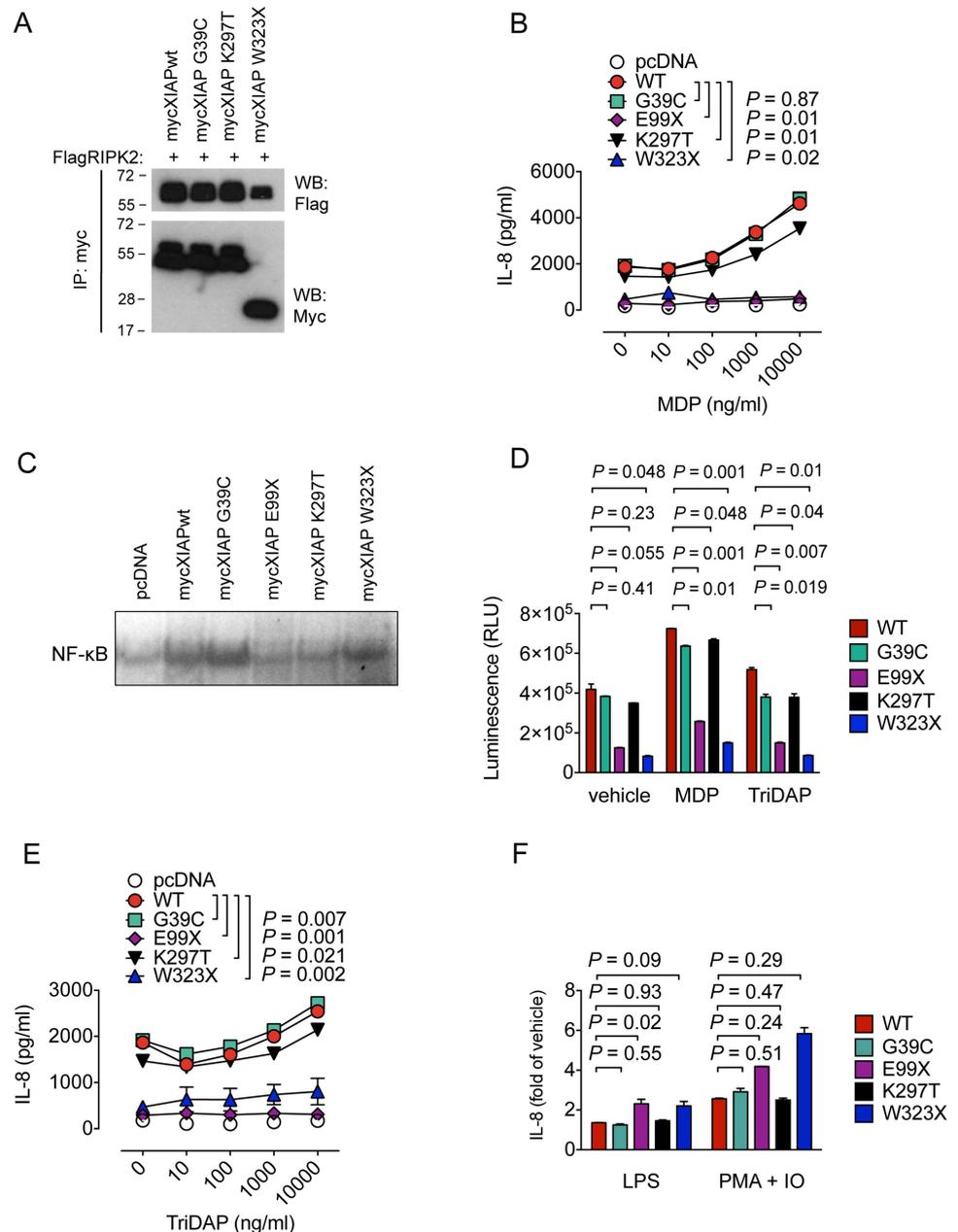


are thus the first disease-associated missense variants described in XIAP, which are not associated with either the BIR2 or the RING domain. Together, a total of four novel, private, non-synonymous XIAP variants (G39C, E99X, K297T, W323X (p.Trp323*)) were identified among 96 male patients with paediatric-onset CD, which highlights their frequent occurrence in this subset of CD patients. Importantly, these four variants are not present in previous datasets from the 1000 Genomes Project and the NHLBI Exome Sequencing Project (ESP).

The patient carrying XIAP W323X showed ileocolonic and perianal disease (A1b, L3, B2B3p¹²). The onset of CD occurred at age 16 years with fulminant colitis requiring colectomy, and a subsequent course of disease characterised by stricturing and fistulising ileitis requiring multiple ileal resections. The patient received continuous corticosteroid treatment between the ages of 16 and 52 years. Azathioprine and infliximab treatment were initiated at 50 and 53 years of age, respectively, and were discontinued due to severe respiratory infections. The patient died

at age 54 years from hospital-acquired pneumonia (see Suppl. Results for further clinical details). The patient carrying XIAP K297T showed gastric, jejunal, ileal and colonic involvement of CD including pyloric stenosis and perianal disease. CD manifested at age 15 years (A1b, L3+L4a/L4b, B2p¹²), and required combined treatment with steroids, azathioprine and infliximab for disease control (see Suppl. Results and see online supplementary figure S5 for further clinical details). Both patients were repeatedly tested negative for EBV and did not exhibit signs of haemophagocytic lymphohistiocytosis (HLH). The patient carrying XIAP G39C is a male CD patient with age of disease onset at 13 years who was enrolled at the age of 38 years in an infliximab study. Further clinical information was not available on this patient. Thus, all patients with clinical data available shared the presence of small and large intestinal involvement, perianal disease and stricturing behaviour, while the location of small intestinal disease and the presence or absence of penetrating behaviour differed among patients.

Figure 5 Impaired NOD1/2-mediated NF- κ B-dependent signalling by XIAP E99X, W323X, and K297T. A: coimmunoprecipitation of FLAG-tagged RIPK2 with WT and mutant myc-tagged XIAP after transfection into 293 T cells. B: IL-8 secretion by XIAP-deficient HCT116 cells transfected with WT or mutant XIAP as well as the empty vector control (pcDNA) and stimulated with the indicated concentration of MDP. C: DNA binding activity of NF- κ B as determined by electrophoretic mobility shift assay in XIAP-deficient HCT116 cells transfected with WT or mutant XIAP and stimulated with MDP (10 μ g/mL). D: NF- κ B-dependent transcription as determined by expression of an NF- κ B-dependent firefly luciferase construct after transfection into XIAP-deficient HCT116 cells along with a renilla luciferase plasmid for normalisation and WT or mutant XIAP. Cells were stimulated as indicated (MDP, Tri-DAP: 10 μ g/mL). E: IL-8 secretion by XIAP-deficient HCT116 cells transfected with WT or mutant XIAP or empty vector control (pcDNA) and stimulated with the indicated concentrations of TriDAP. F: IL-8 secretion by XIAP-deficient HCT116 cells transfected with WT or mutant XIAP and stimulated with LPS (1 μ g/mL) or PMA (50 ng/mL) and ionomycin (1 μ M). Mean \pm sem is shown in B, D, E and F. Results are representative of three independent experiments.



XIAP W323X affects NOD1/NOD2 signalling through truncation of the RING domain

Of the three additional XIAP variants identified, W323X is associated with loss of the XIAP RING domain, but predicted to preserve the BIR2 domain (figure 1A and see online supplementary figure S6). As the patient harbouring XIAP W323X had died before this investigation, we were unable to study the expression and function of this XIAP variant in primary cells. Upon ectopic expression in 293 T cells, XIAP W323X exhibited largely unimpaired expression as well as unaltered coimmunoprecipitation of RIPK2 in line with an intact BIR2 domain (figure 5A). However, despite preserved RIPK2 recruitment, loss of the XIAP RING domain associated with the W323X mutation is expected to affect NOD signalling through impaired RING domain-dependent ubiquitylation of RIPK2.⁴⁷ We therefore investigated whether expression of XIAP W323X can restore NOD2 signalling in XIAP-deficient HCT116 cells.⁵² In accordance with a critical function of the XIAP RING domain in NOD2 signalling, MDP-induced,

NOD2-dependent IL-8 release was only observed with wildtype, but not W323X XIAP (figure 5B). Expression of XIAP W323X was associated with reduced DNA binding activity of NF- κ B and impaired NF- κ B-dependent transcription in response to MDP (figure 5C, D). This suggested that altered IL-8 secretion occurred as a consequence of impaired XIAP- and NOD2-dependent NF- κ B activation. As expected, similar defects in NF- κ B activation and IL-8 secretion were observed with XIAP E99X, which is associated with loss of the BIR2, BIR3 and RING domains of XIAP (figure 5B–D). Of note, reconstitution of XIAP-deficient HCT116 cells with wildtype, but not W323X or E99X XIAP also restored basal IL-8 secretion in the absence of MDP stimulation (figure 5B). These findings resembled observations in primary moDCs harbouring XIAP E99X, where lentiviral transduction of wildtype XIAP restored NOD2-induced secretion and also constitutive IL-8 secretion (figure 4F). As such, these results raised the question of whether XIAP is required for basal and NOD2-induced NF- κ B signalling. In line with this concept, NF- κ B-dependent transcription in the

absence of MDP stimulation was reduced upon expression of W323X and E99X compared to WT XIAP (figure 5D).

Defects observed with XIAP W323X extended to NOD1, as impaired NF- κ B-dependent transcription and IL-8 secretion were also observed in response to the NOD1 agonist Tri-DAP (figure 5D–E). Similar observations were made for XIAP E99X (figure 5D–E). HCT116 cells, even in the presence of wildtype XIAP, did not exhibit IL-8 secretion in response to LPS, Pam3CSK4, and TNF α (figure 5F and data not shown). However, IL-8 production in response to phorbol 12-myristate 13-acetate (PMA) and ionomycin (IO) was unimpaired upon expression of W323X and E99X compared to wildtype XIAP suggesting the absence of general defects in IL-8 production or cell viability (figure 5F). In conclusion, these data demonstrate that XIAP E99X and W323X are associated with impaired NOD1/2- and NF- κ B-dependent cytokine responses as a consequence of loss of the XIAP BIR2 and/or RING domain.

Functional analysis of XIAP variants located in the BIR1 and BIR3 domain of XIAP

XIAP variants previously identified in patients with XLP2 are nonsense and frameshift mutations as well as deletions, which invariably affect the RING domain of XIAP and disrupt NOD1/2 signalling.^{25–30} Additionally, XIAP missense mutations were described, which exclusively affect the BIR2 and RING domains and are associated with impaired RIPK2-mediated NOD1/2 signalling.^{29 31 49} Thus, disease-associated XIAP variants identified so far share the principle that they affect NOD1/2 signalling through direct or indirect interference with the function of the BIR2 or RING domain. By contrast, mutations G39C and K297T described here are the first disease-associated XIAP missense variants, which are located in the BIR1 and BIR3 domains of XIAP, respectively. We therefore first investigated whether these XIAP variants are associated with impaired expression as commonly observed for XIAP missense variants.²⁹ However, XIAP G39C and K297T exhibited expression levels similar to wildtype XIAP upon ectopic expression in 293 T cells (figure 5A). Moreover, in accordance with an intact BIR2 domain, recruitment of RIPK2 by XIAP G39C and K297T was unimpaired (figure 5A). We then investigated whether reconstitution of XIAP-deficient HCT116 cells with G39C and K297T XIAP restores NOD1/2 signalling. Expression of XIAP K297T was associated with reduced MDP-induced DNA binding activity of NF- κ B (figure 5C) as well as a modest reduction in MDP-induced and Tri-DAP-induced NF- κ B-dependent transcription (figure 5D) and IL-8 secretion (figure 5B and E). By contrast, MDP-induced DNA binding activity of NF- κ B (figure 5C) as well as MDP-induced and Tri-DAP-induced IL-8 secretion (figure 5B and E) were unaffected in the presence of XIAP G39C, while a minor reduction in MDP-induced and Tri-DAP-induced NF- κ B-dependent transcription was observed (figure 5D). Thus, XIAP K297T is associated with a modest defect in NOD1/2 signalling, while NOD function is largely preserved in the presence of XIAP G39C.

DISCUSSION

Our studies provide the first systematic analysis of XIAP variants in patients with IBD and reveal their unexpected, frequent occurrence in male patients with paediatric-onset CD. Mutations in XIAP are associated with XLP2, a PID characterised by susceptibility to Epstein Barr virus (EBV) infection and HLH.^{25 45} Chronic intestinal inflammation resembling CD has been observed in XLP2.^{25 27–30} Moreover, relatives of XLP2 patients with shared XIAP variants occasionally developed

intestinal inflammation in the absence of HLH suggesting that XIAP may regulate susceptibility to IBD independent of lymphoproliferative disease.^{27–29} In accordance with this concept, Worthey *et al* described a patient with early onset CD in the absence of lymphoproliferative disease, who carried a missense mutation in XIAP.³¹ Moreover, a recent summary of patients with XIAP mutations highlighted the variability of clinical manifestations and revealed several cases with CD as the sole clinical manifestation.³⁰ Our identification of XIAP E99X in a patient with early onset CD provided the rationale for the first systematic screening for XIAP variants in IBD. These analyses revealed the frequent occurrence of private variants in XIAP, which were restricted to male patients with paediatric-onset CD (age \leq 16 years according to group A1 of the Montreal classification,⁵¹ and A1a/b of the Paris classification¹²). Four novel and private XIAP variants were found among 96 male patients with paediatric-onset CD, together reflecting about 4% of patients in this subset of CD patients. CD patients carrying those variants showed small and large intestinal involvement, perianal disease and stricturing behaviour, while the location of small intestinal disease and the presence of penetrating behaviour differed among patients. Together, these results reveal a common association of XIAP variants with male, paediatric-onset CD independent of the occurrence of lymphoproliferative disease and warrant future systematic sequencing of XIAP in this subset of CD patients. While we did not detect genetic variants in XIAP in a large cohort of adult-onset CD, we observed considerable variation in the age of onset of intestinal inflammation in paediatric patients carrying such variants. Given these findings as well as the previous description of two patients with adult-onset IBD and mutations in XIAP,^{29 30} we suggest that screening for XIAP variants in male patients with severe CD should not be limited to cases of early onset CD, in accordance with previous suggestions.⁵³

The clinical course of disease in CD patients with XIAP mutations is often severe and refractory to treatment with immunomodulators and biologicals, which is reflected in the clinical history of patients reported by us and others.^{30 31} For these patients, allogeneic haematopoietic stem cell transplantation (HSCT) may offer a causal therapeutic approach able to restore XIAP expression and function at least in bone marrow-derived cells. HSCT has thereby demonstrated the potential to provide lasting remission of intestinal inflammation in CD patients carrying XIAP mutations.^{26 30 31} However, considerable morbidity and mortality associated with HSCT in patients with XIAP mutations²⁶ suggests that such treatment should be limited to selected individuals with severe CD refractory to currently available medical treatment.

The association of XIAP variants with IBD and PID is reminiscent of a number of PIDs that manifest as IBD and supports the concept of immunodeficiency underlying a subset of IBD cases.^{8–11 19–24} It is noteworthy in this context that the majority of XIAP variants identified here were associated with defects in NOD2, a microbial pattern recognition receptor and CD susceptibility locus.^{54 55} Immune defects in patients with XIAP mutations may thus extend beyond the known susceptibility to EBV and may contribute to intestinal inflammation. This is supported by the observation that *Xiap*-deficient mice exhibit impaired NOD2 function and susceptibility to bacterial infections in a manner resembling that observed for *Nod2*-deficient mice.^{46 56 57} Recent findings of a critical role of NOD2 in viral recognition^{58 59} and the crosstalk between viruses and bacteria^{60 61} thereby suggest that NOD2-dependent immunodeficiency in patients with XIAP variants may extend to

enteropathogenic viruses with potential implications for the pathogenesis of CD. However, despite a clear association between *NOD2* variants and IBD, particularly in patients homozygous for *NOD2* risk alleles,⁵⁴ it remains to be investigated whether intestinal inflammation in patients harbouring *XIAP* mutations is indeed related to *NOD* dysfunction.

Finally, several observations suggest that the loss of functional *XIAP* is not sufficient to elicit intestinal inflammation but rather requires the presence of other genetic or environmental factors: As such, intestinal inflammation exhibits incomplete penetrance in individuals with shared *XIAP* variants, and neither the location nor type of mutation predicts the clinical manifestations associated with *XIAP* mutations.^{29–30} This contrasts with findings made for homozygous loss-of-function mutations in *IL10*, *IL10RA* and *IL10RB*, which were invariably associated with intestinal inflammation manifesting within the first months of life as well as the absence of homozygosity of these mutations in healthy relatives.^{9–11 13–14} Consistent with these observations in humans, deficiency in *Il10* and *Il10rb* but not in *Xiap* is associated with spontaneous intestinal inflammation in mice.^{15–16} While the association of *XIAP* variants with *NOD1/2* defects and susceptibility to infection are in line with the potential involvement of a microbial trigger in the aetiology of intestinal inflammation, studies by Rigaud *et al* demonstrated that not only environmental but also genetic factors may influence the clinical presentation of genetic variants in *XIAP*.⁶²

In conclusion, our studies reveal an association between *XIAP* variants and male, paediatric-onset CD. The known association between *XIAP* mutations and PID thereby supports the notion that a subset of IBD cases may result from immunodeficiency. Moreover, these findings may provide a mechanistic basis for the unexplained observation of functional *NOD2* defects in the absence of genetic *NOD2* variants.⁶³

Author affiliations

¹Department of Internal Medicine I, University Medical Center Schleswig-Holstein, Kiel, Germany

²Department of General Pediatrics, University Medical Center Schleswig-Holstein, Kiel, Germany

³Institute of Clinical Molecular Biology, Christian-Albrechts-University of Kiel, Kiel, Germany

⁴Sanford-Burnham Medical Research Institute, La Jolla, California, USA

⁵Clinical Bioinformatics, Saarland University, Saarbrücken, Germany

⁶Children's Hospital, Medical Faculty Carl Gustav Carus, Technische Universität Dresden, Germany

⁷Department of General Pediatrics, University Medical Center Schleswig-Holstein, Lübeck, Germany

⁸Institute of Pathology, University Medical Center Schleswig-Holstein, Lübeck, Germany

⁹Institute of Pathology, University Medical Center Schleswig-Holstein, Kiel, Germany

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Authors: Yvonne Zeissig, Britt-Sabina Petersen, Snezana Milutinovic, Esther Bosse, Gabriele

Mayr, Kenneth Peuker, Jelka Hartwig, Andreas Keller, Martina Kohl, Martin W. Laass, Susanne

Billmann-Born, Heide Brandau, Alfred Feller, Christoph Röcken, Martin Schrappe, Philip

Rosenstiel, John C. Reed, Stefan Schreiber, Andre Franke, Sebastian Zeissig

Supplementary Results

Clinical history of patients harboring *XIAP* variants

Patient 1 (XIAP E99X)

A Caucasian male patient presented with bloody diarrhea and anal fissure at the age of nine months. Initially, cow's milk protein intolerance was suspected. However dietary exclusion of milk protein did not resolve symptoms. At the age of 19 months, a colonoscopy was performed, which demonstrated severe, erosive, discontinuous pancolitis with anal fissures and anal stenosis and histological demonstration of superficial, erosive, and ulcerative inflammation with crypt abscesses but without epithelioid granulomas. Histologic and serologic testing for EBV, CMV, and mycobacteria was negative. The family history was negative for immunodeficiency syndromes and inflammatory bowel disease. The subsequent course of disease was refractory to treatment with local and oral mesalazine, corticosteroids, antibiotics, probiotics, and with intolerance to azathioprine (pancreatitis). Anal dilatation was associated with symptomatic relief. Colonoscopy at 31 months of age revealed a similar picture of moderate to severe colitis, anal fissures, and anal stenosis. Upper endoscopy and MR enterography did not reveal signs of macroscopic small intestinal or upper gastrointestinal disease. However, histologic examination demonstrated low-grade, chronic, *Helicobacter pylori*-negative gastritis and duodenitis in accordance with upper gastrointestinal CD. An MRI revealed hepatosplenomegaly. In addition, papular and pustular skin manifestations were found at the back and thighs with histiocytic infiltrates and epithelioid cells in accordance with a cutaneous CD manifestation. Following primary failure of infliximab, the patient received a loop ileostomy at the age of 36 months, which was associated with significant clinical improvement despite persistent mild, discontinuous colitis. At 40 months of age, surgical treatment of an anocutaneous fistula was performed. Since then, the patient has been in clinical remission for more than 17 months without treatment. Self-limiting episodes of fever have occurred, but never fulfilled the criteria of hemophagocytic lymphohistiocytosis (HLH). The patient continues to be negative for EBV. A summary of laboratory findings is shown in Table S2 and S3. The patient was

classified as A1a, L2 + L4a, B2p according to the Paris modification of the Montreal classification [1].

Patient 2 (XIAP G39C)

Patient 2 is a male Caucasian with CD and an indicated age of onset at 13 years, who was enrolled in an infliximab study at the age of 38 years. Further clinical information on this patient was not available.

Patient 3 (XIAP K297T)

In a Caucasian male patient, the diagnosis of CD was made at the age of 16 years after a one year history of abdominal pain, diarrhea, and weight loss of 10 kg (onset of disease at the age of 15 years). Endoscopy of the upper and lower gastrointestinal tract showed discontinuous, ulcerative inflammation of the jejunum, terminal ileum, left colon, sigmoid colon, and rectum as well as of the gastric antrum with mild pyloric stenosis (Fig. S5). Perianal disease was mild with skin tags but no fistulae. Histology showed multiple granulomas in all inflamed parts of the gastrointestinal tract including the gastric antrum. Disease classification according to the Paris modification of the Montreal classification for CD [1] was A1b, L3+L4a/L4b, B2p. The patient was tested positive for ASCA IgA and negative for ANCA antibodies. The personal and family history were negative for immunodeficiencies and inflammatory bowel disease.

Under an exclusive enteral nutrition for 6 weeks, clinical remission was achieved, although laboratory values (elevated C-reactive protein and fecal calprotectin) still showed disease activity. The patient intermittently required corticosteroid treatment. Azathioprine treatment was initiated at the age of 16 years. At the age of 17 (18 month after the diagnosis of CD was made), infliximab was initiated in addition to azathioprine. At the age of 19, infliximab intervals needed to be reduced to 4 weeks. The patient continued to require intermittent corticosteroid treatment (budesonide) and

infusions of iron (ferric carboxymaltose). Currently, the patient is in stable condition and receives infliximab (4-weekly interval), azathioprine, mesalamine, pantoprazole, zinc, vitamin d, folic acid and supplemental enteral nutrition.

The patient was tested negative for three CD-associated variants of NOD2: R702W (SNP8), G908R (SNP12), 3020insC (SNP13).

Patient 4 (W323X)

Patient 4, a male Caucasian, was diagnosed with CD at the age of 16 years in 1966 (A1b, L3, B2B3p according to the Paris modification of the Montreal classification [1]). The family history was negative for IBD. Due to fulminant colitis, the patient underwent colectomy with ileostomy at the age of 16 years. A stenosing and fistulizing course of ileitis required several consecutive ileal resections in the first year following diagnosis and again at the age of 29 years (1979). From the time of diagnosis until the age of 52, the patient received corticosteroids on a continuous basis. Azathioprine was initiated at the age of 50 but was discontinued after 8 months of treatment due to severe leukopenia associated with recurrent and persistent respiratory infections. The patient reported at our center at the age of 53 (2003), when infliximab treatment was initiated due to severe ileitis with enterocutaneous fistulas. Infliximab was discontinued after 4 months of treatment due to severe pneumonia requiring mechanical ventilation but with negative microbiological testing including negative results for *M. tuberculosis*. After recovery, the patient underwent resection of a stenotic jejunal segment with an enterocutaneous fistula. Following surgery, the patient acquired a nosocomial pneumonia and died at the age of 54. The patient was tested negative for EBV at several occasions.

Supplementary Methods

Cell lines

XIAP-deficient and -sufficient HCT116 cells were a gift from B. Vogelstein (Johns Hopkins University) [2]. HCT116 and 293T cells were cultured in RPMI-1640 containing 10% FCS.

Flow cytometry

Flow cytometry was performed as described previously [3]. Briefly, cells were staining with monoclonal antibodies for 20 min at 4°C. For detection of iNKT cells, staining with PBS57-loaded CD1d tetramers (1h, 4°C, NIH Tetramer Core Facility) was performed. Data were collected using a BD Biosciences FACSVerse and were analyzed by Flowjo (Tree Star, Inc., Ashland, OR). For intracellular staining, cells were first surface-stained, then permeabilized with Cytofix/Cytoperm (BD Biosciences) and washed with Perm/Wash buffer (BD Biosciences) according to the manufacturer's instructions. Antibodies were added for 30 min at 4°C before washing with Perm/Wash buffer. Antibodies and reagents used for flow cytometry were obtained from BD Biosciences except for CD1d-PBS57 tetramers (NIH Tetramer Core Facility). Via-Probe (BD Biosciences), and in case of intracellular staining, fixable viability dye eFluor 450 (eBioscience, Frankfurt, Germany), was added to all stainings for exclusion of dead cells. XIAP clone 48 (BD Biosciences) was used for detection of intracellular XIAP according to [4].

Western blotting and immunoprecipitation

Immunoprecipitation was performed as described previously with minor modifications [5]. Briefly, 293T cells were transfected using Lipofectamine 2000 (Invitrogen) with Flag-RIPK2 together with either empty vector (pcDNA), mycXIAPwt or mutant mycXIAP. 24h after transfection, the cells were lysed and protein complexes were immunoprecipitated using anti-myc antibody, followed by SDS-PAGE and immunoblotting using anti-myc and anti-Flag antibodies. Western blotting was performed as described previously [5]. The following antibodies were used: anti-myc (Roche),

anti-flag (M2, Sigma-Aldrich), rabbit anti-caspase-1 (D7F10; Cell Signaling Technology), mouse anti-XIAP (clone 48, BD Biosciences), mouse anti- β -actin (Sigma-Aldrich).

Monocyte-derived DCs and lentiviral reconstitution

Monocytes were extracted from PBMCs by positive selection using CD14 magnetic beads (Miltenyi Biotec) and were cultured at 1×10^6 cells/ml for 5 days in complete RPMI-1640 medium supplemented with 500 U/ml recombinant hIL-4 and 1000 U/ml recombinant hGM-CSF (PeproTech). Lentiviral production and transduction was performed as described previously [3]. Briefly, lentiviral particles were generated in 293T cells using pCMV Δ R8.9 and pMD2.G [6]. Lentiviral particles were concentrated by ultracentrifugation and titered by GFP expression in 293T cells. On day 3 after isolation of monocytes, monocyte-derived DCs were infected with lentiviruses expressing XIAP and GFP [5] or a control virus expressing GFP only at an MOI of 20 in RPMI-1640 containing 10 μ g/ml protamine sulphate (Sigma Aldrich). Immature transduced DCs were cultured at 1×10^6 cells/ml until day 5 when they were investigated in functional studies.

Supplementary Figures 1-6

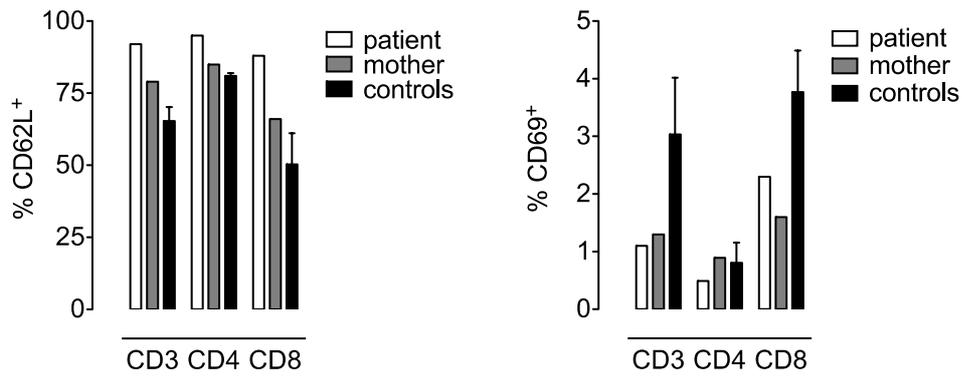


Figure S1. Naive T cell profile in the XIAP E99X patient (Patient 1)

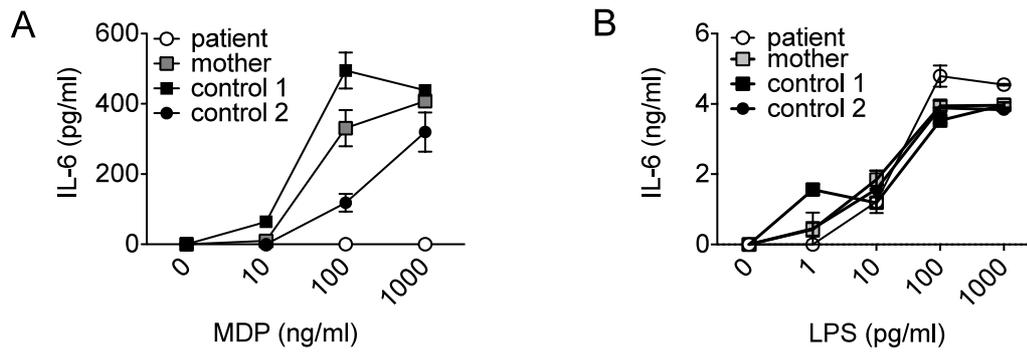


Figure S2. A severe and selective defect in NOD2-dependent IL-6 secretion in the XIAP E99X patient (Patient 1)

IL-6 secretion as determined by ELISA in supernatants of PBMCs treated with the MDP (A) or LPS (B) for 24h. Mean \pm s.e.m. of triplicate cultures are shown.

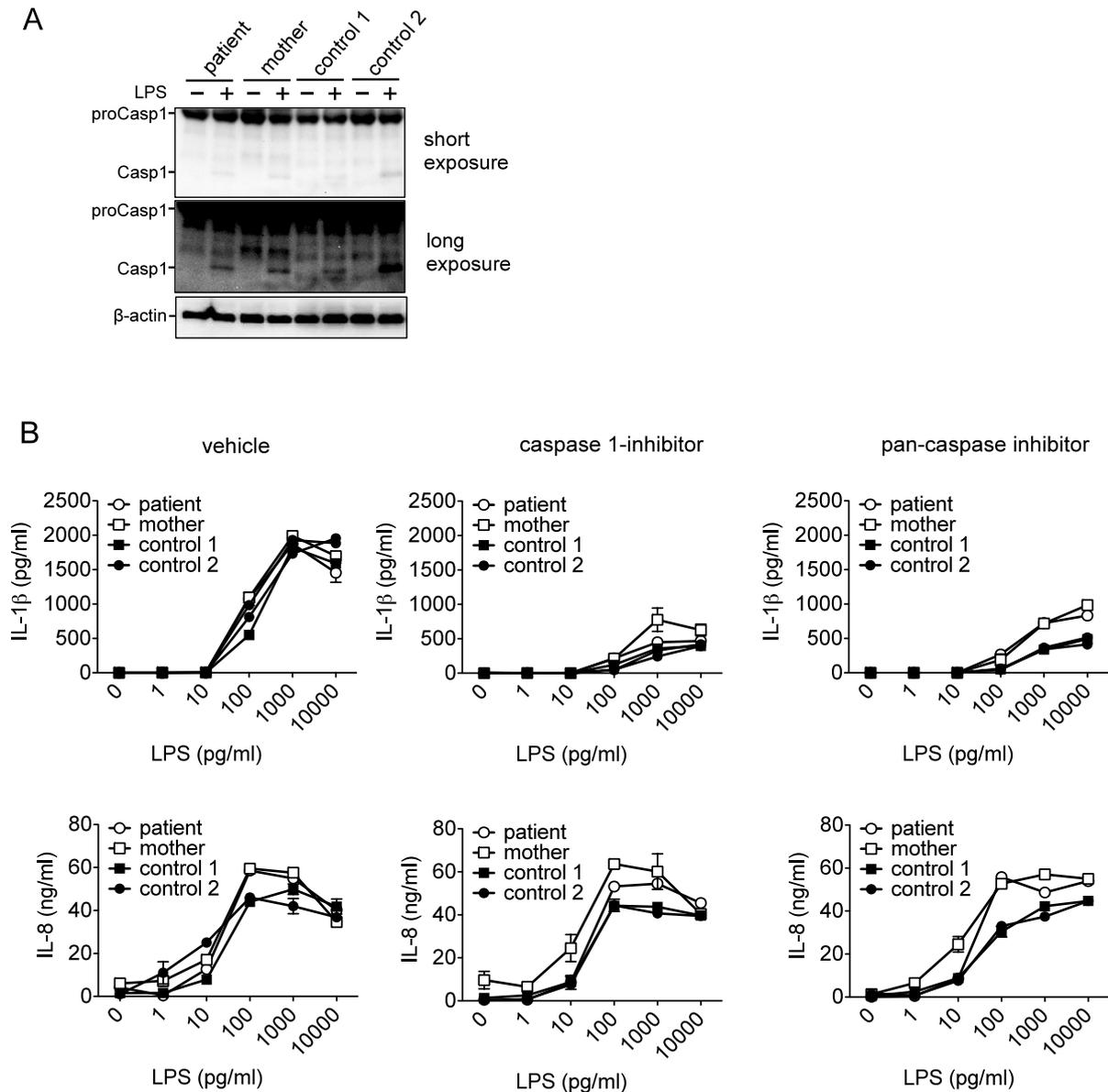


Figure S3. Unaltered caspase-1 cleavage and IL-1 β secretion in the XIAP E99X patient (Patient 1)

A: Caspase-1 cleavage in primary PBMCs induced by treatment with 10 ng/ml LPS for 8 hours as detected by western blotting. B: IL-1 β (upper panel) and IL-8 (lower panel) secretion by PBMCs upon stimulation with LPS at the indicated concentration as determined by ELISA. Where indicated, caspase-1 inhibitor (middle panels, Z-YVAD-fmk, 10 μ M) or pan-caspase inhibitor (right panels; Z-VAD-fmk, 10 μ M) was added to confirm that IL-1 β secretion is mediated by caspase-1. In B, mean \pm s.e.m. of triplicate cultures is shown.

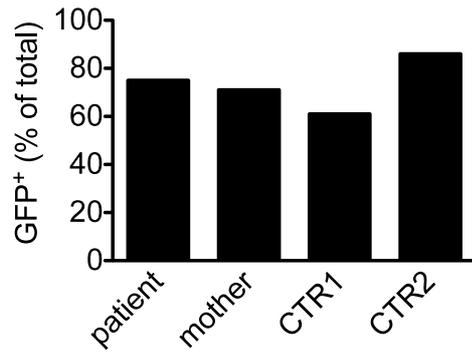


Figure S4. Lentiviral transduction rates of monocyte-derived dendritic cells (moDCs)

The percentage of GFP-positive moDCs 48h after transduction with lentiviruses expressing XIAP is indicated.



Figure S5. Images of upper and lower gastrointestinal endoscopy of patient 3 (XIAP K297T).

A. Gastric antrum with pyloric stenosis. B. Terminal ileitis. C. Colitis of the sigmoid colon.

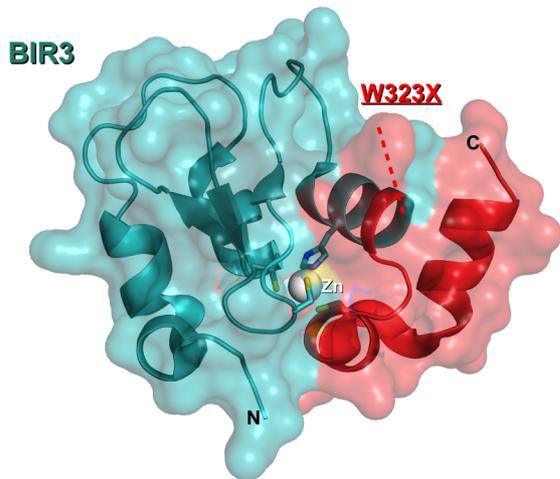


Figure S6. Structural model illustrating the W323X variant (Patient 4). Cartoon and surface representation of the XIAP BIR3 protein domain (PDB ID 2pop, chain B, residues 265-330). Truncation by W323X is indicated by a dashed line and deleted substructures are colored in red. W323X removes the C-terminus of BIR3 including one zinc-coordinating cysteine crucial for domain stability. The mutation likely impairs BIR3 function by preventing proper folding of the domain.

Supplementary Tables 1-5

Table S1. Primer and PCR conditions for Sanger sequencing of *XIAP*.

Primers			
Target	Primer name	F-primer	R-primer
Exon 2, part 1	E2a	TTGCTTGTGTTTCACATTTTG	GTCTGGCCAGTTCTGAAAGG
Exon 2, part2	E2b	TTGAAAATAGTGCCACGCAG	CCATTTTGCTATTGGGGATG
Exon 3	E3	TCTGGGGAACAGAGCCAG	TTTCAAGCCATAGTGAAGGC
Exon 4	E4	TGGGATAGGGAATTGGGTAAC	ACACTGCCCAGCTAGCTCTC
Exon 5	E5	TCCTTCTAGCTCCATCAGGC	CGCAGGCCTGTAGTCCC
Exon 6	E6	GCTTTGCTTTGCTGGTTTG	TGTCCATATCCACCTGTCCC
Exon 7	E7	ACTTGTTGGGTGCTTGGC	CTTGGTAGCAAATGCTAATGG

PCR temperature profile		
95°C	12 min	35 cycles
95°C	30 sec	
60-65°C	30 sec	
72°C	30 sec	
72°C	10 min	
10°C	∞	

Annealing temperatures	
60°C	E2a, E2b
63°C	E3, E6, E7
65°C	E4, E5

Table S2. Summary of microbiological testing of patient 1 (XIAP E99X).

Patient age (months)	20	20	24	24	33	40
Material	blood	stool	rectum biopsy	stool	blood	stool
CMV PCR			neg		neg	
Adenovirus PCR				neg		neg
Norovirus PCR		neg		neg		neg
Rotavirus PCR				neg		neg
Enteropathogenic bacteria culture		neg		neg		neg
EBV-IgG	neg				neg	
EBV-IgM	neg				neg	
EBV-PCR	neg					
Atypical mycobacteria - PCR			neg			

neg = negative; gray fields indicate data not available

Table S3. Summary of clinical laboratory data from patient 1 (XIAP E99X).

Pat. age (months)	19	20	22	27	31	34	38	40	53	Reference
Hb (g/dl)	9.1	10.2		13.1		11.1	15.4		10.2	10.7-13.9
Hkt (%)	27.6	31.5		38.4			46.5		32.9	32.5-41.5
MCV (fl)	76	74		74			67		65	74-89
Leukocytes (/nl)	16.3	7.1		12.7		12.0	10.7	7.5	11.7	5.1-12.9
Lymphocytes (/µl)	11224	7106								3000-9500
CD3 ⁺ cells (/µl)				3103						1800-3000
CD4 ⁺ cells (/µl)				2288						700-1800
CD8 ⁺ cells (/µl)				720						500-1500
CD4/CD8 ratio				3.18						0.865-1.885
Natural Killer cells (/µl)				247						200-600
CD19 ⁺ cells (/µl)				804						300-1300
Monocytes (/µl)				797						50-1000
Eosinophils (/µl)		258								40-650
Basophils (/µl)		129		159						0-70
Thrombocytes (/nl)	287	135		330		428			277	286-509
CRP (mg/l)		3.9				11.5		2.1	19.2	< 5
Antibodies & functional analysis										
IgA (g/l)	0.533			0.857						0.3-1.2
IgG (g/l)	6.61			10.2						3.5-10
IgM (g/l)				1.86						0.4-1.4
IgG1 (g/l)	4.36			7.3						2.4-7.8
IgG2 (g/l)	0.417			0.627						0.55-2
IgG3 (g/l)	0.087			0.225						0.15-0.93
IgG4 (g/l)	0.015			0.046						0.006-0.69
IgE (kU/l)	41.3									< 100
anti-Transglutaminase IgA (kU/l)	2.1									0-9
anti-Transglutaminase IgG (kU/l)	1.8									0-9
ANA			neg							
pANCA			neg							
cANCA			neg							
anti-myeloperoxidase			neg							
anti-proteinase			neg							
anti-lactoferrin			neg							
anti-elastase			neg							
anti-cathepsin G			neg							
anti-lysozyme			neg							
anti-mitochondrial			neg							
ASCA IgG			pos							
ASCA IgM			pos							
SAP staining					n					
Oxidative burst					n					
CD107 degranulation					n					
Stool										
Calprotectin (mg/kg)						592				
Elastase (µg/g stool)								280		> 200

n = normal, neg = negative, pos = positive, pat. = patient; gray fields indicate data not available

Table S4. Exome sequencing coverage (patient 1)

Sample	Coverage of exome target				
	% covered $\geq 1x$	% covered $\geq 8x$	% covered $\geq 20x$	mean coverage	Gb of coverage
father	96.98	92.87	86.88	69.45	4.31
mother	97.17	93.67	88.78	80.54	5.00
child	96.38	91.02	80.72	44.10	2.74

Table S5. Information on variants in *FAM151A*, *RAPGEF4*, and *DCHS2* (patient 1, XIAP E99X)

Gene	Type	Exonic Function	Status in son	Status in parents	Amino Acid Change	SIFT	Poly-Phen2	PhyloP	Mutation Taster
FAM151A	exonic	frameshift deletion	homozygous	both parents het	NM_176782:c.1003_1031del;p.335_344del				
RAPGEF4	exonic	nonsynonymous SNV	heterozygous	only father het	NM_007023:c.G215A;p.R72H	0,14	1	1,000	1
RAPGEF4	exonic	nonsynonymous SNV	heterozygous	only mother het	NM_001100397:c.G1504T;p.V502F	0,27	0,05	1,000	0,9953
DCHS2	exonic	nonsynonymous SNV	heterozygous	only father het	NM_017639:c.G7513A;p.A2505T	0	0,980	0,999	0,4202
DCHS2	exonic	stopgain SNV	heterozygous	only mother het	NM_017639:c.T6155G;p.L2052X	0,75	0,565	0,844	1
DCHS2	exonic	nonsynonymous SNV	heterozygous	only mother het	NM_017639:c.G4561T;p.A1521S	0,92	0	0,149	2,70E-05
Gene	Chr	Start	End	Obs	Ref				
FAM151A	chr1	55076138	55076166	-	CACTCCACATTCAGACCGTCATCCCCAGG				
RAPGEF4	chr2	173662259	173662259	A	G				
RAPGEF4	chr2	173882160	173882160	T	G				
DCHS2	chr4	155156926	155156926	T	C				
DCHS2	chr4	155158284	155158284	C	A				
DCHS2	chr4	155219540	155219540	A	C				

Supplementary References

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